DOI: 10.2903/j.efsa.2025.9305

SCIENTIFIC OPINION



Commodity risk assessment of *Populus alba*, *Populus nigra* and *Populus tremula* plants from the UK

EFSA Panel on Plant Health (PLH) | Antonio Vicent Civera | Paula Baptista | Anna Berlin | Elisavet Chatzivassiliou | Jaime Cubero | Nik Cunniffe | Eduardo de la Peña | Nicolas Desneux | Francesco Di Serio | Anna Filipiak | Beata Hasiów-Jaroszewska | Hervé Jactel | Blanca B. Landa | Lara Maistrello | David Makowski | Panagiotis Milonas | Nikos Papadopoulos | Roel Potting | Hanna Susi | Dirk Jan van Der Gaag | Andrea Battisti | Claude Bragard | Christer Sven Magnusson | Hugo Mas | Daniel Rigling | Massimo Faccoli | Alžběta Mikulová | Fabio Stergulc | Eugen Christoph | Olaf Mosbach-Schulz | Franz Streissl | Paolo Gonthier

Correspondence: plants@efsa.europa.eu

The declarations of interest of all scientific experts active in EFSA's work are available at https://open.efsa.europa.eu/experts

Abstract

The European Commission requested the EFSA Panel on Plant Health to prepare and deliver risk assessments for commodities listed in Commission Implementing Regulation (EU) 2018/2019 as 'High risk plants, plant products and other objects'. This Scientific Opinion covers plant health risks posed by plants of *Populus alba*, Populus nigra and Populus tremula imported from the United Kingdom (UK) as: (a) 1to 7-year-old bare root plants, (b) 3- to 15-year-old plants in pots, (c) 1- to 2-year-old cell grown plants and (d) bundles of 1- to 2-year-old cuttings/graftwood (only for P. nigra and P. tremula), taking into account the available scientific information, including the technical information provided by the UK. All pests associated with the commodity were evaluated against specific criteria for their relevance for this Opinion. Two EU protected zone quarantine pests, i.e. Bemisia tabaci (European populations) and Entoleuca mammata, fulfilled all relevant criteria and were selected for further evaluation. For the selected pests, the risk mitigation measures implemented in the technical dossier from the UK were evaluated taking into account the possible limiting factors. Expert judgements were given on the likelihood of pest freedom taking into consideration the risk mitigation measures acting on the pest, including uncertainties associated with the assessment. The age of the plants was considered, reasoning that older trees are more likely to be infested mainly due to longer exposure time and larger size. The degree of pest freedom varies between the pests evaluated, with E. mammata being the pest most frequently expected on the imported plants. The Expert Knowledge Elicitation (EKE) indicated with 95% certainty that between 9730 and 10,000 per 10,000 P. tremula rooted plants in pots (3 to 15 year old) will be free from *E. mammata*.

KEYWORDS

commodity risk assessment, European Union, plant health, plant pest, poplar

This is an open access article under the terms of the Creative Commons Attribution-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited and no modifications or adaptations are made. © 2025 European Food Safety Authority. *EFSA Journal* published by Wiley-VCH GmbH on behalf of European Food Safety Authority.

CONTENTS

Ab	stract		1				
1.	Intro	duction	4				
	1.1.	Background and Terms of Reference as provided by European Commission					
		1.1.1. Background	4				
		1.1.2. Terms of Reference	4				
	1.2.	Interpretation of the Terms of Reference	4				
2.	Data	and Methodologies	5				
	2.1.	Data provided by DEFRA of the UK	5				
	2.2.	Literature searches performed by EFSA	7				
	2.3.	Methodology	8				
		2.3.1. Commodity data	8				
		2.3.2. Identification of pests potentially associated with the commodity	8				
		2.3.3. Listing and evaluation of risk mitigation measures	8				
		2.3.4. Expert Knowledge Elicitation	g				
3.	Com	modity Data	10				
	3.1.	Description of the commodity	10				
	3.2.	Description of the production areas	11				
	3.3.	Production and handling processes	11				
		3.3.1. Source of planting material	11				
		3.3.2. Production cycle	12				
		3.3.3. Pest monitoring during production	12				
		3.3.4. Pest management during production	13				
		3.3.5. Inspections before export	13				
		3.3.6. Export procedure	14				
4.	Iden	tification of pests potentially associated with the commodity	14				
	4.1.	Selection of relevant EU-quarantine pests associated with the commodity	14				
	4.2.	Selection of other relevant pests (non-regulated in the EU) associated with the commodity	18				
	4.3.	Overview of interceptions	18				
	4.4.	List of potential pests not further assessed	19				
	4.5.	Summary of pests selected for further evaluation	19				
5.	Risk	mitigation measures	19				
	5.1.	Risk mitigation measures applied in the UK	19				
	5.2.	Evaluation of the current measures for the selected relevant pests including uncertainties	21				
		5.2.1. Overview of the evaluation of <i>Bemisia tabaci</i> (European populations) (Hemiptera; Aleyrodidae)	21				
		5.2.2. Overview of the evaluation of <i>Entoleuca mammata</i> (Xylariales; Xylariaceae)	23				
		5.2.3. Outcome of Expert Knowledge Elicitation	26				
6.	Con	-lusions	30				
Glo	ossary		30				
Ab	brevia	tions	31				
Ac	knowl	edgements	31				
Re	questo)r	31				
Qu	estior	numbers	31				
Со	pyrigh	t for non-EFSA content	31				
Pai	nel me	mbers	31				
Ret	ferenc	es	31				
Ap	pendi	x A	34				
' An	pendi	x B	95				
-12							

T

Appendix C	
Appendix D	
Appendix E	
Appendix F	

1 | INTRODUCTION

1.1 | Background and Terms of Reference as provided by European Commission

1.1.1 | Background

The Plant Health Regulation (EU) 2016/2031,¹ on the protective measures against pests of plants, has been applied from December 2019. Provisions within the above Regulation are in place for the listing of 'high risk plants, plant products and other objects' (Article 42) on the basis of a preliminary assessment, and to be followed by a commodity risk assessment. A list of 'high risk plants, plant products and other objects' has been published in Regulation (EU) 2018/2019.² Scientific opinions are therefore needed to support the European Commission and the Member States in the work connected to Article 42 of Regulation (EU) 2016/2031, as stipulated in the terms of reference.

1.1.2 | Terms of Reference

In view of the above and in accordance with Article 29 of Regulation (EC) No 178/2002,³ the Commission asks EFSA to provide scientific opinions in the field of plant health.

In particular, EFSA is expected to prepare and deliver risk assessments for commodities listed in the relevant Implementing Act as 'High risk plants, plant products and other objects'. Article 42, paragraphs 4 and 5, establishes that a risk assessment is needed as a follow-up to evaluate whether the commodities will remain prohibited, removed from the list and additional measures will be applied or removed from the list without any additional measures. This task is expected to be on-going, with a regular flow of dossiers being sent by the applicant required for the risk assessment.

Therefore, to facilitate the correct handling of the dossiers and the acquisition of the required data for the commodity risk assessment, a format for the submission of the required data for each dossier is needed.

Furthermore, a standard methodology for the performance of 'commodity risk assessment' based on the work already done by Member States and other international organizations needs to be set.

In view of the above and in accordance with Article 29 of Regulation (EC) No 178/2002, the Commission asks EFSA to provide scientific opinion in the field of plant health for *Populus alba*, *Populus nigra* and *Populus tremula* from the UK taking into account the available scientific information, including the technical dossier provided by the UK.

1.2 Interpretation of the Terms of Reference

The EFSA Panel on Plant Health (hereafter referred to as 'the Panel') was requested to conduct a commodity risk assessment of *Populus alba, Populus nigra* and *Populus tremula* from the UK following the Guidance on commodity risk assessment for the evaluation of high risk plant dossiers (EFSA PLH Panel, 2019) and the protocol for commodity risk assessments as presented in the EFSA standard protocols for scientific assessments (EFSA PLH Panel, 2024; Gardi et al., 2024), taking into account the available scientific information, including the technical information provided by the UK.

The EU quarantine pests that are regulated as a group in the Commission Implementing Regulation (EU) 2019/2072⁴ were considered and evaluated separately at species level.

Annex II of Implementing Regulation (EU) 2019/2072 lists certain pests as non-European populations or isolates or species. These pests are regulated quarantine pests. Consequently, the respective European populations, or isolates, or species are non-regulated pests.

Annex VII of the same Regulation, in certain cases (e.g. point 32) makes reference to the following countries that are excluded from the obligation to comply with specific import requirements for those non-European populations, or isolates, or species: Albania, Andorra, Armenia, Azerbaijan, Belarus, Bosnia and Herzegovina, Canary Islands, Faeroe Islands, Georgia, Iceland, Liechtenstein, Moldova, Monaco, Montenegro, North Macedonia, Norway, Russia (only the following parts: Central Federal District (Tsentralny federalny okrug), Northwestern Federal District (Severo-Zapadny federalny okrug), Southern Federal District (Yuzhny federalny okrug), North Caucasian Federal District (Severo-Kavkazsky federalny okrug)

¹Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants, amending Regulations (EU) 228/2013, (EU) 652/2014 and (EU) 1143/2014 of the European Parliament and of the Council and repealing Council Directives 69/464/EEC, 74/647/EEC, 93/85/EEC, 98/57/EC, 2000/29/EC, 2006/91/EC and 2007/33/EC. OJ L 317, 23.11.2016, pp. 4–104.

²Commission Implementing Regulation (EU) 2018/2019 of 18 December 2018 establishing a provisional list of high risk plants, plant products or other objects, within the meaning of Article 42 of Regulation (EU) 2016/2031 and a list of plants for which phytosanitary certificates are not required for introduction into the Union, within the meaning of Article 73 of that Regulation C/2018/8877. OJ L 323, 19.12.2018, pp. 10–15.

³Regulation (EC) No 178/2002 of the European Parliament and of the Council of 28 January 2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. OJ L 31, 1.2.2002, pp. 1–24.

⁴Commission Implementing Regulation (EU) 2019/2072 of 28 November 2019 establishing uniform conditions for the implementation of Regulation (EU) 2016/2031 of the European Parliament and the Council, as regards protective measures against pests of plants, and repealing Commission Regulation (EC) No 690/2008 and amending Commission Implementing Regulation (EU) 2018/2019. OJ L 319, 10.12.2019, p. 1–279.

and Volga Federal District (Privolzhsky federalny okrug), San Marino, Serbia, Switzerland, Türkiye, Ukraine and the United Kingdom (except Northern Ireland⁵).

Consequently, for those countries,

- (i) any pests identified, which are listed as non- European species in Annex II of Implementing Regulation (EU) 2019/2072 should be investigated as any other non-regulated pest.
- (ii) any pest found in a European country that belongs to the same denomination as the pests listed as non-European populations or isolates in Annex II of Implementing Regulation (EU) 2019/2072, should be considered as European populations or isolates and should not be considered in the assessment of those countries.

Pests listed as 'Regulated Non-Quarantine Pest' (RNQP) in Annex IV of the Commission Implementing Regulation (EU) 2019/2072, and deregulated pests (i.e. pest which were listed as quarantine pests in the Council Directive 2000/29/EC and were deregulated by Commission Implementing Regulation (EU) 2019/2072) were not considered for further evaluation. In case a pest is at the same time regulated as a RNQP and as a Protected Zone Quarantine pest, in this Opinion it should be evaluated as Quarantine pest.

In its evaluation the Panel:

- Checked whether the information in the technical dossier (hereafter referred to as 'the Dossier') provided by the applicant (United Kingdom, Department for Environment Food and Rural Affairs – hereafter referred to as 'DEFRA') was sufficient to conduct a commodity risk assessment. When necessary, additional information was requested to the applicant.
- Selected the relevant Union quarantine pests and protected zone quarantine pests (as specified in Commission Implementing Regulation (EU) 2019/2072, hereafter referred to as 'EU quarantine pests') and other relevant pests present in the UK and associated with the commodity.
- Did not assess the effectiveness of measures for Union quarantine pests for which specific measures are in place for the import of the commodity from the UK in Commission Implementing Regulation (EU) 2019/2072 and/or in the relevant legislative texts for emergency measures and if the specific country is in the scope of those emergency measures. The assessment was restricted to whether or not the applicant country implements those measures.
- Assessed the effectiveness of the measures described in the Dossier for those Union quarantine pests for which no specific measures are in place for the importation of the commodity from the UK and other relevant pests present in the UK and associated with the commodity.

Risk management decisions are not within EFSA's remit. Therefore, the Panel provided a rating based on expert judgement regarding the likelihood of pest freedom for each relevant pest given the risk mitigation measures proposed by DEFRA of the UK.

2 | DATA AND METHODOLOGIES

2.1 Data provided by DEFRA of the UK

The Panel considered all the data and information (hereafter called 'the Dossier') provided by DEFRA of the United Kingdom (UK) in September 2023 including the additional information provided in October and November 2024, after EFSA's request. The Dossier is managed by EFSA.

The structure and overview of the Dossier is shown in Table 1. The number of the relevant section is indicated in the Opinion when referring to a specific part of the Dossier.

Dossier Section	Overview of contents	Filename
1.1	Technical dossier for Populus alba	Populus alba commodity information final
1.2	Technical dossier for Populus nigra	Populus nigra commodity information final
1.3	Technical dossier for Populus tremula	Populus tremula commodity information final
2.0	Pest list	Populus Pest List_Final
3.1	Producers sample product list for Populus alba	Populus_alba_producers_sample_product_list
3.2	Producers sample product list for Populus nigra	Populus_nigra_producers_sample_product_list
3.3	Producers sample product list for Populus tremula	Populus_tremula_producers_sample_product_list
		(Continues)

 TABLE 1
 Structure and overview of the Dossier.

⁵In accordance with the Agreement on the withdrawal of the United Kingdom of Great Britain and Northern Ireland from the European Union and the European Atomic Energy Community, and in particular Article 5(4) of the Windsor Framework in conjunction with Annex 2 to that Framework, for the pruposes of this Opinion, references to the United Kingdom do not include Northern Ireland.

Dossier Section	Overview of contents	Filename
4.1	Distribution of Populus alba	Populus_alba_distribution
4.2	Distribution of Populus nigra	Populus_nigra_distribution
4.3	Distribution of Populus tremula	Populus_tremula_distribution
5.1	Additional information: answers	Populus additional information 17 October 2024
5.2	Additional information: pests	Pest_Query_Populus_2024
5.3	Additional information: answers	Populus additional information 26 Nov 2024

The data and supporting information provided by DEFRA of the UK formed the basis of the commodity risk assessment. Table 2 shows the main data sources used by DEFRA of the UK to compile the Dossier (Dossier Sections 1.1, 1.2, 1.3, 2.0, 3.1, 3.2, 3.3, 4.1, 4.2, 4.3, 5.1, 5.2 and 5.3).

TABLE 2	Databases used in the literature searches by DEFRA	of the UK
---------	--	-----------

Database	Platform/link
AHDB	https://ahdb.org.uk/
Aphids on the World's Plants	https://www.aphidsonworldsplants.info/
Bark and Ambrosia Beetles of the Americas	https://www.barkbeetles.info/americas_index.php
British Bugs	https://www.britishbugs.org.uk/index.html
British leafminers	https://www.leafmines.co.uk/index.htm
CABI Crop Protection Compendium	https://www.cabi.org/cpc/
CABI Plantwise Plus	https://plantwiseplusknowledgebank.org/
Checklist of the British & Irish Basidiomycota	https://basidiochecklist.science.kew.org/
Current British Aphid Checklist	https://influentialpoints.com/aphid/Checklist_of_aphids_in_Britain.htm
Database of Insects and their Food Plants	https://dbif.brc.ac.uk/homepage.aspx
Descriptions of Plant Viruses	https://www.dpvweb.net/
EPPO Global Database	https://gd.eppo.int/
EU-Nomen	https://www.eu-nomen.eu/portal/index.php
FAO	https://agris.fao.org/
Fera	https://www.fera.co.uk/ncppb
GBIF	https://www.gbif.org/
Hantsmoths	https://www.hantsmoths.org.uk/index.php
HOSTS - a Database of the World's Lepidopteran Hostplants	https://data.nhm.ac.uk/dataset/hosts
Index Fungorum	https://www.indexfungorum.org/names/Names.asp
Insects (Insecta) of the World	https://insecta.pro/
Lepidoptera and some other life forms	https://ftp.funet.fi/pub/sci/bio/life/intro.html
Lepidoptera and their ecology	https://www.pyrgus.de/index_en.php
Lepiforum e.V.	https://lepiforum.org/
Mycobank	https://www.mycobank.org/
Natural History Museum	https://www.nhm.ac.uk/
NBN atlas	https://nbnatlas.org/
NorfolkMoths	https://www.norfolkmoths.co.uk/
Plant Parasites of Europe	https://bladmineerders.nl/
Scalenet	https://scalenet.info/catalogue/
Spider Mites Web	https://www1.montpellier.inra.fr/CBGP/spmweb/
The leaf and stem mines of British flies and other isects	http://www.ukflymines.co.uk/index.php
The Sawflies (Symphyta) of Britain and Ireland	https://www.sawflies.org.uk/
Thrips of the British Isles	https://keys.lucidcentral.org/keys/v3/british_thrips/overview.html
TortAI	https://idtools.org/id/leps/tortai/index.html
Tortricid.net	http://www.tortricidae.com/
UK Beetle Recording	https://coleoptera.org.uk/home

TABLE 2 (Continued)							
Database	Platform/link						
UKmoths	https://ukmoths.org.uk/						
UK Plant Health Risk Register	https://planthealthportal.defra.gov.uk/pests-and-diseases/uk-plant-healt h-risk-register/index.cfm						
USDA Fungal Databases	https://fungi.ars.usda.gov/						
Woodland trust	https://www.woodlandtrust.org.uk/						

2.2 | Literature searches performed by EFSA

Literature searches in different databases were undertaken by EFSA to complete a list of pests potentially associated with *Populus alba, Populus nigra* and *Populus tremula*. The following searches were combined: (i) a general search to identify pests reported on *P. alba, P. nigra* and *P. tremula* in the databases, (ii) a search to identify any EU quarantine pest reported on *Populus* as genus and subsequently (iii) a tailored search to identify whether the above pests are present or not in the UK. The searches were run between May and June 2024. No language, date or document type restrictions were applied in the search strategy.

The Panel used the databases indicated in Table 3 to compile the list of pests associated with *P. alba, P. nigra* and *P. tremula*. As for Web of Science, the literature search was performed using a specific, ad hoc established search string (see Appendix B). The string was run in 'All Databases' with no range limits for time or language filters. This is further explained in Section 2.3.2.

Database	Platform/link
Aphids on World Plants	https://www.aphidsonworldsplants.info/C_HOSTS_AAIntro.htm
BIOTA of New Zealand	https://biotanz.landcareresearch.co.nz/
CABI Crop Protection Compendium	https://www.cabi.org/cpc/
Database of Insects and their Food Plants	https://www.brc.ac.uk/dbif/hosts.aspx
Database of the World's Lepidopteran Hostplants	https://www.nhm.ac.uk/our-science/data/hostplants/search/index.dsml
EPPO Global Database	https://gd.eppo.int/
EUROPHYT	https://food.ec.europa.eu/plants/plant-health-and-biosecurity/europhyt_en
Leaf-miners	https://www.leafmines.co.uk/html/plants.htm
Nemaplex	https://nemaplex.ucdavis.edu/Nemabase2010/PlantNematodeHostStatusDD Query.aspx
Plant Parasites of Europe	https://bladmineerders.nl/
Plant Pest Information Network	https://www.mpi.govt.nz/news-and-resources/resources/registers-and-lists/ plant-pest-information-network/
Scalenet	https://scalenet.info/associates/
Scolytinae hosts and distribution database	https://www.scolytinaehostsdatabase.eu/site/it/home/
Spider Mites Web	https://www1.montpellier.inra.fr/CBGP/spmweb/
USDA ARS Fungal Database	https://fungi.ars.usda.gov/
Web of Science: All Databases (Web of Science Core Collection, CABI: CAB Abstracts, BIOSIS Citation Index, Chinese Science Citation Database, Current Contents Connect, Data Citation Index, FSTA, KCI-Korean Journal Database, Russian Science Citation Index, MEDLINE, SciELO Citation Index, Zoological Record)	Web of Science https://www.webofknowledge.com
World Agroforestry	https://www.worldagroforestry.org/treedb2/speciesprofile.php?Spid=1749

TABLE 3 Databases used by EFSA for the compilation of the pest list associated with Populus alba, P. nigra and P. tremula.

Additional documents were retrieved when developing the Opinion. The available scientific information, including previous EFSA opinions on the relevant pests and diseases (see pest data sheets in Appendix A) and the relevant literature and legislation (e.g. Regulation (EU) 2016/2031; Commission Implementing Regulations (EU) 2018/2019; (EU) 2018/2018 and (EU) 2019/2072) were taken into account.

2.3 | Methodology

When developing the Opinion, the Panel followed the EFSA Guidance on commodity risk assessment for the evaluation of high risk plant dossiers (EFSA PLH Panel, 2019).

In the first step, pests potentially associated with the commodity in the country of origin (EU-quarantine pests and other pests) that may require risk mitigation measures are identified. The EU non-quarantine pests not known to occur in the EU were selected based on evidence of their potential impact in the EU. After the first step, all the relevant pests that may need risk mitigation measures were identified.

In the second step, the implemented risk mitigation measures for each relevant pest were evaluated.

A conclusion on the pest freedom status of the commodity for each of the relevant pests was determined and uncertainties identified using expert judgements.

Pest freedom was assessed by estimating the number of infested/infected units out of 10,000 exported units. Further details on the methodology used to estimate the likelihood of pest freedom are provided in Section 2.3.4.

2.3.1 | Commodity data

Based on the information provided by DEFRA of the UK the characteristics of the commodity were summarised.

2.3.2 | Identification of pests potentially associated with the commodity

To evaluate the pest risk associated with the importation of the commodity from the UK, a pest list was compiled. The pest list is a compilation of all identified plant pests reported as associated with *P. alba, P. nigra* and *P. tremula* based on information provided in the Dossier Sections 1.1, 1.2, 1.3, 2.0, 3.1, 3.2, 3.3, 4.1, 4.2, 4.3, 5.1, 5.2 and 5.3, on searches performed by the Panel, and of EU quarantine and protected zone quarantine pests associated with *Populus* as a genus. The search strategy and search syntax were adapted to each of the databases listed in Table 3, according to the options and functionalities of the different databases and CABI keyword thesaurus.

The scientific names of the host plant (i.e. *Populus alba, Populus nigra, Populus tremula, Populus*) were used when searching in the EPPO Global database and CABI Crop Protection Compendium. The same strategy was applied to the other databases excluding EUROPHYT and Web of Science.

EUROPHYT was investigated by searching for the interceptions associated with *P. alba, P. nigra* and *P. tremula* imported from the whole world from 1995 to May 2020 and TRACES-NT from May 2020 to 30 November 2024, respectively. For the pests selected for further evaluation, a search in the EUROPHYT and/or TRACES-NT was performed for the interceptions from the whole world, at species level, for all the available years until 30 September 2024.

The search strategy used for Web of Science Databases was designed combining English common names for pests and diseases, terms describing symptoms of plant diseases and the scientific and English common names of the commodity and excluding pests which were identified using searches in other databases. The established search strings are detailed in Appendix B and they were run on 14 June 2024.

The titles and abstracts of the scientific papers retrieved were screened and the pests associated with *P. alba, P. nigra* and *P. tremula* were included in the pest list. The pest list was eventually further compiled with other relevant information (e.g. EPPO code per pest, taxonomic information, categorisation, distribution) useful for the selection of the pests relevant for the purposes of this Opinion.

The compiled pest list (see Microsoft Excel[®] in Appendix F) includes all identified pests that use as host *P. alba, P. nigra* and *P. tremula* as well as all EU quarantine pests and protected zone quarantine pests found to be associated with *Populus* as a genus.

The evaluation of the compiled pest list was done in two steps: first, the relevance of the EU-quarantine pests was evaluated (Section 4.1); second, the relevance of any other plant pest was evaluated (Section 4.2).

Pests for which limited information was available on one or more criteria used to identify them as relevant for this Opinion, e.g. on potential impact, are listed in Appendix E (List of pests that can potentially cause an effect not further assessed).

2.3.3 | Listing and evaluation of risk mitigation measures

All implemented risk mitigation measures were listed and evaluated. When evaluating the likelihood of pest freedom of the commodity, the following types of potential infection/infestation sources for *P. alba, P. nigra* and *P. tremula* in export nursery were considered (see also Figure 1):

- pest entry from surrounding areas,
- · pest entry with new plants/seeds,
- pest spread within the nursery.



FIGURE 1 Conceptual framework to assess likelihood that plants are exported free from relevant pests. (Source: EFSA PLH Panel, 2019).

The risk mitigation measures proposed by DEFRA of the UK were evaluated with Expert Knowledge Elicitation (EKE) according to the Guidance on uncertainty analysis in scientific assessment (EFSA Scientific Committee, 2018).

Information on the biology, likelihood of entry of the pest to the export nursery, of its spread inside the nursery and the effect of measures on the specific pests were summarised in data sheets of pests selected for further evaluation (see Appendix A).

2.3.4 | Expert Knowledge Elicitation

To estimate the pest freedom of the commodities an EKE was performed following EFSA guidance (Annex B.8 of EFSA Scientific Committee, 2018). The specific question for EKE was: 'Taking into account (i) the risk mitigation measures in place in the nurseries and (ii) other relevant information, how many of 10,000 commodity units, either single plants or bundles of plants will be infested with the relevant pest when arriving in the EU? A unit is defined as either single plants or bundles of plants, bare root plants or plants in pots, depending on the commodity.

For the purpose of the EKE, the commodities (see Section 3.1) were grouped as follows:

- 1. Cuttings/Graftwood of 1-2 years, in bundles of 10-20 items.
- 2. Bare root plants of 1–7 years as single trees or in bundles of 5, 10, 15, 25, 50 plants depending on the species and plant size.
- 3. Cell grown plants of 1–2 years as single plants or bundled in 5–10 plants depending on the plant size.
- 4. Single rooted plants of 3–15 years in pots.

Single plants and bundles of plants were considered together during the EKE. The following reasoning is given for not distinguishing bundles of bare root plants and bundles of cell grown plants from their respective single plants:

- (i) There is no quantitative information available regarding clustering of plants during production;
- (ii) Single plants are grouped in bundles after sorting;
- (iii) For the pests under consideration, a cross-contamination during transport is possible;
- (iv) Bundles of small plants resemble in their risk larger single plants.

The uncertainties associated with the EKE were taken into account and quantified in a probability distribution fitted to the elicited percentiles, applying the semi-formal method described in Section 3.5.2 of the EFSA-PLH Guidance on quantitative pest risk assessment (EFSA PLH Panel, 2018). Finally, the EKE results were reported in terms of the likelihood of pest freedom, calculated by 1 minus the likelihood to be infested. The lower 5% percentile of the uncertainty distribution reflects the opinion that pest freedom is with 95% certainty above this limit.

3 | COMMODITY DATA

3.1 | Description of the commodity

The commodities to be imported from the UK to the EU are cuttings/graftwood, bare root plants, cell grown plants and rooted plants up to 15 years in pots of *P. alba* (common names: white poplar, silver-leaved poplar; Family: Salicaceae), *P. nigra* (common names: black poplar; Family: Salicaceae) and *P. tremula* (common names: aspen, european aspen, trembling poplar; Family: Salicaceae) as described in the details below:

- 1. Cuttings/Graftwood (only for *P. nigra* and *P. tremula*): the age of cuttings/graftwood is between 1 and 2 years (Dossier Sections 1.2 and 1.3). The diameter is between 0.8 and 1.2 cm. They are grouped in bundles of 10–20 items. Cuttings/Graftwoods are strong young shoots bearing buds which are suitable for use in chip budding or grafting. The shoots are approximately between 35 and 40 cm long and will typically have 9, 10 or more buds present (Dossier Sections 1.2, 1.3 and 5.1).
- **2. Bare root plants**: the age of plants is between 1 and 7 years (Dossier Sections 1.1, 1.2, 1.3 and 5.1). The diameter is between 0.5 and 4 cm and height is between 50 and 200 cm. Bare root plants may have some leaves at the time of export, in particular when exported in early winter (Dossier Sections 1.1, 1.2, 1.3 and 5.1). Bare root plants will be exported as single trees or in bundles of 5, 10, 15, 25, 50. (Dossier Sections 1.1, 1.2 and 5.1).
- **3. Cell grown plants**: the age of plants is between 1 and 2 years. The diameter is between 0.5 and 1 cm and height between 40 and 60 cm. Cell grown plants are plants grown in cells at one plant per cell, using EU-compliant growing media. These may be grown in greenhouses initially but are subsequently grown outdoors in containers in metal frames above the ground. Cell grown plants may be traded as individual plants or as bundles. Typically, bundles will include 5–10 plants depending on the size of plant. The cell grown plants may be exported with leaves based on the picture 'cell grown plants bundled ready for dispatch' provided by the applicant country (Dossier Sections 1.1, 1.2, 1.3 and 5.1).
- 4. Rooted plants in pots: the age of plants is between 3 and 15 years (Dossier Section 5.1). The diameter is between 1.5 and 12 cm and height between 1 m and 10 m. Rooted plants in pots may be either grown in EU-compliant growing media in pots for their whole life, or initially grown in the field before being lifted, root-washed to remove any soil and then potted in EU-compliant growing media. The trees will be lifted from the field a minimum of one growing season prior to export at no more than 6 years old. The plants in pots may be exported with leaves, depending on the timing of the export (Dossier Sections 1.1, 1.2, 1.3 and 5.1).

The growing media used is either virgin peat or peat-free compost (a mixture of coir, tree bark, wood fibre, etc.) complying with the requirements for growing media as specified in the Annex VII of the Commission Implementing Regulation 2019/2072. This growing media is certified and heat-treated by commercial suppliers during production to eliminate pests and diseases (Dossier Sections 1.1, 1.2 and 1.3).

According to ISPM 36 (FAO, 2019), the commodities can be classified as 'bare root plants' and 'rooted plants in pots'. The yearly average trade volume of the different commodities to the EU is reported in Dossier Sections 1.1, 1.2 and 1.3 and summarised in Table 4. The trade of these commodities will mainly be to Northern Ireland and the Republic of Ireland.

Type of plant	Number of items	Seasonal timing
Populus alba		
Bare root plants	20,000	November to April
Rooted plants in pots (including cell grown plants)	25,000	Mainly September to May
Populus nigra		
Cuttings/graftwood	500	January to March
Bare root plants	10,000	November to April
Rooted plants in pots (including cell-grown plants)	20,000	Mainly September to May
Populus tremula		
Cuttings/graftwood	500	January to March
Bare root plants	50,000	November to April
Rooted plants in pots (including cell grown plants)	20,000	Mainly September to May

TABLE 4 Trade volumes of *Populus alba, Populus nigra* and *Populus tremula* commodities.

According to the Dossier Sections 1.1, 1.2 and 1.3, the intended use of the commodities is as follows. Plants are supplied directly to professional operators and traders. Uses may include propagation, growing-on, onward trading or onward sales to final customers but will generally fall into the following categories:

Tree production and further growing-on by professional operators; Landscapers and garden centres, for woodland and ornamental/landscape planting; Direct sales to final users as ornamentals.

3.2 | Description of the production areas

There are four nurseries specified the technical dossier from the UK producing the commodities (Dossier Sections 1.1, 1.2 and 1.3). *Populus* species are grown in Great Britain in line with the Plant Health (Amendment etc.) (EU Exit) Regulations 2020⁶ and the Plant Health (Phytosanitary Conditions) (Amendment) (EU Exit) Regulations 2020.⁷ These regulations are broadly similar to the EU phytosanitary regulations. All plants within the UK nurseries are grown under the same phytosanitary measures, meeting the requirements of the UK Plant Passporting regime (Dossier Sections 1.1, 1.2 and 1.3).

The size of the nurseries is between 8 and 150 ha for container stock (plants in pots) and up to 325 ha for field-grown stock (Dossier Sections 1.1, 1.2 and 1.3).

The nurseries also grow other plant species as shown in the Appendix C. The minimum and maximum proportion of *Populus* compared to the other plant species grown in the nurseries is between 0.5% and 1% for *P. alba* and *P. nigra*, and between 0.5% and 3% for *P. tremula* (Dossier Sections 1.1, 1.2 and 1.3). The following plant species may be grown in some of the nurseries: *Castanea sativa*, *Larix* spp., *Fagus sylvatica*, *Fagus* spp., *Malus* spp., *Quercus petraea*, *Quercus pubescens*, *Quercus robur*, *Quercus* spp., *Rosa* spp., *Sorbus* spp., *Ulmus* spp. and *Viburnum* spp. (Dossier Section 5.1). There are nurseries which also produce plants for the local market, and there is no distancing between production areas for the export and the local market (Dossier Sections 1.1, 1.2 and 1.3).

Non-cultivated herbaceous plants grow on less than 1% of the nursery area. The predominant species is rye grass (*Lolium* spp.). Other identified species include dandelions (*Taraxacum officinale*), hairy bittercress (*Cardamine hirsuta*), common daisy (*Bellis perennis*), creeping cinquefoil (*Potentilla reptans*) and bluebells (*Hyacinthoides non-scripta*). These are all extremely low in number (Dossier Sections 1.1, 1.2 and 1.3). In access areas, non-cultivated herbaceous plants are kept to a minimum and only exist at nursery boundaries.

There are hedges surrounding the export nurseries made up of a range of species including hazel (*Corylus avellana*), yew (*Taxus baccata*), holly (*Ilex* spp.), ivy (*Hedera* spp.), alder (*Alnus glutinosa*), cherry laurel (*Prunus laurocerasus*), hawthorn (*Crataegus* spp.), blackthorn (*Prunus spinosa*) and leylandii (*Cupressus × leylandii*) (Dossier Sections 1.1, 1.2 and 1.3).

The minimum distance in a straight line, between the growing area in the nurseries and the closest *P. alba* plants in the local surroundings is 10 metres and the closest *P. nigra* or *P. tremula* plants in the local surroundings is 20 metres (Dossier Sections 1.1, 1.2 and 1.3).

Nurseries are predominately situated in rural areas. The surrounding land tend to be arable farmland with some pasture for animals and small areas of woodland. Hedges are often used to define field boundaries and grown along roadsides (Dossier Sections 1.1, 1.2 and 1.3).

Arable crops present around the nurseries are rotated in line with good farming practices and could include oilseed rape (*Brassica napus*), wheat (*Triticum spp.*), barley (*Hordeum vulgare*), turnips (*Brassica rapa subsp. rapa*), potatoes (*Solanum tuberosum*) and maize (*Zea mays*) (Dossier Sections 1.1, 1.2 and 1.3).

Pastures present around the nurseries are predominantly ryegrass (Lolium spp.) (Dossier Sections 1.1, 1.2 and 1.3).

Woodland is present around the nurseries. Woodlands tend to be a standard UK mixed woodland, with a range of the UK native trees such as oak (*Quercus robur*), pine (*Pinus spp.*), poplar (*Populus spp.*), ash (*Fraxinus spp.*), sycamore (*Acer pseudo-platanus*), holly (*Ilex spp.*), Norway maple (*Acer platanoides*) and field maple (*Acer campestre*). The nearest woodland to one of the nurseries borders the boundary fence (Dossier Sections 1.1, 1.2 and 1.3).

It is not possible to identify the plant species growing within the gardens of private dwellings around the nurseries (Dossier Sections 1.1, 1.2 and 1.3).

The following plant species may be grown within a 2 km zone surrounding the nurseries: *Brassica* spp. (cultivated species), *Camellia* spp., *Castanea* sativa, Larix kaempferi, Larix spp., *Fagus* sylvatica, *Fagus* spp., *Malus* spp., *Morus* spp., *Quercus* petraea, *Quercus* pubescents, *Quercus* robur, *Quercus* spp., *Rhododendron* spp., *Rosa* spp., *Rubus* spp., *Solanum* lycopersicum, *Sorbus* spp., *Taraxacum* officinalis, Ulmus spp., Urtica dioica and Viburnum spp. (Dossier Section 5.1).

Based on the global Köppen–Geiger climate zone classification (World Maps of Köppen-Geiger climate classification), the climate of the production areas of *P. alba, P. nigra* and *P. tremula* in the UK is classified as Cfb, i.e. main climate (C): warm temperate; precipitation (f): fully humid; temperature (b): warm summer.

3.3 Production and handling processes

3.3.1 | Source of planting material

The starting material of the commodities is a mix of seeds and seedlings depending on the nursery (Dossier Sections 1.1, 1.2 and 1.3).

Seeds purchased in the UK are certified under the Forest Reproductive Material (Great Britain) Regulations 2002. Seedlings sourced in the UK are certified with the UK Plant Passports. A small percentage of seedlings are obtained from EU countries (the Netherlands, Belgium, France) and they are certified with phytosanitary certificates (Dossier Sections 1.1, 1.2 and 1.3).

⁶Plant Health (Amendment etc.) (EU Exit) Regulations 2020 of 14 December 2020, No. 1482, 80 pp. https://www.legislation.gov.uk/uksi/2020/1482/contents/made ⁷Plant Health (Phytosanitary Conditions) (Amendment) (EU Exit) Regulations 2020, No. 1527, 276 pp. https://www.legislation.gov.uk/uksi/2020/1527/contents/made

None of the nurseries expected to export to the EU produce plants from grafting, they use only seed, seedlings and cuttings, therefore there are no mother plants of *P. alba*, *P. nigra* or *P. tremula* present in the nurseries (Dossier Sections 1.1, 1.2 and 1.3).

3.3.2 | Production cycle

Plants are either grown in containers (cells, pots, tubes, etc.) or in the field. Cell grown plants can be grown in greenhouses; however, most plants will be field-grown or field-grown in containers (Dossier Sections 1.1 and 1.2). The minimum distance between greenhouses and production fields of *Populus* is 30 m (Dossier Section 5.2).

As the plants are intended for outdoor cultivation it is normally only the early growth stages that are maintained under protection, such as young plants where there is an increased vulnerability due to climatic conditions including frost. The commodity to be exported should therefore be regarded as outdoor grown. Growth under protection is primarily to protect against external climatic conditions rather than protection from pests. The early stages of plants grown under protection are maintained in plastic polytunnels, or in glasshouses which typically consist of a metal or wood frame construction and glass panels (Dossier Sections 1.1, 1.2, 1.3 and 5.1).

Rooted plants in pots may be either grown in EU-compliant growing media in pots for their whole life, or initially grown in the field before being lifted, root-washed to remove the soil and then potted in EU-compliant growing media. Trees will be lifted from the field, root-washed to remove the soil and transplanted into pots at least one growing season before export (Dossier Section 5.1).

Specimen trees may either be grown in pots in EU-compliant media their whole life or be initially grown in the field, lifted at no more than 6 years old, root-washed and subsequently grown from that point on in pots in EU-compliant growing media (Dossier Sections 1.1, 1.2, 1.3 and 5.1). Trees will be lifted from the field, root-washed to remove the soil and transplanted into pots at least one growing season before export (Dossier Section 5.1).

Pruning is done on the commodities 1, 2 and 4 described above in 3.1 Pruning frequency depends on growth, age of plant, nursery and customer preference. Cell grown plants are not pruned (Dossier Sections 1.1, 1.2, 1.3 and 5.1).

According to the Dossier Sections 1.1, 1.2 and 1.3, bare root plants are harvested in winter to be able to lift plants from the field, and because this is the best time to move dormant plants. Rooted plants in pots can be moved at any point in the year to fulfil customer demand.

The growing media is virgin peat or peat-free compost. This compost is heat-treated by commercial suppliers during production to eliminate pests and diseases. It is supplied in sealed bulk bags or shrink-wrapped bales and stored off the ground on pallets, these are free from contamination. Where delivered in bulk, compost is kept in a dedicated bunker, either indoors, or covered by tarpaulin outdoors, and with no risk of contamination with soil or other material (Dossier Sections 1.1, 1.2 and 1.3).

Overhead, sub irrigation or drip irrigation is applied. Water used for irrigation can be drawn from several sources, the mains supply, bore holes or from rainwater collection or watercourses (Dossier Sections 1.1, 1.2 and 1.3). Additional information on water used for irrigation is provided in Appendix D. Regardless of the source of the water used to irrigate, none of the nurseries are known to have experienced the introduction of a pest/disease because of contamination of the water supply (Dossier Sections 1.1, 1.2 and 1.3).

Growers are required to assess whether water sources, irrigation and drainage systems used in plant production could harbour and transmit plant pests. Water is routinely sampled and sent for analysis (Dossier Sections 1.1, 1.2 and 1.3).

Growers must have an appropriate programme of weed management in place in the nursery (Dossier Sections 1.1, 1.2 and 1.3).

General hygiene measures are undertaken as part of routine nursery production, including disinfection of tools and equipment between batches/lots and different plant species. The tools are dipped in a disinfectant solution and wiped with a clean cloth between trees to reduce the risk of viral and bacterial transfer between subjects. There are various disinfectants available, with Virkon S (active substance: potassium peroxymonosulfate and sodium chloride) being a common example (Dossier Sections 1.1, 1.2 and 1.3).

Growers keep records to allow traceability for all plant material handled. These records must allow a consignment or consignment in transit to be traced back to the original source, as well as forward to identify all trade customers to which those plants have been supplied (Dossier Sections 1.1, 1.2 and 1.3).

3.3.3 | Pest monitoring during production

All producers are registered as professional operators with the UK Competent Authority via the Animal and Plant Health Agency (APHA) for England and Wales, or with Science and Advice for Scottish Agriculture (SASA) for Scotland, and are authorised to issue UK plant passports, verifying they meet the required national sanitary standards. The Competent Authority inspects crops at least once a year to check they meet the standards set out in the guides. The UK surveillance is based on visual inspection with samples taken from symptomatic material, and where appropriate, samples are also taken from asymptomatic material (e.g. plants, tubers, soil, watercourses) (Dossier Sections 1.1, 1.2 and 1.3).

The sanitary status of production areas is controlled by the producers as part of these schemes, as well as via official inspections by APHA Plant Health and Seeds Inspectors (PHSI; England and Wales) or with SASA (Scotland) (Dossier Sections 1.1, 1.2 and 1.3).

13 of 120

Plant material is regularly monitored for plant health issues. Pest monitoring is carried out visually by trained nursery staff via regular crop walking and records are kept of this monitoring. Qualified agronomists also undertake crop walks to verify the producer's assessments. Curative or preventative actions as described below are implemented together with an assessment of phytosanitary risk. Unless a pest can be immediately and definitively identified as non-quarantine, growers are required to treat it as a suspect quarantine pest and notify the Competent Authority. All plants are also carefully inspected by nurseries on arrival and dispatch for any plant health issues (Dossier Sections 1.1, 1.2 and 1.3).

The nurseries follow the Plant Health Management Standard issued by the Plant Healthy Certification Scheme which DEFRA, the Royal Horticultural Society (Dossier Sections 1.1, 1.2 and 1.3).

During production, in addition to the general health monitoring of the plants by the nurseries, official growing season inspections are undertaken by the UK Plant Health Service at an appropriate time, taking into consideration factors such as the likelihood of pest presence and growth stage of the crop. Where appropriate this could include sampling and laboratory analysis. Official sampling and analysis could also be undertaken nearer to the point of export depending on the type of analysis and the import requirements of the country being exported to. Samples are generally taken on a representative sample of plants, in some cases however where the consignment size is quite small all plants are sampled. Magnification equipment is provided to all inspectors as part of their standard equipment and is used during inspections when appropriate (Dossier Sections 1.1, 1.2 and 1.3).

In the Dossier it is reported that in the last 3 years there has been a substantial level of inspection of registered *P. alba, P. nigra* and *P. tremula* producers, both in support of the Plant Passporting scheme (checks are consistent with EU legislation, with a minimum of 1 a year for authorised operators) and as part of the Quarantine Surveillance programme (Great Britain uses the same framework for its surveillance programme as the EU) The number of inspected nurseries were 4 in 2020 and 5 in 2021 and 2022. Inspections targeted *P. ramorum* but plants were also inspected for symptoms and signs of other pests including quarantine pests. (Dossier Sections 1.1, 1.2 and 1.3).

All residues or waste materials are reported to be assessed for the potential to host, harbour and transmit pests (Dossier Sections 1.1, 1.2 and 1.3).

Incoming plant material and other goods such as packaging material and growing media, that have the potential to be infected or harbour pests, are checked on arrival. Growers have procedures in place to quarantine any suspect plant material and to report findings to the authorities (Dossier Sections 1.1, 1.2 and 1.3).

3.3.4 | Pest management during production

Crop protection is achieved using a combination of measures including approved plant protection products, biological control or physical measures. Plant protection products are only used when necessary and records of all plant protection treatments are kept (Dossier Sections 1.1, 1.2 and 1.3).

Pest and disease pressure varies from season to season. Product application takes place only when required and depends on situation (disease pressure, growth stage etc. and environmental factors) at that time. Subject to this variation in pest pressure, in some seasons few, if any, pesticides are applied; in others it is sometimes necessary to apply preventative and/or control applications of pesticides. In many circumstances also, biological control rather than chemical control is reported to be used to manage pest outbreaks (Dossier Sections 1.1, 1.2 and 1.3).

Examples of typical treatments used against rust, leaf spot, canker, spider mites, aphids and weeds are listed in the Dossier Sections 1.1, 1.2, 1.3 and 5.1. These would be applied at the manufacturers recommended rate and intervals (Dossier Sections 1.1, 1.2 and 1.3).

There are no specific measures/treatments against soil pests. However, containerised plants are grown in trays on top of protective plastic membranes to prevent contact with soil. Membranes are regularly refreshed when needed. Alternatively, plants may be grown on raised galvanised steel benches stood on gravel as a barrier between the soil and bench feet and/ or concreted surfaces (Dossier Sections 1.1, 1.2 and 1.3).

Post-harvest and through the autumn and winter, nursery management is centred on pest and disease prevention and maintaining good levels of nursery hygiene. Leaves, pruning residues and weeds are all removed from the nursery to reduce the number of over wintering sites for pests and diseases (Dossier Sections 1.1, 1.2 and 1.3).

3.3.5 | Inspections before export

The UK NPPO carries out inspections and testing where required by the country of destination's plant health legislation, to ensure all requirements are fulfilled and a valid phytosanitary certificate with the correct additional declarations is issued (Dossier Sections 1.1, 1.2 and 1.3).

Separate to any official inspection, plant material is checked by growers for plant health issues prior to dispatch (Dossier Sections 1.1, 1.2 and 1.3).

A final pre-export inspection is undertaken as part of the process of issuing a phytosanitary certificate. These inspections are generally undertaken as near to the time of export as possible, usually within 1–2 days and not more than 2 weeks before export. Phytosanitary certificates are only issued if the commodity meets the required plant health standards after inspection and/or testing according to appropriate official procedures (Dossier Sections 1.1, 1.2 and 1.3).

The protocol for plants infested by pests during inspections before export is to treat the plants, if they are on site for a sufficient period of time or to destroy any plants infested by pests otherwise. All other host plants in the nursery would be treated. The phytosanitary certificate for export will not be issued until the UK Plant Health inspectors confirm that the plants are free from pests (Dossier Sections 1.1, 1.2 and 1.3).

3.3.6 | Export procedure

Bare root plants, harvested from November to March, are lifted and washed free from soil with a low-pressure washer in the outdoors nursery area away from packing/cold store area. In some cases, the plants may be kept in a cold store for up to 5 months after harvesting prior to export (Dossier Sections 1.1, 1.2 and 1.3).

Rooted plants in pots can be moved at any point in the year to fulfil customer demand. These will likely be destined for garden centre trade rather than nurseries (Dossier Sections 1.1, 1.2 and 1.3).

Graftwood/budwood is wrapped in plastic and packed in cardboard boxes or Dutch crates on ISPM certified wooden pallets, or metal pallets, dependant on quantity. Graftwood/budwood may be exported in bundles of 10–20 items (Dossier Sections 1.1, 1.2 and 1.3).

Cell grown plants may be traded as individual plants or as bundles. Typically, bundles will include 5–10 plants depending on the size of plant (Dossier Section 5.1).

Prior to export bare root plants can be placed in bundles 5–50 plants, depending on their size or single bare root trees. They are then wrapped in polythene and packed and distributed on ISPM 15 certified wooden pallets, or metal pallets. Alternatively, they may be placed in pallets which are then wrapped in polythene. Small volume orders may be packed in waxed cardboard cartons or polythene bags and dispatched via courier (Dossier Sections 1.1, 1.2 and 1.3).

Rooted plants in pots are transported on Danish trolleys for smaller containers, or ISPM 15 certified pallets, or individually in pots for larger containers (Dossier Sections 1.1, 1.2 and 1.3).

The preparation of the commodities for export is carried out inside the nurseries in a closed environment, e.g. packing shed (Dossier Sections 1.1, 1.2 and 1.3).

Plants are transported by lorry (size dependant on load quantity). Cold sensitive plants are occasionally transported by temperature-controlled lorry if weather conditions during transit are likely to be very cold (Dossier Sections 1.1, 1.2 and 1.3).

4 | IDENTIFICATION OF PESTS POTENTIALLY ASSOCIATED WITH THE COMMODITY

The search for potential pests associated with the commodity rendered 1657 species (see Microsoft Excel® file in Appendix F).

4.1 Selection of relevant EU-quarantine pests associated with the commodity

The EU listing of union quarantine pests and protected zone quarantine pests (Commission Implementing Regulation (EU) 2019/2072) is based on assessments concluding that the pests can enter, establish, spread and have potential impact in the EU.

Fifty-five EU-quarantine species that are reported to use commodity as a host plant were evaluated (Table 5) for their relevance of being included in this Opinion.

The relevance of an EU-quarantine pest for this Opinion was based on evidence that:

a. the pest is present in the UK;

b. the commodity is host of the pest;

c. one or more life stages of the pest can be associated with the specified commodity.

Pests that fulfilled all criteria were selected for further evaluation.

Table 5 presents an overview of the evaluation of the 55 EU-quarantine pest species that are reported as associated with the commodity.

Of these 55 EU-quarantine pest species evaluated, 2 (*Bemisia tabaci* (European populations) and *Entoleuca mammata*) are present in the UK and can be associated with the commodity and hence were selected for further evaluation.

There were two EU quarantine pests, despite being reported to be associated with Populus were not further evaluated (*Agrilus anxius* and *Phytophthora ramorum*).

An association with *Populus* was reported in CABI for *A. anxius* without providing the original source (CABI, 2020). The consultation of other literature revealed that *Populus* is not reported as a host of *A. anxius* and that *A. anxius* is a specialist on *Betula*. Moreover, the pest is not known to be present in the UK. *Populus deltoides* was reported to be a putative host of *P. ramorum*. However, the host status was not confirmed by the fulfilment of the Koch's postulates (Vettraino et al., 2010). Therefore, there is no indication that poplars are hosts of the pathogen.

TABLE 5 Overview of the evaluation of the 55 EU-quarantine pest species for which information was found in the Dossier, databases and literature searches that use *Populus* as a host plant for their relevance for this Opinion.

No.	Pest name according to EU legislation ^a	EPPO code	Group	Pest present in the UK	Populus confirmed as a host (reference)	Pest can be associated with the commodity	Pest relevant for the Opinion
1	Acleris issikii	ACLRIS	Insects	No	Populus nigra (Byun & Yan, 2004)	Not assessed	No
2	Aleurocanthus woglumi	ALECWO	Insects	No	Populus spp. (Shaw, 1950)	Not assessed	No
3	Anoplophora chinensis	ANOLCN	Insects	No	Populus alba, P. nigra (Sjöman et al., 2014)	Not assessed	No
4	Anoplophora glabripennis	ANOLGL	Insects	No	Populus alba, P. nigra, P. tremula (Sjöman et al., 2014)	Not assessed	No
5	Apriona cinerea	APRICI	Insects	No	Populus alba (EPPO, 2024), P. nigra (Singh et al., 2004)	Not assessed	No
6	Apriona germari	APRIGE	Insects	No	Populus alba (Lim et al., 2014)	Not assessed	No
7	Apriona rugicollis	APRIJA	Insects	No	Populus (EPPO, 2024)	Not assessed	No
8	Arrhenodes minutus	ARRHMI	Insects	No	Populus (Kissinger, 1993)	Not assessed	No
9	<i>Begomovirus caricae</i> as Begomoviruses	PALCUV	Viruses	No	Populus alba (Mustafa et al., 2022)	Not assessed	No
10	<i>Begomovirus solanumkeralaense</i> as Begomoviruses	TOLCKA	Viruses	No	Populus alba (Mustafa et al., 2022)	Not assessed	No
11	Bemisia tabaci (non-European populations)	BEMITA	Insects	No	Populus nigra (Samin et al., 2015)	Not assessed	No
12	<i>Bemisia tabaci</i> (European populations) ^b	BEMITA	Insects	Yes	Populus nigra (Samin et al., 2015)	Yes	Yes
13	Candidatus Phytoplasma fraxini	PHYPFR	Phytoplasmas	No	Populus nigra (Franco-Lara et al., 2020)	Not assessed	No
14	Choristoneura conflictana	ARCHCO	Insects	No	Populus alba (EPPO, <mark>2024</mark>)	Not assessed	No
15	Choristoneura rosaceana	CHONRO	Insects	No	Populus sp. (Ferguson, 1975)	Not assessed	No
16	Entoleuca mammata	ΗΥΡΟΜΑ	Fungi	Yes	Populus alba, P. tremula (Kasanen et al., 2004)	Yes	Yes
17	Eotetranychus lewisi	EOTELE	Mites	No	Populus deltoides (Estebanes-Gonzalez & Baker, 1968)	Not assessed	No
18	Euwallacea fornicatus sensu lato	XYLBFO	Insects	No	Populus alba (van Rooyen et al., 2021), Populus nigra (DAFNAE, 2024)	Not assessed	No
19	Homalodisca vitripennis	HOMLTR	Insects	No	Populus sp. (Hoddle et al., 2003)	Not assessed	No
20	Lopholeucaspis japonica	LOPLJA	Insects	No	Populus alba (Batsankalashvili et al., 2017)	Not assessed	No
21	Lycorma delicatula	LYCMDE	Insects	No	Populus alba (Dara et al., 2015)	Not assessed	No
22	Oemona hirta	OEMOHI	Insects	No	Populus alba, P. nigra (EPPO, 2024)	Not assessed	No
23	Phymatotrichopsis omnivora	PHMPOM	Fungi	No	Populus alba, P. nigra (Anonymous, 1960)	Not assessed	No
24	Phytophthora ramorum (non-EU isolates)	PHYTRA	Oomycetes	Yes	Populus deltoides (Vettraino et al., 2010)	No ^c	No
25	Popillia japonica	POPIJA	Insects	No	Populus nigra (Clausen et al., 1927)	Not assessed	No
26	Sphaerulina musiva	MYCOPP	Fungi	No	Populus nigra (Anonymous, 1960)	Not assessed	No
27	Trirachys sartus	AELSSA	Insects	No	Populus alba, P. nigra (Ahmad et al., 1977)	Not assessed	No

TABLE 5(Continued)

No.	Pest name according to EU legislation ^a	EPPO code	Group	Pest present in the UK	Populus confirmed as a host (reference)	Pest can be associated with the commodity	Pest relevant for the Opinion
28	<i>Xiphinema rivesi</i> (non-European populations)	XIPHRI	Nematodes	No	Populus sp. (Xu & Zhao, 2019)	Not assessed	No
29	Xylella fastidiosa	XYLEFA	Bacteria	No	Populus tremula (Casarin et al., 2023)	Not assessed	No
Scolyti	nae spp. (non-European)						
30	Ambrosiodmus apicalis as Scolytinae spp. (non-European)	_	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No
31	Ambrosiodmus lewisi as Scolytinae spp. (non-European)	AMBDLE	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No
32	Ambrosiodmus rubricollis as Scolytinae spp. (non-European)	AMBDRU	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No
33	Anisandrus maiche as Scolytinae spp. (non-European)	ANIDMA	Insects	No	Populus tremula (Terekhova & Skrylnik, 2012)	Not assessed	No
34	Anisandrus obesus as Scolytinae spp. (non-European)	ANIDOB	Insects	No	Populus tremuloides (Wood & Bright, 1992)	Not assessed	No
35	<i>Debus emarginatus</i> as Scolytinae spp. (non-European)	DEBUEM	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No
36	<i>Dryoxylon onoharaense</i> as Scolytinae spp. (non-European)	DRYXON	Insects	No	Populus deltoides (Atkinson, 2024)	Not assessed	No
37	<i>Euwallacea interjectus</i> as Scolytinae spp. (non-European)	XYLBIN	Insects	No	Populus spp. (Wood & Bright, 1992)	Not assessed	No
38	<i>Euwallacea validus</i> as Scolytinae spp. (non-European)	XYLBVA	Insects	No	Populus deltoides (Wood & Bright, 1992)	Not assessed	No
39	Gnathotrichus retusus as Scolytinae spp. (non-European)	GNAHRE	Insects	No	Populus tristis (Atkinson, 2024)	Not assessed	No
40	Hylocurus hirtellus as Scolytinae spp. (non-European)	-	Insects	No	Populus sp. (Atkinson, 2024)	Not assessed	No
41	Hypothenemus seriatus as Scolytinae spp. (non-European)	STEHSE	Insects	No	Populus deltoides (Wood & Bright, 1992)	Not assessed	No
42	<i>Micracis swainei</i> as Scolytinae spp. (non-European)	_	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No
43	Procryphalus mucronatus as Scolytinae spp. (non-European)	-	Insects	No	Populus spp., Populus tremuloides (Atkinson, 2024)	Not assessed	No
44	Pycnarthrum hispidum as Scolytinae spp. (non-European)	_	Insects	No	Populus sp. (Gomez et al., 2020)	Not assessed	No
45	<i>Scolytoplatypus tycoon</i> as Scolytinae spp. (non-European)	-	Insects	No	Populus (Wood & Bright, 1992)	Not assessed	No

TABLE 5 (Continued)

No.	Pest name according to EU legislation ^a	EPPO code	Group	Pest present in the UK	Populus confirmed as a host (reference)	Pest can be associated with the commodity	Pest relevant for the Opinion
46	Trypodendron retusum as Scolytinae spp. (non-European)	-	Insects	No	Populus deltoides, P. grandidentata, P. tremuloides (Wood & Bright, 1992)	Not assessed	No
47	Trypophloeus klimeschi as Scolytinae spp. (non-European)	TRYOKL	Insects	No	Populus diversifolia (Wood & Bright, 1992)	Not assessed	No
48	Trypophloeus kurenzovi as Scolytinae spp. (non-European)	-	Insects	No	Populus tremula (Wood & Bright, 1992)	Not assessed	No
49	Trypophloeus populi as Scolytinae spp. (non-European)	CRYHPO	Insects	No	Populus angustifolia, P. tremuloides, P. tristis (Wood & Bright, 1992)	Not assessed	No
50	Trypophloeus thatcheri as Scolytinae spp. (non-European)	-	Insects	No	Populus tremuloides, P. tristis (Wood & Bright, 1992)	Not assessed	No
51	Trypophloeus tremulae as Scolytinae spp. (non-European)	-	Insects	No	Populus alba, P. nigra, P. tremula (DAFNAE, 2024)	Not assessed	No
52	<i>Xyleborus affinis</i> as Scolytinae spp. (non-European)	XYLBAF	Insects	No	Populus deltoides (Atkinson, 2024)	Not assessed	No
53	<i>Xyleborus perforans</i> as Scolytinae spp. (non-European)	XYLBPE	Insects	No	Populus deltoides (DAFNAE, 2024)	Not assessed	No
54	<i>Xyleborus volvulus</i> as Scolytinae spp. (non-European)	XYLBTO	Insects	No	Populus tremula (DAFNAE, 2024)	Not assessed	No
55	<i>Xyloterinus politus</i> as Scolytinae spp. (non-European)	XYORPO	Insects	No	Populus sp. (Mayers et al., 2020)	Not assessed	No

^aCommission Implementing Regulation (EU) 2019/2072.

^b *B. tabaci* (European population) is regulated as a protected zone quarantine pest. Therefore *B. tabaci* is listed twice, as European and non-European population. The association with *P. nigra* was assessed at the species level and not at the population level.

^c*Populus deltoides* was reported to be a putative host of the pathogen: However, host status was not confirmed by the fulfilment of the Koch's postulates (Vettraino et al., 2010). Therefore, there is no indication that populars are hosts of the pathogen.

4.2 | Selection of other relevant pests (non-regulated in the EU) associated with the commodity

The information provided by the UK, integrated with the search performed by EFSA, was evaluated in order to assess whether there are other relevant pests potentially associated with the commodity species present in the country of export. For these potential pests that are non-regulated in the EU, pest risk assessment information on the probability of entry, establishment, spread and impact is usually lacking. Therefore, these pests were also evaluated to determine their relevance for this Opinion based on evidence that:

- a. the pest is present in the UK;
- b. the pest is (i) absent or (ii) has a limited distribution in the EU;
- c. commodity is a host of the pest;
- d. one or more life stages of the pest can be associated with the specified commodity;
- e. the pest may have an impact in the EU.

For non-regulated species with a limited distribution (i.e. present in one or a few EU MSs) and fulfilling the other criteria (i.e. c, d and e), either one of the following conditions should be additionally fulfilled for the pest to be further evaluated:

- official phytosanitary measures have been adopted in at least one EU MS;
- any other reason justified by the working group (e.g. recent evidence of presence).

Pests that fulfilled the above listed criteria were selected for further evaluation.

Based on the information collected, 1602 potential pests known to be associated with the species commodity were evaluated for their relevance to this Opinion. Pests were excluded from further evaluation when at least one of the conditions listed above (1–5) was not met. Details can be found in Appendix F (Microsoft Excel® file). None of the pests not regulated in the EU was selected for further evaluation because none of them met all selection criteria.

There were two pests, i.e. *Colletotrichum populi* and *Pemphigus populitransversus* that were initially considered potentially relevant, but they were discarded at later stage because of the following reasons.

Colletotrichum populi

Colletotrichum populi was reported on *P. nigra* var. *italica* in China (Li et al., 2012). There was also one report from the UK on *Fragaria* × *ananassa* as *Colletotrichum aenigma* (Baroncelli et al., 2015), which according to USDA Fungal Database is the current fungus name (Farr & Rossman, 2024). However, according to the Index Fungorum (2024) and MyCoBank (2024) *C. aenigma* and *C. populi* are two separate species. The Panel, decided to follow the Index Fungorum and MyCoBank and therefore, the pest is not considered to be present in the UK.

Pemphigus populitransversus

The pest meets the criteria of being considered for further evaluation because *P. populitransversus* is reported to be associated with *P. nigra* in South Africa and *Populus* sp. in the Azores (Aphids on World's Plants, 2024), besides being impactful on *Brassica* in North America (Wene & White, 1953; Sokal et al., 1991; Coyle et al., 2005). However, there is no report on the pest producing galls on poplars in the UK (Dossier Section 2.0) suggesting a lack of association with the commodities. Available information suggests that the population in the UK is anholocyclic and only infests the secondary host (Brassicaceae) (Aphids on World's plants, 2024; Blackman & Eastop, 2006). Because of the high uncertainties and the lack of information a quantitative assessment is not conducted. A pest categorisation is also likely to be unconclusive based on the currently available information. The Panel proposes to include the pest in the horizon scanning programme.

4.3 **Overview of interceptions**

Data on the interception of harmful organisms on plants of *Populus* can provide information on some of the organisms that can be present on *Populus* despite the current measures taken. According to EUROPHYT (2024) (accessed on 10 December 2024) and TRACES-NT (2024) (accessed on 10 December 2024), there were no interceptions of plants for planting of *Populus* from the UK destined to the EU Member States due to the presence of harmful organisms between the years 1995 and 30 November 2024. It should be noted that since Brexit the movement of *Populus* from UK to the EU has been banned according to the current plant health legislation and therefore it is not expected to have interceptions after Brexit.

4.4 | List of potential pests not further assessed

From the list of pests not selected for further evaluation, the Panel highlighted two species (see Appendix E) for which currently available evidence provides no reason to select these species for further evaluation in this Opinion. A specific justification of the inclusion in this list is provided for each species in Appendix E.

4.5 | Summary of pests selected for further evaluation

The two pests satisfying all the relevant criteria listed above in the Sections 4.1 and 4.2 are included in Table 6. The effectiveness of the risk mitigation measures applied to the commodity was evaluated for these selected pests.

Number	Current scientific name	EPPO code	Name used in the EU legislation	Taxonomic information	Group	Regulatory status
1	Bemisia tabaci	BEMITA	<i>Bemisia tabaci</i> Genn. (European populations)	Hemiptera Aleyrodidae	Insects	Protected Zone Quarantine Pest according to Commission Implementing Regulation (EU) 2019/2072
2	Entoleuca mammata	НҮРОМА	Entoleuca mammata (Wahlenb.) Rogers and Ju	Xylariales Xylariaceae	Fungi	Protected Zone Quarantine Pest according to Commission Implementing Regulation (EU) 2019/2072

TABLE 6 List of relevant pests selected for further evaluation.

5 | RISK MITIGATION MEASURES

For the selected pests (Table 7), the Panel evaluated the likelihood that it could be present in the *P alba*, *P. nigra* and *P. trem-ula* nurseries by evaluating the possibility that the commodity in the export nurseries is infested either by:

- introduction of the pest from the environment surrounding the nursery;
- introduction of the pest with new plants/seeds;
- spread of the pest within the nursery.

The information used in the evaluation of the effectiveness of the risk mitigation measures is summarised in pest data sheets (see Appendix A).

5.1 | Risk mitigation measures applied in the UK

With the information provided by the UK (Dossier Sections 1.1, 1.2, 1.3, 2.0, 3.1, 3.2, 3.3, 4.1, 4.2, 4.3, 5.1, 5.2 and 5.3), the Panel summarised the risk mitigation measures (see Table 7) that are implemented in the production nursery.

Number	Risk mitigation measure	Implementation in the UK
1	Registration of production sites	All producers are registered as professional operators with the UK Competent Authority via APHA for England and Wales, or SASA for Scotland, and are authorised to issue the UK plant passports, verifying they meet the required national sanitary standards (Dossier Sections 1.1 1.2 and 1.3).
2	Physical separation	Most of the nurseries also produce plants for the local market, and there is no distancing between production areas for the export and the local market. All plants within UK nurseries are grown under the same phytosanitary measures, meeting the requirements of the UK Plant Passporting regime (Dossier Sections 1.1, 1.2 and 1.3).
3	Certified plant material	Populus seeds purchased in the UK are certified under The Forest Reproductive Material (Great Britain) Regulations 2002 (legislation.gov.uk); seedlings sourced in the UK are certified with UK Plant Passports. A small percentage of seed and young plants may be obtained from EU (Netherlands, Belgium and France); seeds and planting material from the EU countries are certified with phytosanitary certificates (Dossier Sections 1.1, 1.2 and 1.3).

TABLE 7 Overview of implemented risk mitigation measures for *Populus alba, Populus nigra* and *Populus tremula* plants designated for export to the EU from the UK.

TABLE 7	(Continued)	
Number	Risk mitigation measure	Implementation in the UK
4	Growing media	The growing media is virgin peat or peat-free compost. This compost is heat-treated by commercial suppliers during production to eliminate pests and diseases. It is supplied in sealed bulk bags or shrink-wrapped bales and stored off the ground on pallets, these are free from contamination. Where delivered in bulk, compost is kept in a dedicated bunker, either indoors, or covered by tarpaulin outdoors, and with no risk of contamination with soil or other material (Dossier Sections 1.1, 1.2 and 1.3).
5	Surveillance, monitoring and sampling	For additional information see Section 3.3.3 Pest monitoring during production.
6	Hygiene measures	 All nurseries have plant hygiene and housekeeping rules and practices in place, which are communicated to all relevant employees. General hygiene measures are undertaken as part of routine nursery production, including disinfection of tools and equipment between batches/lots and different plant species. The tools are dipped in a disinfectant solution and wiped with a clean cloth between trees to reduce the risk of transfer of pests between subjects. There are various disinfectants available, with Virkon S (active substance: potassium peroxymonosulfate and sodium chloride) being a common example. Growers must have an appropriate programme of weed management in place on the nursery (Dossier Sections 1.1, 1.2 and 1.3).
7	Removal of infested plant material	 Post-harvest and through the autumn and winter, nursery management is centred on pest and disease prevention and maintaining good levels of nursery hygiene. Leaves, pruning residues and weeds are all removed from the nursery to reduce the number of over wintering sites for pests and diseases. All residues or waste materials shall be assessed for the potential to host, harbour or transmit pests (Dossier Sections 1.1, 1.2 and 1.3).
8	Irrigation water	Water for irrigation is routinely sampled and sent for analysis (Dossier Sections 1.1, 1.2 and 1.3).
9	Application of pest control products	 Crop protection is achieved using a combination of measures including approved plant protection products, biological control or physical measures. Plant protection products are only used when necessary and records of all plant protection treatments are kept. Pest and disease pressure varies from season to season. Plant protection products are applied application takes place only when required and depends on situation (disease pressure, growth stage etc. and environmental factors) at that time. Subject to this variation in pest pressure, in some seasons few, if any, pesticides are applied; in others it is sometimes necessary to apply preventative and/or control applications of pesticides. In many circumstances also, biological control is reported to be used to control outbreaks, rather than using chemical treatments. Examples of typical products used against rusts, leafspots and canker fungi, spider mites, aphids and weeds are provided in the Dossier Sections 1.1, 1.2, 1.3 and 5.1. These would be applied at the manufacturers recommended rate and intervals (Dossier Sections 1.1, 1.2, and 1.3).
10	Measures against soil pests	There are no specific measures/treatments against soil pests. However, containerised plants are grown in trays on top of protective plastic membranes to prevent contact with soil. Membranes are regularly refreshed when needed. Alternatively, plants may be grown on raised galvanised steel benches stood on gravel as a barrier between the soil and bench feet and/or concreted surfaces (Dossier Sections 1.1, 1.2 and 1.3).
11	Inspections and management of plants before export	 The UK NPPO carries out inspections and testing where required by the country of destination's plant health legislation, to ensure all requirements are fulfilled and a valid phytosanitary certificate with the correct additional declarations is issued (Dossier Sections 1.1, 1.2 and 1.3). Separate to any official inspection, plant material is checked by growers for plant health issues prior to dispatch (Dossier Sections 1.1, 1.2 and 1.3). A final pre-export inspection is undertaken as part of the process of issuing a phytosanitary certificate. These inspections are generally undertaken usually within 1–2 days, and not more than 2 weeks before export. Phytosanitary certificates are only issued if the commodity meets the required plant health standards after inspection and/or testing according to appropriate official procedures (Dossier Sections 1.1, 1.2 and 1.3). The protocol for plants infested by pests during inspections before export is to treat the plants, if they are on site for a sufficient period of time or to destroy any plants infested by pests otherwise. All other host plants in the nursery would be treated. The phytosanitary certificate for export will not be issued until the UK Plant Health inspectors confirm that the plants are free from pests (Dossier Sections 1.1, 1.2 and 1.3).

TABLE 7 (Continued)

Number Risk mitigation measure Implementation in the UK	
12 Separation during transport to the destination The commodities are dispatched as single plant plants or in bundles as follows: 12 destination - 10-20 for cuttings/graftwood; -5, 10, 15, 25, 50 for bare root plants; - 5-10 for cell grown plants. Cuttings/graftwood is wrapped in plastic and p 15 certified wooden pallets, or metal pallets and 1.3). Bare root plants are then wrapped in polythene wooden pallets, or metal pallets. Alternative wrapped in polythene bags and dispatched via (Dossier Rooted plants in pots are transported on Danis pallets, or individually in pots for larger con The preparation of the commodities for export environment, e.g. packing shed, except for an open field due to their dimensions (Doss Plants are transported by lorry (size dependant transported by temperature-controlled lorr	nts in pots, single cell grown plants, single bare root packed in cardboard boxes or Dutch crates on ISPM ts, dependant on quantity (Dossier Sections 1.1, 1.2 ne and packed and distributed on ISPM 15 certified vely, they may be placed in pallets which are then rs may be packed in waxed cardboard cartons or er Sections 1.1, 1.2 and 1.3). ish trolleys for smaller containers, or ISPM 15 certified ntainers (Dossier Sections 1.1, 1.2 and 1.3). rt is carried out inside the nurseries in a closed r the specimen trees, which are prepared outside in ssier Sections 1.1, 1.2 and 1.3). it on load quantity). Sensitive plants are occasionally rry if weather conditions during transit are likely to be

5.2 | Evaluation of the current measures for the selected relevant pests including uncertainties

For each evaluated pest, the relevant risk mitigation measures acting on the pest were identified. Any limiting factors on the effectiveness of the measures were documented.

All the relevant information including the related uncertainties deriving from the limiting factors used in the evaluation are summarised in a pest data sheet provided in Appendix A. Based on this information, for each selected relevant pest, an expert judgement is given for the likelihood of pest freedom taking into consideration the risk mitigation measures and their combination acting on the pest.

An overview of the evaluation of each relevant pest is given in the sections below (Sections 5.2.1–5.2.2). The outcome of the EKE regarding pest freedom after the evaluation of the currently proposed risk mitigation measures is summarised in Section 5.2.3.

Overview of the evaluatio	Overview of the evaluation of <i>B. tabaci</i> for bare root plants of 1–7 years						
Rating of the likelihood of pest freedom	Pest free with some exe	ceptional cases (based o	n the median).				
Percentile of the distribution	5%	25%	Median	75%	95%		
Proportion of pest-free plants/bundles	9959 out of 10,000 plants/bundles	9976 out of 10,000 plants/bundles	9987 out of 10,000 plants/bundles	9994 out of 10,000 plants/bundles	9999 out of 10,000 plants/bundles		
Percentile of the distribution	5%	25%	Median	75%	95%		
Proportion of infected plants/bundles	1 out of 10,000 plants/bundles	6 out of 10,000 plants/bundles	13 out of 10,000 plants/bundles	25 out of 10,000 plants/bundles	41 out of 10,000 plants/bundles		
Summary of the information used for the evaluation	 Possibility that the pest could become associated with the commodity The pest is present in the UK, with few occurrences but continuously intercepted. The UK outbreaks of <i>B. tabaci</i> have been restricted to greenhouses. The pest is extremely polyphagous with a very wide host range. Other traded plants present in the surroundings of the nursery could be a source of the pest. Polytunnels and glasshouses in the nurseries could act as a reservoir of the pest. The pest could go undetected during inspections. Measures taken against the pest and their efficacy General measures taken by the nurseries are effective against the pest. These measures include (a) inspections, surveillance, monitoring, sampling and laboratory testing; (b) hygiene measures; (c) application of pest control products and (d) removal of infested plant material. 						

5.2.1 | Overview of the evaluation of *Bemisia tabaci* (European populations) (Hemiptera; Aleyrodidae)

\sim						1
(വ	nti	n	116	ചറ	11
\sim				u		·/

Interception records

In the EUROPHYT/TRACES-NT database there are no records of notification of *Populus* plants for planting neither from the UK or from other countries due to the presence of *B. tabaci* between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024).

There were four interceptions of *B. tabaci* from the UK in 2007 and 2015 on non *Populus* plants, (EUROPHYT, 2024). **Shortcomings of current measures/procedures**

None.

Main uncertainties

- Possibility of development of the pest outside greenhouses.
- Pest abundance in the nursery and the surroundings.
- The precision of surveillance and the application of measures targeting the pest.
- Host status of Populus spp. (other than P. nigra) to the pest.

Overview of the evaluation of B. tabaci for cell grown plants of 1-2 years

Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the median).				
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of pest-free plants/bundles	9943 out of 10,000 bundles	9966 out of 10,000 bundles	9981 out of 10,000 bundles	9992 out of 10,000 bundles	9998 out of 10,000 bundles
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	2 out of 10,000 bundles	8 out of 10,000 bundles	19 out of 10,000 bundles	34 out of 10,000 bundles	57 out of 10,000 bundles
Summary of the information used for the evaluation	 Possibility that the pest could become associated with the commodity The pest is present in the UK, with few occurrences but continuously intercepted. The UK outbreaks of <i>B. tabaci</i> has been restricted to greenhouses. The pest is extremely polyphagous with a very wide host range. Other traded plants present in the surroundings of the nursery could be a source of the pest. Polytunnels and glasshouses in the nurseries could act as a reservoir of the pest. The pest could go undetected during inspections. Measures taken against the pest and their efficacy General measures taken by the nurseries are effective against the pest. These measures include (a) inspections, surveillance, monitoring, sampling and laboratory testing; (b) hygiene measures; (c) application of pest control products and (d) removal of infested plant material. Interception records In the EUROPHYT/TRACES-NT database there are no records of notification of <i>Populus</i> plants for planting neither from the UK or from other countries due to the presence of <i>B. tabaci</i> between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024). There were four interceptions of <i>B. tabaci</i> from the UK in 2007 and 2015 on non <i>Populus</i> plants, (EUROPHYT, 2024). Shortcomings of current measures/procedures None. Main uncertainties Possibility of development of the pest outside greenhouses. Pest abundance in the nursery and the surroundings. The precision of surveillance and the application of measures targeting the pest. 				eaks of <i>B. tabaci</i> have ange. Other traded and glasshouses in ections. e (a) inspections, tion of pest control r planting neither 5 and September (EUROPHYT, 2024).

Overview of the evaluation of B. tabaci for plants in pots of 3–15 years

Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the median).					
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of pest-free plants/bundles	9937 out of 10,000 plants	9961 out of 10,000 plants	9978 out of 10,000 plants	9991 out of 10,000 plants	9999 out of 10,000 plants	
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of infected plants/bundles	1 out of 10,000 plants	9 out of 10,000 plants	22 out of 10,000 plants	39 out of 10,000 plants	63 out of 10,000 plants	
Summary of the information used for the evaluation	Possibility that the pest could become associated with the commodity The pest is present in the UK, with few occurrences but continuously intercepted. The UK outbreaks of <i>B. tabaci</i> have been restricted to greenhouses. The pest is extremely polyphagous with a very wide host range. Other traded plants present in the surroundings of the nursery could be a source of the pest. Polytunnels and glasshouses in the nurseries could act as a reservoir of the pest. The pest could go undetected during inspections.					

(Continued)	
	Measures taken against the pest and their efficacy General measures taken by the nurseries are effective against the pest. These measures include (a) inspections, surveillance, monitoring, sampling and laboratory testing; (b) hygiene measures; (c) application of pest control products and (d) removal of infested plant material.
	Interception records
	In the EUROPHYT/TRACES-NT database there are no records of notification of <i>Populus</i> plants for planting neither from the UK or from other countries due to the presence of <i>B. tabaci</i> between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024).
	There were four interceptions of <i>B. tabaci</i> from the UK in 2007 and 2015 on non <i>Populus</i> plants, (EUROPHYT, 2024).
	Shortcomings of current measures/procedures
	None.
	Main uncertainties
	 Possibility of development of the pest outside greenhouses.
	 Pest abundance in the nursery and the surroundings.
	 The precision of surveillance and the application of measures targeting the pest.
	- Host status of Populus spp. to the pest Host status of Populus spp. (other than P. nigra) to the pest.

For more details, see relevant pest data sheet on Bemisia tabaci (European populations) (Section A.1 in Appendix A).

5.2.2 | Overview of the evaluation of *Entoleuca mammata* (Xylariales; Xylariaceae)

Overview of the evaluation	n of <i>E. mammata</i> for cu	ttings/graftwood P. nig	gra of 1–2 years			
Rating of the likelihood of pest freedom	Pest free with some ex	ceptional cases (based o	on the median).			
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of pest-free plants/bundles	9947 out of 10,000 plants/bundles	9971 out of 10,000 plants/bundles	9983 out of 10,000 plants/bundles	9992 out of 10,000 plants/bundles	9998 out of 10,000 plants/bundles	
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of infected plants/bundles	2 out of 10,000 plants/bundles	8 out of 10,000 plants/bundles	17 out of 10,000 plants/bundles	29 out of 10,000 plants/bundles	63 out of 10,000 plants/bundles	
Overview of the evaluation	n of <i>E. mammata</i> for cuttings/graftwood <i>P. tremula</i>					
Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the Median).					
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of pest-free plants/bundles	9894 out of 10,000 plants/bundles	9941 out of 10,000 plants/bundles	9967 out of 10,000 plants/bundles	9985 out of 10,000 plants/bundles	9997 out of 10,000 plants/bundles	
Percentile of the distribution	5%	25%	Median	75%	95%	
Proportion of infected plants/bundles	3 out of 10,000 plants/bundles	15 out of 10,000 plants/bundles	33 out of 10,000 plants/bundles	59 out of 10,000 plants/bundles	106 out of 10,000 plants/bundles	
Summary of the information used for the evaluation	 Possibility that the pest could become associated with the commodity Entoleuca mammata is present in the UK, although not widely distributed. All poplars (Populus spp.) are suitable hosts, although <i>P. tremula</i> is a major one and <i>P. nigra</i> and <i>P. alba</i> may be minor hosts. Mechanical wounds including pruning wounds are expected to be present and may represent infection courts. The hosts can be present either inside or in the surroundings of the nurseries. Altogether, this suggests that an association with the commodity is possible. Measures taken against the pest and their efficacy General measures taken by the nurseries have an effect against the pathogen. These measures include (a) the use of certified plant material; (b) inspections, surveillance, monitoring, sampling and laboratory testing; (c) the removal of infected plant material and (d) application of pest control products. Interception records In the EUROPHYT/TRACES-NT database there are no records of notification of <i>Populus</i> plants for planting neither from the UK nor from other countries due to the presence of <i>E. mammata</i> between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024). Shortcomings of current measures/procedures None observed. Main uncertainties The presence/abundance of the pathogen in the area where the nurseries are located. Effect of fungicide treatments against the pathogen. 					

Overview of the evaluation of <i>E. mammata</i> for bare root plants <i>P. alba, P. nigra</i> of 1–7 years							
Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the median).						
Percentile of the distribution	5%	25%	Median	75%	95%		
Proportion of pest-free plants/bundles	9911 out of 10,000 plants/bundles	9950 out of 10,000 plants/bundles	9971 out of 10,000 plants/bundles	9985 out of 10,000 plants/bundles	9996 out of 10,000 plants/bundles		
Percentile of the distribution	5%	25%	Median	75%	95%		
Proportion of infected plants/bundles	4 out of 10,000 plants/bundles	15 out of 10,000 plants/bundles	29 out of 10,000 plants/bundles	50 out of 10,000 plants/bundles	89 out of 10,000 plants/bundles		
Overview of the evaluation of Entoleuca mammata for bare root plants P. tremula							
Rating of the likelihood of pest freedom	Extremely frequently pest free (based on the Median).						
Percentile of the distribution	5%	25%	Median	75%	95%		

alstribution					
Proportion of pest-free plants/bundles	9822 out of 10,000 plants/bundles	9900 out of 10,000 plants/bundles	9942 out of 10,000 plants/bundles	9970 out of 10,000 plants/bundles	9992 out of 10,000 plants/bundles
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	8 out of 10,000 plants/bundles	30 out of 10,000 plants/bundles	58 out of 10,000 plants/bundles	100 out of 10,000 plants/bundles	178 out of 10,000 plants/bundles
Summary of the information used	Possibility that the pest could become associated with the commodity <i>Entoleuca mammata</i> is present in the UK, although not widely distributed. All poplars (<i>Populus</i> spp.) are suitable				

Entoleuca mammata is present in the UK, although not widely distributed. All poplars (*Populus* spp.) are suitable hosts, although *Populus tremula* is a major one and *P. nigra* and *P. alba* may be minor hosts. The hosts can be present either inside or in the surroundings of the nurseries. Infection may occur through mechanically-induced wounds such as pruning wounds. Altogether, this suggests that an association with the commodity is possible. **Measures taken against the pest and their efficacy**

General measures taken by the nurseries are effective against the pathogen. These measures include (a) the use of certified plant material; (b) inspections, surveillance, monitoring, sampling and laboratory testing; (c) the removal of infected plant material and (d) application of pest control products.

Interception records

for the evaluation

In the EUROPHYT/TRACES-NT database there are no records of notification of *Populus* plants for planting neither from the UK nor from other countries due to the presence of *E. mammata* between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024).

Shortcomings of current measures/procedures

None observed.

Main uncertainties

- The presence/abundance of the pathogen in the area where the nurseries are located.

- Whether the pest can reliably be detected via visual inspection.

- Effect of fungicide treatments against the pathogen.

Overview of the evaluation of E. mammata for cell grown plants P. alba, P. nigra of 1-2 years

Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the median).				
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of pest-free plants/bundles	9930 out of 10,000 bundles	9961 out of 10,000 bundles	9979 out of 10,000 bundles	9991 out of 10,000 bundles	9998 out of 10,000 bundles
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	2 out of 10,000 bundles	9 out of 10,000 bundles	21 out of 10,000 bundles	39 out of 10,000 bundles	70 out of 10,000 bundles
Overview of the evaluation of <i>E. mammata</i> for cell grown plants <i>P. tremula</i>					
Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the Median).				
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of pest-free plants/bundles	9861 out of 10,000 bundles	9922 out of 10,000 bundles	9958 out of 10,000 bundles	9981 out of 10,000 bundles	9997 out of 10,000 bundles

COMMODITY RISK ASSESSMENT OF POPULUS PLANTS FROM THE UK

(Continued)					
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	3 out of 10,000 bundles	19 out of 10,000 bundles	42 out of 10,000 bundles	78 out of 10,000 bundles	139 out of 10,000 bundles
Summary of the information used for the evaluation	Possibility that the p Entoleuca mammata is hosts, although Po plants are in close conditions for E. m hosts can be prese association with th Measures taken agai General measures take of certified plant m removal of infected Interception records In the EUROPHYT/TRA from the UK nor fro September 2024 (E Shortcomings of curr None observed. Main uncertainties – The presence/abun – Whether the pest ca – Effect of fungicide t	est could become asso present in the UK, alth- pulus tremula is a major proximity to each other ammata. Mechanical w ant either inside or in the ecommodity may be p nst the pest and their en by the nurseries are of naterial; (b) inspections, d plant material and (d) CES-NT database there on other countries due CUROPHYT, 2024; TRACE rent measures/proced dance of the pathogen an reliably be detected treatments against the	boiated with the common bugh not widely distrib one and <i>P. nigra</i> and <i>P.</i> which increases the hu ounds could be present e surroundings of the ni ossible. efficacy effective against the pa surveillance, monitorir application of pest con are no records of notifit to the presence of <i>E. ma</i> (S-NT, 2024). ures in the area where the ni via visual inspection. pathogen.	nodity uted. All poplars (<i>Popul</i> <i>alba</i> may be minor hos imidity and hence prov t and may represent info urseries. Altogether, thi thogen. These measure ig, sampling and labora itrol products. cation of <i>Populus</i> plants <i>ammata</i> between the your nurseries are located.	us spp.) are suitable ts. Cell grown ides good growth action courts. The s suggests that an s include (a) the use tory testing; (c) the for planting neither ears 1995 and

Overview of the evaluation of <i>E. mammata</i> for plants in pots (<i>Populus alba, P. nigra</i>) of 3–15 years	

Rating of the likelihood of pest freedom	Pest free with some exceptional cases (based on the median).				
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of pest-free plants/bundles	9865 out of 10,000 plants	9923 out of 10,000 plants	9958 out of 10,000 plants	9982 out of 10,000 plants	9997 out of 10,000 plants
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	3 out of 10,000 plants	18 out of 10,000 plants	42 out of 10,000 plants	77 out of 10,000 plants	135 out of 10,000 plants
Overview of the evaluation of E	. <i>mammata</i> for plants	in pots (P. tremula)			
Rating of the likelihood of pest freedom	Very frequently pest f	ree (based on the Medi	an).		
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of pest-free plants/bundles	9730 out of 10,000 plants	9845 out of 10,000 plants	9916 out of 10,000 plants	9964 out of 10,000 plants	9994 out of 10,000 plants
Percentile of the distribution	5%	25%	Median	75%	95%
Proportion of infected plants/bundles	6 out of 10,000 plants	36 out of 10,000 plants	84 out of 10,000 plants	155 out of 10,000 plants	270 out of 10,000 plants
Summary of the information used for the evaluation	plantsplantsplantsplantsplantsPossibility that the pest could become associated with the commodityEntoleuca mammata is present in the UK, although not widely distributed. All poplars (Populus spp.) are suitabhosts, although P. tremula is a major one and P. nigra and P. alba may be minor hosts. Mechanical woundsincluding pruning wounds are expected to be present and may represent infection courts. The hosts can bepresent either inside or in the surroundings of the nurseries. Altogether, this suggests that an associationwith the commodity may be possible.Measures taken against the pest and their efficacyGeneral measures taken by the nurseries are effective against the pathogen. These measures include (a) the us of certified plant material; (b) inspections, surveillance, monitoring, sampling and laboratory testing; (c) th removal of infected plant material and (d) application of pest control products.Interception recordsIn the EUROPHYT/TRACES-NT database there are no records of notification of Populus plants for planting neither from the UK nor from other countries due to the presence of <i>E. mammata</i> between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024).Shortcomings of current measures/proceduresNone observed.Main uncertainties				lus spp.) are suitable echanical wounds rts. The hosts can be nat an association es include (a) the use atory testing; (c) the s for planting een the years 1995

For more details, see relevant pest data sheet on Entoleuca mammata (Section A.2 in Appendix A).

5.2.3 | Outcome of Expert Knowledge Elicitation

Table 8 and Figure 2 show the outcome of the EKE regarding pest freedom after the evaluation of the implemented risk mitigation measures for all the evaluated pests.

Figure 3 provides an explanation of the descending distribution function describing the likelihood of pest freedom after the evaluation of the implemented risk mitigation measures for *P. tremula* plants in pots up to 15 years old designated for export to the EU for *E. mammata*.

TABLE 8 Assessment of the likelihood of pest freedom following evaluation of current risk mitigation measures against pests on *Populus alba, P. nigra* and *P. tremula* plants designated for export to the EU. In panel A, the median value for the assessed level of pest freedom for each pest is indicated by 'M', the 5% percentile is indicated by 'L' and the 95% percentile is indicated by 'U'. The percentiles together span the 90% uncertainty range regarding pest freedom. The pest freedom categories are defined in panel B of the table.

Number	Group	Pest species	Sometimes pest free	More often than not pest free	Frequently pest free	Very frequently pest free	Extremely frequently pest free	Pest free with some exceptional cases	Pest free with few exceptional cases	Almost always pest free
1	Insects	Bemisia tabaci, P. alba, P.nigra, P. tremula, bare root plants						LM		U
2	Insects	Bemisia tabaci, P. alba, P.nigra, P. tremula, cell grown plants					L	м		U
3	Insects	Bemisia tabaci, P. alba, P.nigra, P. tremula, plants in pots					L	м		U
4	Fungi	Entoleuca mammata, P. nigra, cuttings/graftwood					L	м		U
5	Fungi	Entoleuca mammata, P. alba, P. nigra, bare root plants					L	м		U
6	Fungi	Entoleuca mammata, P. alba, P. nigra, cell grown plants					L	м		U
7	Fungi	Entoleuca mammata, P alba, P. nigra, plants in pots				L		м		U
8	Fungi	Entoleuca mammata, P. tremula, cuttings/graftwood				L		м		U
9	Fungi	<i>Entoleuca mammata, P tremula,</i> bare root plants				L	м		U	
10	Fungi	Entoleuca mammata, P tremula, cell grown plants				L		м		U
11	Fungi	Entoleuca mammata, P tremula, plants in pots				L	м		U	

PANEL A

Pest freedom category	Pest fee plants out of 10,000
Sometimes pest free	≤5000
More often than not pest free	5000-≤9000
Frequently pest free	9000-≤9500
Very frequently pest free	9500-≤9900
Extremely frequently pest free	9900 -≤ 9950
Pest free with some exceptional cases	9950-≤9990
Pest free with few exceptional cases	9990-≤9995
Almost always pest free	9995-≤10,000

PANEL B

Legend of pest freedom categories			
L	Pest freedom category includes the elicited lower bound of the 90% uncertainty range		
М	Pest freedom category includes the elicited median		
U	Pest freedom category includes the elicited upper bound of the 90% uncertainty range		



Uncertainty distributions of pest freedom for different pests

Categories of pest freedom

[pestfree plants out of 10,000] (logarithmic scale: - LOG(1-PF))

FIGURE 2 Elicited certainty (y-axis) of the number of pest-free plants/bundles of *Populus alba*, *Populus nigra* and *Populus tremula* (x-axis; log-scaled) out of 10,000 plants/bundles designated for export to the EU from the UK for all evaluated pests visualised as descending distribution function. Horizontal lines indicate the reported certainty levels (starting from the bottom 5%, 25%, 50%, 75%, 95%). Please see reading instructions below.



Uncertainty distributions of pest freedom of plants in pots Populus tremula for Entoleuca mammata

Categories of pest freedom

[pestfree plants out of 10,000] (logarithmic scale: - LOG(1-PF))

FIGURE 3 Explanation of the descending distribution function describing the likelihood of pest freedom after the evaluation of the implemented risk mitigation measures for plants designated for export to the EU based on based on the example of *Entoleuca mammata* on *Populus tremula* plants in pots of 3–15 years old.

6 | CONCLUSIONS

There are two pests identified to be present in the UK and considered to be potentially associated with the commodities imported from the UK and relevant for the EU.

These pests are *Bemisia tabaci* (European populations) and *Entoleuca mammata*. The likelihood of the pest freedom after the evaluation of the implemented risk mitigation measures for the commodities designated for export to the EU was estimated. In the assessment of risk, the age of the plants was considered, reasoning that older trees are more likely to be infested mainly due to longer exposure time and larger size.

For *B. tabaci* the likelihood of pest freedom for bare root plants/trees up to 7 years old of *P. alba, P. nigra* and *P. tremula* was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'pest free with some exceptional cases' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9959 and 10,000 bare root plants/trees up to 7 years old per 10,000 will be free from *B. tabaci*. The likelihood of pest freedom for cell grown plants of *P. alba, P. nigra* and *P. tremula* up to 2 years old was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9943 and 10,000 cell grown plants in pots up to 2 years old per 10,000 will be free from *B. tabaci*. The likelihood of pest freedom for rooted plants in pots of *P. alba, P. nigra* and *P. tremula* up to 2 years old was estimated as 'pest free'. The EKE indicated, with 95% certainty, that between 9943 and 10,000 cell grown plants in pots up to 2 years old per 10,000 will be free from *B. tabaci*. The likelihood of pest freedom for rooted plants in pots of *P. alba, P. nigra* and *P. tremula* from three to 15 years old was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9943 and 10,000 cell grown plants in pots of *P. alba, P. nigra* and *P. tremula* from three to 15 years old was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9937 and 10,000 rooted plants in pots from three to 15 years old per 10,000 will be free from *B. tabaci*.

For *E. mammata* the likelihood of pest freedom for cuttings/graftwood of *of P. nigra*, following evaluation of current risk mitigation measures, was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9947 and 10,000 cuttings/graftwood per 10,000 will be free from *E. mammata*. The likelihood of pest freedom for bare root plants/trees up to 7 years old of *P. alba* and *P. nigra* was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9911 and 10,000 bare root plants/trees up to 7 years old per 10,000 will be free from *E. mammata*. The likelihood of pest freedom for cell grown plants of *P. alba* and *P. nigra* up to 2 years old was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently range reaching from 'extremely free with some exceptional cases' with the 90% uncertainty range reaching from 'extremely frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9930 and 10,000 cell grown plants in pots up to 2 years old per 10,000 will be free from *E. mammata*. The likelihood of pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9930 and 10,000 cell grown plants in pots of *P. alba* and *P. nigra* from three to 15 years old was estimated as 'pest free'. The EKE indicated, with 95% certainty range spanning from 'very frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that

For *E. mammata* the likelihood of pest freedom for cuttings/graftwood of *P. tremula*, following evaluation of current risk mitigation measures, was estimated as 'pest free with some exceptional cases' with the 90% uncertainty range spanning from 'very frequently pest free' to 'almost always pest free'. The EKE indicated, with 95% certainty, that between 9894 and 10,000 cuttings/graftwood per 10,000 will be free from *E. mammata*. The likelihood of pest freedom for bare root plants/ trees up to 7 years old of *P. tremula* was estimated as 'extremely frequently pest free' with the 90% uncertainty range reaching from 'very frequently pest free' to 'pest free with few exceptional cases'. The EKE indicated, with 95% certainty, that between 9822 and 10,000 bare root plants/trees up to 7 years old per 10,000 will be free from *E. mammata*. The likelihood of pest free with some exceptional cases' with the 90% uncertainty range spanning from 'very frequently pest free' to 'pest free up to 7 years old per 10,000 will be free from *E. mammata*. The likelihood of pest freedom for cell grown plants of *P. tremula* up to 2 years old was estimated as 'pest free'. The EKE indicated, with 95% certainty, that between 9861 and 10,000 cell grown plants in pots up to 2 years old per 10,000 will be free from *E. mammata*. The likelihood of pest freedom for rooted plants in pots of *P. tremula* from three to 15 years old was estimated as 'extremely frequently pest free' with the 90% uncertainty range reaching from 'very frequently pest free' with few exceptional cases'. The EKE indicated, with 95% certainty, that between 9861 and 10,000 cell grown plants in pots of *P. tremula* from three to 15 years old was estimated as 'extremely frequently pest free' with the 90% uncertainty range reaching from 'very frequently pest free' to 'pest free with few exceptional cases'. The EKE indicated, with 95% certainty, that between 9861 and 10,000 cell grown plants in pots of *P. tremula* from three to 15 years old was estimated as 'extremely fr

GLOSSARY	
Control (of a pest)	Suppression, containment or eradication of a pest population (FAO, 2024a, 2024b).
Entry (of a pest)	Movement of a pest into an area where it is not yet present, or present but not widely distributed and being officially controlled (FAO, 2024b).
Establishment (of a pest)	Perpetuation, for the foreseeable future, of a pest within an area after entry (FAO, 2024b).
Impact (of a pest)	The impact of the pest on the crop output and quality and on the environment in the occupied spatial units.
Introduction (of a pest)	The entry of a pest resulting in its establishment (FAO, 2024b).
Measures	Control (of a pest) is defined in ISPM 5 (FAO, 2024b) as 'Suppression, containment or eradication of a pest population' (FAO, 2024a). Control measures are measures that have a direct effect on pest abundance. Supporting measures are organisational measures or procedures supporting the choice of appropriate risk mitigation measures that do not directly affect pest abundance.
Pathway	Any means that allows the entry or spread of a pest (FAO, 2024b).

Phytosanitary measures	Any legislation, regulation or official procedure having the purpose to prevent the in- troduction or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests (FAO, 2024b).
Protected zone	A Protected zone is an area recognised at EU level to be free from a harmful organism, which is established in one or more other parts of the Union.
Quarantine pest	A pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled (FAO, 2024b).
Regulated non-quarantine pest	A non-quarantine pest whose presence in plants for planting affects the intended use of those plants with an economically unacceptable impact and which is therefore regulated within the territory of the importing contracting party (FAO, 2024b).
Risk mitigation measure	A measure acting on pest introduction and/or pest spread and/or the magnitude of the biological impact of the pest should the pest be present. A risk mitigation measure may become a phytosanitary measure, action or procedure according to the decision of the risk manager.
Spread (of a pest)	Expansion of the geographical distribution of a pest within an area (FAO, 2024b).

ABBREVIATIONS

APHA	Animal and Plant Health Agency
CABI	Centre for Agriculture and Bioscience International
DEFRA	Department for Environment Food and Rural Affairs
EKE	Expert Knowledge Elicitation
EPPO	European and Mediterranean Plant Protection Organization
FAO	Food and Agriculture Organization
ISPM	International Standards for Phytosanitary Measures
NPPO	National Plant Protection Organisation
PHSI	Plant Health and Seeds Inspectorate
PLH	Plant Health
PRA	Pest Risk Assessment
RNQPs	Regulated Non-Quarantine Pests
SASA	Science and Advice for Scottish Agriculture

ACKNOWLEDGEMENTS

The Scientific Opinion was prepared in cooperation with the Universita degli studi di Padova, Dipartimento Agronomia, Animali, Alimenti, Risorse Naturali e Ambiente (Italy) under the EFSA Art. 36 Framework Partnership Agreement 'GP/EFSA/ PLANTS/2022/11' commodity risk assessment for forestry plants.

REQUESTOR

European Commission

QUESTION NUMBERS

EFSA-Q-2023-00588, EFSA-2023-00589, EFSA-2023-00590.

COPYRIGHT FOR NON-EFSA CONTENT

EFSA may include images or other content for which it does not hold copyright. In such cases, EFSA indicates the copyright holder and users should seek permission to reproduce the content from the original source.

PANEL MEMBERS

Antonio Vicent Civera, Paula Baptista, Anna Berlin, Elisavet Chatzivassiliou, Jaime Cubero, Nik Cunniffe, Eduardo De La Peña, Nicolas Desneux, Francesco Di Serio, Anna Filipiak, Paolo Gonthier, Beata Hasiów-Jaroszewska, Hervé Jactel, Blanca B. Landa, Lara Maistrello, David Makowski, Panagiotis Milonas, Nikos Papadopoulos, Roel Potting, Hanna Susi and Dirk Jan van Der Gaag.

REFERENCES

Ahmad, M. I., Hafiz, I. A., & Chaudhry, M. I. (1977). Biological studies of *Aeolesthes sarta* Solksy attacking poplars in Pakistan. *Pakistan Journal of Forestry*, 27, 123–129.

Anonymous. (1960). Index of plant diseases in the United States. Agriculture Handbook no 165, USDA-ARS (US), 531 pp.

Aphids on World's Plants. (2024). An online identification and information guide. https://aphidsonworldsplants.info (accessed: 2024-10-13).

Atkinson, T. H. (2024). Bark and Ambrosia Beetles of the Americas. https://www.barkbeetles.info/index.php (accessed: 2024-12-10).

Baroncelli, R., Zapparata, A., Sarrocco, S., Sukno, S. A., Lane, C. R., Thon, M. R., Vannacci, G., Holub, E., & Sreenivasaprasad, S. (2015). Molecular diversity of anthracnose pathogen populations associated with UK strawberry production suggests multiple introductions of three different *Colletotrichum* species. *PLoS One*, *10*(6), e0129140. https://doi.org/10.1371/journal.pone.0129140

- Batsankalashvili, M., Kaydan, M. B., Kirkitadze, G., & Japoshvili, G. (2017). Updated checklist of scale insects (Hemiptera: Coccomorpha) in Sakartvelo (Georgia). Annals of Agrarian Science, 15(2), 252–268. https://doi.org/10.1016/j.aasci.2017.05.002
- Blackman, R. L., & Eastop, V. F. (2006). Aphids on the World's herbaceous plants and shrubs (Vol. 1 and 2, p. 1450). John Wiley and Sons.
- Brown, J. K., Paredes-Montero, J. R., & Stocks, I. C. (2023). The *Bemisia tabaci* cryptic (sibling) species group Imperative for a taxonomic reassessment, current opinion in insect. *Science*, *57*, 101032. https://doi.org/10.1016/j.cois.2023.101032
- Byun, B. K., & Yan, S. (2004). Check list of the tribe Tortricini (Lepidoptera: Tortricidae) in Northeast China, with two newly recorded species from China. *Korean Journal of Applied Entomology*, 43(2), 91–101.
- CABI (Centre for Agriculture and Bioscience International). (2020). Agrilus anxius (bronze birch borer). https://www.cabidigitallibrary.org/doi/full/10.1079/ cabicompendium.3771 (accessed: 2024-12-10).
- Casarin, N., Hasbroucq, S., Pesenti, L., Géradin, A., Emond, A., López-Mercadal, J., Miranda, M. Á., Grégoire, J. C., & Bragard, C. (2023). Salicaceae as potential host plants of *Xylella fastidiosa* in European temperate regions. *European Journal of Plant Pathology*, *165*(3), 489–507. https://doi.org/10.1007/ s10658-022-02622-7
- Clausen, C. P., King, J. L., & Teranishi, C. (1927). The parasites of *Popillia japonica* in Japan and chosen (Korea), and their introduction into the United States. US Department of Agriculture, department bulletin, 1429, 156 pp.
- Coyle, D. R., Nebeker, T. E., Hart, E. R., & Mattson, W. J. (2005). Biology and management of insect pests in North American intensively managed hardwood forest systems. *Annual Review of Entomology*, 50(1), 1–29. https://doi.org/10.1146/annurev.ento.50.071803.130431
- DAFNAE (Dipartimento di Agronomia, Animali, Alimenti, Risorse naturali e Ambiente). (2024). Scolytinae hosts and distribution database. https://www. scolytinaehostsdatabase.eu/site/it/home/ (accessed 2024-12-10).
- Dara, S. K., Barringer, L., & Arthurs, S. P. (2015). Lycorma delicatula (Hemiptera: Fulgoridae): A new invasive pest in the United States. Journal of Integrated Pest Management, 6(1), 20. https://doi.org/10.1093/jipm/pmv021
- EFSA PLH Panel (EFSA Panel on Plant Health). (2018). Guidance on quantitative pest risk assessment. EFSA Journal, 16(8), 5350. https://doi.org/10.2903/j. efsa.2018.5350
- EFSA PLH Panel (EFSA Panel on Plant Health). (2019). Guidance on commodity risk assessment for the evaluation of high risk plants dossiers. *EFSA Journal*, 17(4), 5668. https://doi.org/10.2903/j.efsa.2019.5668
- EFSA PLH Panel (EFSA Panel on Plant Health). (2024). Standard protocols for plant health scientific assessments. *EFSA Journal*, 22(9), e8891. https://doi. org/10.2903/j.efsa.2024.8891
- EFSA Scientific Committee. (2018). Scientific Opinion on the principles and methods behind EFSA's guidance on uncertainty analysis in scientific assessment. EFSA Journal, 16(1), 5122. https://doi.org/10.2903/j.efsa.2018.5122

EPPO (European and Mediterranean plant protection organization). (2024). EPPO Global Database. https://gd.eppo.int/ (accessed 2024-12-11).

- Estebanes-Gonzalez, M. L., & Baker, E. W. (1968). Arañas rojas de Mexico (Acarina: Tetranychidae). Anales. Escuela Nacional de Ciencias Biológicas, 15, 61–133.
- EUROPHYT (European Union Notification System for Plant Health Interceptions). (2024). https://ec.europa.eu/food/plants/plant-health-and-biosecurity/European-union-notification-system-plant-health-interceptionsen (accessed 2024-12-10).
- FAO (Food and Agriculture Organization of the United Nations). (2019). *ISPM (international standards for phytosanitary measures)* No 36. Integrated measures for plants for planting. https://www.ippc.int/en/publications/636
- FAO (Food and Agriculture Organization of the United Nations). (2024a). *ISPM (international standards for phytosanitary measures) No 4*. Requirements for the establishment of pest free areas. https://www.ippc.int/en/publications/614/
- FAO (Food and Agriculture Organization of the United Nations). (2024b). ISPM (International standards for phytosanitary measures) No. 5. Glossary of phytosanitary terms. FAO, Rome. https://www.ippc.int/en/publications/622/
- Farr, D. F., & Rossman, A. Y. (2024). Fungal Databases. U.S. https://fungi.ars.usda.gov/ (accessed 2024-12-11).
- Ferguson, D. C. (1975). Host records for Lepidoptera reared in eastern North America. USDA Technical Bulletin, 1521, 1–49.
- Franco-Lara, L., García, J. A., Bernal, Y. E., & Rodríguez, R. A. (2020). Diversity of the 'Candidatus Phytoplasma asteris' and 'Candidatus Phytoplasma fraxini'isolates that infect urban trees in Bogotá, Colombia. International Journal of Systematic and Evolutionary Microbiology, 70(12), 6508–6517. https:// doi.org/10.1099/ijsem.0.004553
- Index Fungorum. (2024). https://www.indexfungorum.org/ (accessed 2024-12-11).
- Gardi, C., Kaczmarek, A., Streissl, F., Civitelli, C., Do Vale Correia, C., Mikulová, A., Yuen, J., & Stancanelli, G. (2024). EFSA standard protocol for commodity risk assessment. Zenodo. https://doi.org/10.5281/zenodo.13149775
- Gomez, D. F., Johnson, A. J., De Grammont, P. C., Alfonso-Simonetti, J., Montaigne, J., Elizondo, A. I., Muiño, B. L., Ojeda, D., Vidal, J., & Hulcr, J. (2020). New records of bark and ambrosia beetles (Coleoptera: Scolytinae) from Cuba with description of a new species. *Florida Entomologist*, *102*(4), 717–724. https://doi.org/10.1653/024.102.0408
- Hoddle, M. S., Triapitsyn, S. V., & Morgan, D. J. W. (2003). Distribution and plant association records for *Homalodisca coagulata* (Hemiptera: Cicadellidae) in Florida. *Florida Entomologist*, 86(1), 89–91. https://doi.org/10.1653/0015-4040(2003)086[0089:daparf]2.0.co;2
- Kasanen, R., Hantula, J., Ostry, M., Pinon, J., & Kurkela, T. (2004). North American populations of *Entoleuca mammata* are genetically more variable than populations in Europe. *Mycological Research*, 108(7), 766–774.
- Kissinger, D. G. (1993). The insects and arachnids of Canada. Part 21. The weevils of Canada and Alaska: Volume 1-Coleoptera: Curculionoidea, excluding Scolytidae and Curculionidae. Annals-entomological society of. *America*, 86, 844.
- Li, Z., Liang, Y. M., & Tian, C. M. (2012). Characterization of the causal agent of poplar anthracnose occurring in the Beijing region. *Mycotaxon*, 120(1), 277–286. https://doi.org/10.5248/120.277
- Lim, J., Jung, S. Y., Lim, J. S., Jang, J., Kim, K. M., Lee, Y. M., & Lee, B. W. (2014). A review of host plants of Cerambycidae (Coleoptera: Chrysomeloidea) with new host records for fourteen cerambycids, including the Asian longhorn beetle (Anoplophora glabripennis Motschulsky). Korea. Korean Journal of Applied Entomology, 53(2), 111–133. https://doi.org/10.5656/ksae.2013.11.1.061
- Mayers, C. G., Harrington, T. C., Mcnew, D. L., Roeper, R. A., Biedermann, P. H., Masuya, H., & Bateman, C. C. (2020). Four mycangium types and four genera of ambrosia fungi suggest a complex history of fungus farming in the ambrosia beetle tribe Xyloterini. *Mycologia*, *112*(6), 1104–1137. https://doi. org/10.1080/00275514.2020.1755209
- Mustafa, R., Hamza, M., Rehman, A. U., Kamal, H., Tahir, M. N., Mansoor, S., Scheffler, B. E., Briddon, R. W., & Amin, I. (2022). Asymptomatic populus alba: A tree serving as a reservoir of begomoviruses and associated satellites. *Australasian Plant Pathology*, *51*(6), 577–586. https://doi.org/10.1007/s1331 3-022-00886-5
- MyCoBank (2024). https://www.mycobank.org/ (accessed 2024-12-11).
- Samin, N., Ghahari, H., & Behnood, S. (2015). A contribution to the knowledge of whiteflies (Hemiptera: Aleyrodidae) in Khorasan and Semnan provinces, Iran. Acta Phytopathologica et Entomologica Hungarica, 50(2), 287–295. https://doi.org/10.1556/038.50.2015.2.12
- Shaw, J. G. (1950). Hosts of the citrus blackfly in Mexico. Bureau of Entomology and Plant Quarantine, E-793, 16.
- Singh, A. P., Bhandari, R. S., & Verma, T. D. (2004). Important insect pests of poplars in agroforestry and strategies for their management in northwestern India. Agroforestry Systems, 63, 15–26. https://doi.org/10.1023/b:agfo.0000049429.37483.47

Sjöman, H., Östberg, J., & Nilsson, J. (2014). Review of host trees for the wood-boring pests Anoplophora glabripennis and Anoplophora chinensis: An urban forest perspective. Arboriculture & Urban Forestry, 40(3), 143–164. https://doi.org/10.48044/jauf.2014.016

Sokal, R. R., Unnash, R. S., & Thomson, B. A. (1991). *Pemphigus* revisited: Changes in geographical variation but constancy in variability and covariation. *Evolution*, 45, 1585–1605.

Terekhova, V. V., & Skrylnik, Y. (2012). Biological peculiarities of the alien for Europe Anisandrus maiche Stark (Coleoptera: Curculionidae: Scolytinae) bark beetle in Ukraine. Russian Journal of Biological Invasions, 3(2), 139–144. https://doi.org/10.1134/s2075111712020105

TRACES-NT. (2024). TRAde Control and Expert System. https://webgate.ec.europa.eu/tracesnt (accessed: 2024-12-10).

Van Rooyen, E., Paap, T., De Beer, W., Townsend, G., Fell, S., Nel, W. J., Morgan, S., Hill, M., Gonzalez, A., & Roets, F. (2021). The polyphagous shot hole borer beetle: Current status of a perfect invader in South Africa. South African Journal of Science, 117(11–12), 1–10. https://doi.org/10.17159/sajs.2021/9736

Vettraino, A. M., Sukno, S., Vannini, A., & Garbelotto, M. (2010). Diagnostic sensitivity and specificity of different methods used by two laboratories for the detection of *Phytophthora ramorum* on multiple natural hosts. *Plant Pathology*, *59*(2), 289–300. https://doi.org/10.1111/j.1365-3059.2009.02209.x
 Wene, G. P., & White, A. N. (1953). The cabbage root aphid. *The Ohio Journal of Science*, *53*(6), 332–334.

Wood, S. L., & Bright, D. E. (1992). A catalog of Scolytidae and Platypodidae (Coleoptera). Part 2: Taxonomic index. Great Basin Naturalist Memoirs, 13, 1241–1348.

Xu, Y. M., & Zhao, Z. Q. (2019). Longidoridae and Trichodoridae (Nematoda: Dorylaimida and Triplonchida). Fauna of New Zealand, 79, 149.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: EFSA PLH Panel (EFSA Panel on Plant Health), Vicent Civera, A., Baptista, P., Berlin, A., Chatzivassiliou, E., Cubero, J., Cunniffe, N., de la Peña, E., Desneux, N., Di Serio, F., Filipiak, A., Hasiów-Jaroszewska, B., Jactel, H., Landa, B. B., Maistrello, L., Makowski, D., Milonas, P., Papadopoulos, N., Potting, R., ... Gonthier, P. (2025). Commodity risk assessment of *Populus alba, Populus nigra* and *Populus tremula* plants from the UK. *EFSA Journal*, 23(3), e9305. <u>https://doi.org/10.2903/j.efsa.2025.9305</u>

APPENDIX A

Data sheets of pests selected for further evaluation

A.1 | BEMISIA TABACI (EUROPEAN POPULATIONS)

A.1.1 | Organism information

Taxonomic information	Current valid scientific name: Bemisia tabaci Synonyms: Aleurodes inconspicua, Aleurodes tabaci, Bemisia achyranthes, Bemisia bahiana, Bemisia costa-limai, Bemisia emiliae, Bemisia goldingi, Bemisia gossypiperda, Bemisia gossypiperda mosaicivectura, Bemisia hibisci, Bemisia inconspicua, Bemisia longispina, Bemisia lonicerae, Bemisia manihotis, Bemisia minima, Bemisia minuscula, Bemisia nigeriensis, Bemisia rhodesiaensis, Bemisia signata, Bemisia vayssieri Name used in the EU legislation: Bemisia tabaci Genn. (European populations) Order: Hemiptera Family: Aleyrodidae Common name: Cassava whitefly, cotton whitefly, silver-leaf whitefly, sweet-potato whitefly, tobacco whitefly Name used in the dossier: –
Group	Insects
EPPO code	BEMITA
Regulated status	 Bemisia tabaci Genn. (European populations) is listed in Annex III of Commission Implementing Regulation (EU) 2019/2072 as protected zone quarantine pest for Ireland and Sweden. The non-European populations of Bemisia tabaci are listed in Annex II. Bemisia tabaci is included in the EPPO A2 list (EPPO, 2024a). The species is a quarantine pest in Belarus, Moldova, Norway and New Zealand. It is on A1 list of Azerbaijan, Chile, Georgia, Kazakhstan, Switzerland, Ukraine and the UK. It is on A2 list of Bahrain, Russia, Türkiye, EAEU (= Eurasian Economic Union – Armenia, Belarus, Kazakhstan, Kyrgyzstan and Russia) and OIRSA (= Organismo Internacional Regional de Sanidad Agropecuaria – Belize, Costa Rica, the Dominican Republic, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama) (EPPO, 2024b).
Pest status in the UK	 Bemisia tabaci (European populations) is present in the UK, with few occurrences (CABI, 2015; EPPO, 2024c) and it is continuously intercepted in commodities imported to the UK. The intercepted populations were identified as B biotype Middle East-Asia Minor 1 (=MEAM1) and Q biotype Mediterranean (=MED) (Cuthbertson, 2013). From 1998 to 2015 there were between 7 and 35 outbreaks per year of <i>B. tabaci</i> in the UK and all the findings were subject to eradication. The UK outbreaks of <i>B. tabaci</i> have been restricted to greenhouses and there are no records of the whitefly establishing outdoors during summer (Bradshaw et al., 2019; Cuthbertson & Vänninen, 2015). According to the additional information received by the applicant: the pest is absent, pest eradicated. Sporadic and regular outbreaks of <i>B. tabaci</i> that occur under glass in Great Britain and which are subject to control measures and eradication are all derived from European populations introduced with plants from Europe (Dossier Section 5.1).
Pest status in the EU	 Bemisia tabaci is an alien species widespread in the EU – Austria, Belgium, Bulgaria, Croatia, the Republic of Cyprus, Czechia, Finland, France, Germany, Greece, Hungary, Italy, Malta, the Netherlands, Poland, Portugal, Romania, Slovenia and Spain (CABI, 2015; EPPO, 2024c). It is absent from Denmark, Estonia, Ireland, Latvia, Lithuania, Luxembourg, Slovakia and Sweden (CABI, 2015; EPPO, 2024c). In the EU, <i>B. tabaci</i> is mainly present in the greenhouses, with exception of Mediterranean coastal region (Cyprus, Greece, Malta, Italy, south of France, certain parts of Spain and Portugal), where the whitefly occurs also outdoors (EFSA PLH Panel, 2013).
Host status on Populus alba, P. nigra, P. tremula	<i>Bemisia tabaci</i> was found on <i>Populus nigra</i> in Iran in 2009 (Samin et al., 2015). There is no information on whether <i>B. tabaci</i> can also attack <i>Populus alba, P. tremula</i> or other <i>Populus</i> species.
PRA information	 Available Pest Risk Assessments: Scientific Opinion on the risks to plant health posed by <i>Bemisia tabaci</i> species complex and viruses it transmits for the EU territory (EFSA PLH Panel, 2013); UK Risk Register Details for <i>Bemisia tabaci</i> European populations (DEFRA, 2022); UK Risk Register Details for <i>Bemisia tabaci</i> non-European populations (DEFRA, 2023).
Other relevant information fe	or the assessment
Biology	 Bemisia tabaci is a cosmopolitan whitefly present on almost all continents except for Antarctica (CABI, 2015; EPPO, 2024c). In the literature it is reported as either native to Africa, Asia, India, North America or South America (De Barro et al., 2011). However, based on mtCO1 (mitochondrial cytochrome oxidase 1) sequence its origin is most likely to be sub-Saharan Africa (De Barro, 2012). Bemisia tabaci is a complex of at least 40 cryptic species that are morphologically identical but distinguishable at molecular level (Khatun et al., 2018). The species differ from each other in host association, spread capacity, transmission of viruses and resistance to insecticides (De Barro et al., 2011, Brown et al., 2023).

(Continued)

Continued)			
	 Bemisia tabaci develops through three life stages: egg, nymph (four instars) and adult (Walker et al., 2010). Nymphs of <i>B. tabaci</i> mainly feed on phloem in minor veins of the underside leaf surface (Cohen et al., 1996). Adults feed on both phloem and xylem of leaves (Janssen et al., 1989; Lei et al., 1997, 2001; Jiang et al., 1999 cited in Walker et al., 2010). Honeydew is produced by both nymphs and adults (Davidson et al., 1994). <i>Bemi tabaci</i> is multivoltine with up to 15 generations per year (Ren et al., 2001). The life cycle from egg to adult requires from 2.5 weeks up to 2 months depending on the temperature (Norman et al., 1995) and the host plant (Coudriet et al., 1985). In the southern California desert on field-grown lettuce (from 27 October 1983 to 4 January 1984), <i>B. tabaci</i> completed at least one generation (Coudriet et al., 1985). In Israel the reproduction of <i>B. tabaci</i> was much reduced in winter months, but adults emerging in December survived and started ovipositing at the end of th cold season (Avidov, 1956). The most cold-tolerant stage are eggs (-2^o, -6^o, -10^oC) and the least tolerant are lar nymphs. Short periods of exposure in 0° to -6°C have little effect on mortality. As the temperature lowers to -10°C, the duration of time required to cause significant mortality shortens dramatically (Simmons & Elsey, 195 Females can lay more than 300 eggs (Gerling et al., 1986), which can be found mainly on the underside of the leaves (CABI, 2015). Females develop from fertilised and males four unstality and the set urg oblaw brown. Their size is about 0.19–0.20 mm long and 0.10–0.12 mm wide. First instar nymph (=crawler) is scale-like, elliptical, darker yellow in colour and about 0.26 mm long and 0.63 mm wide) 0.24 mm wide), third (0.55 mm long and 0.55 mm wide) and fourth instar nymph (0.86 mm long and 0.63 mm wide) (Hill, 1969). Fourth instar nymph (=pupa) stops feedi and moults into an adult (Walker et al., 2009, citing othe		
Symptoms	Main type of symptoms	Main symptoms of <i>B. tabaci</i> on plants are chlorotic spotting, decrease of plant growth, deformation of fruits, deformation of leaves, intervein yellowing, leaf yellowing, leaf curling, leaf crumpling, leaf vein thickening, leaf enations, leaf cupping, leaf loss, necrotic lesions on stems, plant stunting, reduced flowering, reduced fruit development, silvering of leaves, stem twisting, vein yellowing, wilting, yellow blotching of leaves, yellow mosaic of leaves, presence of honeydew and sooty mould. These symptoms are plant responses to the feeding of the whitefly and to the presence of transmitted viruses (EPPO, 2004; EFSA PLH Panel, 2013; CABI, 2015).	
	Presence of	Symptoms of <i>B. tabaci</i> being present on the plants are usually visible. However, <i>B.</i>	
	asymptomatic plants Confusion with other pests	 <i>tabaci</i> is a vector of several viruses and their infection could be asymptomatic. <i>Bemisia tabaci</i> can be easily confused with other whitefly species such as <i>B.</i> <i>afer, Trialeurodes lauri, T. packardi, T. ricini, T. vaporariorum</i> and <i>T. variabilis</i>. A microscopic slide is needed for morphological identification (EPPO, 2004). Different species of <i>B. tabaci</i> complex can be distinguished using molecular methods (Brown et al., 2023; De Barro et al., 2011). 	
Host plant range	Bemisia tabaci has a wide hos	st range, including more than 1000 different plant species (Abd-Rabou & Simmons,	
	 2010). Some of the many hosts of <i>B. tabaci</i> are <i>Abelmoschus esculentus</i>, <i>Amaranthus blitoides</i>, <i>A. retroflexus</i>, <i>Aracl hypogaea</i>, <i>Atriplex semibaccata</i>, <i>Bellis perennis</i>, <i>Borago officinalis</i>, <i>Brassica oleracea</i> var. <i>botrytis</i>, <i>B. olera gemmifera</i>, <i>B. oleracea</i> var. <i>italica</i>, <i>Bryonia dioica</i>, <i>Cajanus cajan</i>, <i>Capsella bursa-pastoris</i>, <i>Capsicum anne Citrus spp.</i>, <i>Crataegus spp.</i>, <i>Cucumis sativus</i>, <i>Cucurbita pepo</i>, <i>Erigeron canadensis</i>, <i>Euphorbia pulcherrime Gerbera jamesonii</i>, <i>Glycine max</i>, <i>Gossypium spp.</i>, <i>G. hirsutum</i>, <i>Hedera helix</i>, <i>Ipomoea batatas</i>, <i>Lactuca sa serriola</i>, <i>Lavandula coronopifolia</i>, <i>Ligustrum lucidum</i>, <i>L. quihoui</i>, <i>L. vicaryiis</i>, <i>Manihot esculenta</i>, <i>Melissa c Nicotiana tabacum</i>, <i>Ocimum basilicum</i>, <i>Origanum majorana</i>, <i>Oxalis pes-caprae</i>, <i>Phaseolus spp.</i>, <i>P. vulga nigrum</i>, <i>Potentilla spp.</i>, <i>Prunus spp.</i>, <i>Rosa spp.</i>, <i>Rubus fruticosus</i>, <i>Salvia officinalis</i>, <i>S. rosmarinus</i>, <i>Senecio Sinningia speciosa</i>, <i>Solanum lycopersicum</i>, <i>S. melongena</i>, <i>S. nigrum</i>, <i>S. tuberosum</i>, <i>Sonchus oleraceus</i>, <i>Stemedia</i>, <i>Tagetes erecta</i>, <i>Taraxacum officinale</i>, <i>Thymus serpyllum</i>, <i>Urtica urens</i>, <i>Vitis vinifera</i> and many mor PLH Panel, 2013; CABI, 2015; EPPO, 2024c; Li et al., 2011). For a full host list refer to CABI (2015), EFSA PLH Panel (2013) and EPPO (2024c) and Li et al. (2011) 		

(Continued) Reported evidence of impact	Bemisia tabaci (European populations) is EU protected zone quarantine pest.
Evidence that the commodity is a pathway	<i>Bemisia tabaci</i> is continuously intercepted in the EU on different commodities including plants for planting (EUROPHYT, 2024; TRACES-NT, 2024). Therefore, the commodity is a potential pathway for <i>B. tabac</i> . as <i>B. tabaci</i> was found to be associated with <i>Populus nigra</i> in Iran (Samin et al., 2015). Plants can carry leaves at the time of export which can host all life stages of the pest.
Surveillance information	<i>Bemisia tabaci</i> is regulated quarantine pest in the UK. As such, the policy for any outbreak is to eradicate the population. The UK makes many interceptions of <i>B. tabaci</i> and experiences a few outbreaks each year (356 interceptions and outbreaks in 2021), but all outbreaks are under protection and subject to eradication measures. This pest has never established outdoors in the UK (EFSA PLH Panel, 2024).

COMMODITY RISK ASSESSMENT OF POPULUS PLANTS FROM THE UK

A.1.2 | Possibility of pest presence in the nursery

A.1.2.1 | Possibility of entry from the surrounding environment

Bemisia tabaci (European populations) is present in glasshouses in the UK with few occurrences (location not specified) (CABI, 2015; EPPO, 2024c) and is continuously intercepted on commodities to the UK. The UK outbreaks of *B. tabaci* have been restricted to glasshouses and there are no records of *B. tabaci* establishing outdoors during summer (Bradshaw et al., 2019; Cuthbertson & Vänninen, 2015). Bradshaw et al. (2019) indicate that theoretically *B. tabaci* (in summertime) could complete one generation across most of Scotland, and one to three generations over England and Wales. However, the temperatures experienced during the cold days and nights during summer may be low enough to cause chilling injury to *B. tabaci*, thereby inhibiting development and preventing establishment in the UK. It is unlikely, therefore, that this pest will establish outdoors in the UK under current climate conditions.

The possible entry of *B. tabaci* from surrounding environment to the nurseries may occur through adult dispersal and passively on wind currents (Byrne, 1999; Cohen et al., 1988; EFSA PLH Panel, 2013).

Bemisia tabaci is a polyphagous species that can infest a number of different plants. Suitable hosts of *B. tabaci* like Brassica rapa, Fraxinus spp., Ilex spp., Quercus spp., Solanum spp. and Triticum spp. are present within 2 km from the nurseries (Dossier Sections 1.1, 1.2 and 1.3).

Uncertainties

36 of 120

- Exact locations where the whitefly is present.
- Possibility of spread beyond the infested greenhouses.
- Possibility of the whitefly to survive the UK summer in outdoor conditions.

Taking into consideration the above evidence and uncertainties, the Panel considers that it is possible for the pest to enter the nurseries from surrounding environment, even though it is only reported to be present in greenhouses. In the surrounding area suitable hosts are present and the pest can spread by wind and adult flight.

A.1.2.2 | Possibility of entry with new plants/seeds

The starting materials of *P. alba, P. nigra and P. tremula* are either seeds, seedlings or cuttings. Seeds and seedlings are either from the UK (certified with UK Plant Passports) or the EU (mostly the Netherlands, Belgium and France) (certified with phytosanitary certificates) (Dossier Sections 1.1, 1.2 and 1.3). Seeds are not a pathway for the whitefly.

In the nurseries many other plants are cultivated (Dossier Sections 3.1, 3.2 and 3.3). Out of them *Acer* spp., *Acacia* spp., *Crataegus* spp., *Hedera* spp., *Prunus* spp., *Pyrus* spp., *Rosa* spp., *Salvia* spp., *Viburnum* spp. and many more plants are potential suitable hosts of the whitefly. However, there is no information on how and where the plants are produced. Therefore, if the plants are first produced in another nursery, the whitefly could possibly travel with them.

The nurseries are using virgin peat or peat-free compost as a growing media, which is a mixture of coir, tree bark, wood fibre, etc., heat-treated by commercial suppliers during production to eliminate pests and diseases (Dossier Sections 1.1, 1.2 and 1.3). Growing media is not a pathway for the whitefly.

Uncertainties

 No information is available on the provenance of plants other than *Populus* used for plant production in the nurseries.

Taking into consideration the above evidence and uncertainties, the Panel considers that it is possible for the pest to enter the nurseries with new seedlings of *Populus* and new plants of other species used for plant production in the area. The entry of the pest with seeds and the growing media the Panel considers as not possible.
A.1.2.3 | Possibility of spread within the nursery

Populus plants are grown both in containers outdoors and in fields. There are no mother plants present in the nurseries and none of the nurseries expected to export to the EU produce plants from grafting (Dossier Sections 1.1, 1.2 and 1.3).

The whitefly can attack other suitable plants (such as Acer spp., Acacia spp., Crataegus spp., Hedera spp., etc.) and noncultivated herbaceous plants (Bellis perennis, Potentilla spp., Taraxacum officinale) present within the nurseries and hedges surrounding the nurseries (Crataegus spp., Hedera helix, llex spp. and Prunus spp.).

There are greenhouses within the nurseries (Dossier Sections 1.1, 1.2 and 1.3).

The whitefly can spread within the nurseries by adult flight or wind. Spread within the nurseries through equipment and tools is not relevant.

Uncertainties

- Possibility of the whitefly to survive the UK summer in outdoor conditions.
- Possibility that greenhouses are heated which allows the pest to overwinter.

Taking into consideration the above evidence and uncertainties, the Panel considers that the spread of the pest within the nurseries is possible either by wind or by active flight.

A.1.3 | Information from interceptions

In the EUROPHYT/TRACES-NT database there are no records of notification of *Populus, P. alba, P. nigra* and *P. tremula* plants for planting neither from the UK nor from other countries due to the presence of *B. tabaci* between the years 1995 and October 2024 (EUROPHYT, 2024; TRACES-NT, 2024).

There were two interceptions of *B. tabaci* from the UK in 2007 and 2015 on other plants already planted likely produced under protected conditions (EUROPHYT, 2024) and one interception on other live plants (including their roots) in October 2024 (TRACES-NT, 2024).

A.1.4 | Evaluation of the risk mitigation measures

In the table below, all risk mitigation measures currently applied in the UK are listed and an indication of their effectiveness on *B. tabaci* is provided. The description of the risk mitigation measures currently applied in the UK is provided in Table 7.

N	Risk mitigation measure	Effect on the pest	Evaluation and uncertainties
1	Registration of production sites	Yes	As the plant passport is very similar to the EU one, plants shall be free from quarantine pests. <u>Uncertainties</u> : – None
2	Physical separation	No	Not relevant, there is no separation between production areas for the export and the local market.
3	Certified plant material	Yes	 Seeds are not a pathway for <i>B. tabaci</i>. As the plant passport is very similar to the EU one, seedlings shall be free from quarantine pests. Phytosanitary certificates should ensure that seedlings are free from quarantine pests. <u>Uncertainties</u>: None
4	Growing media	No	Not relevant, growing media is not a pathway of <i>B. tabaci</i> .
5	Surveillance, monitoring and sampling	Yes	 Plant material is regularly monitored for plant health issues. They must meet the required national sanitary standards. Monitoring should be affective in finding infestation of <i>B. tabaci</i>. <u>Uncertainties</u>: Difficulty of detecting low levels of infestation. Difficulty in the identification by morphological traits.
6	Hygiene measures	Yes	 Weeding can have some effect on the reduction of <i>B. tabaci</i> populations. The other measures are not relevant. <u>Uncertainties</u>: None
7	Removal of infested plant material	Yes	Removing infested plant material can have some effect on the reduction of <i>B. tabaci</i> populations. <u>Uncertainties</u> : - None
8	Irrigation water	No	Not relevant, water is not a pathway of <i>B. tabaci</i> .

(Continued)

N	Risk mitigation measure	Effect on the pest	Evaluation and uncertainties
9	Application of pest control products	Yes	 Plant protection products are only used when necessary and records of all plant protection treatments are kept. It may have an effect on the pest. <u>Uncertainties</u>: No information about the specific treatments. No information on the effect of treatments against the pest.
10	Measures against soil pests	No	Not relevant to the pest.
11	Inspections and management of plants before export	Yes	 Exporting plants should meet phytosanitary certificate requirements. Inspection before export should be affective in finding infestation of <i>B. tabaci</i>. However, a low level of infestation by <i>B. tabaci</i> could go undetected. Inspection is performed between 1 day and 2 weeks before the export, but a reinfestation can occur during this period. <u>Uncertainties:</u> Capacity of detection of low levels of infestation. Difficulty in the identification by morphological traits. Exact duration of the period between inspection and export.
12	Separation during transport to the destination	Yes	 The pest could spread from infested plants to non-infested plants during transport to the destination. <u>Uncertainties</u>: None

A.1.5 | Overall likelihood of pest freedom for bare root plants

A.1.5.1 | Reasoning for a scenario which would lead to a reasonably low number of infected bare root plants

This scenario assumes that the pest is not present in the nursery area.

A.1.5.2 | Reasoning for a scenario which would lead to a reasonably high number of infected bare root plants

This scenario assumes high pest pressure in and around nurseries especially when in proximity of greenhouses. Leaves may be present and there is a high uncertainty of probability of detection in the canopies. Seven years old plants have more leaves compared to younger plants and hence more possibilities for the pest to hide and being overlooked. Reasoning for a central scenario equally likely to over- or underestimate the number of infected bundles of whips and seedlings (Median).

The scenario assumes low values for the central scenario because *B. tabaci* is not expected to be present outdoors and because of the uncertainty about the host status of *B. tabaci* on *Populus* species other than *Populus nigra*. However, it has been considered also that pest outbreaks are reported in greenhouses in the UK, and that that visual inspections could overlook the pest.

A.1.5.3 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The Panel expresses the maximum uncertainty with the first quartile, and a lower uncertainty with the third quartile, mainly because there is relatively high distance between the greenhouse and the commodity outside. It is very unlikely to be present outdoors and *Populus* is not a major host. It is a quarantine pest in the UK and therefore more likely to be detected in the greenhouse where measures must be taken.

A.1.5.4 | Elicitation outcomes of the assessment of the pest freedom for *Bemisia tabaci* (European populations) on bare root plants

The following Tables show the elicited and fitted values for pest infection (Table A.1) and pest freedom (Table A.2).

TABLE A.1 Elicited and fitted values of the uncertainty distribution of pest infestation by *Bemisia tabaci* per 10,000 plants/bundles.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50 %	67 %	75%	83%	90%	95%	97.5%	99 %
Elicited values	0					6		12		25					50
EKE	0.137	0.384	0.839	1.85	3.35	5.42	7.70	13.1	19.9	24.1	29.4	35.0	41.0	45.6	49.9

Note: The EKE results is the BetaGeneral (0.89141, 2.423, 0, 59.5) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants/bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.2.

TABLE A.2 The uncertainty distribution of plants free of Bemisia tabaci per 10,000 plants/bundles calculated by Table A.1.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95 %	97.5 %	99 %
Values	9950					9975		9988		9994					10,000
EKE results	9950	9954	9959	9965	9971	9976	9980	9987	9992	9995	9997	9998	9999.2	9999.6	9999.9







Pestfree plants/bundles [number out of 10,000]

FIGURE A.1 (Continued)



FIGURE A.1 (A) Elicited uncertainty of pest infection per 10,000 plants/bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free bare root plants/bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants/bundles.

A.1.6 | Overall likelihood of pest freedom for cell grown plants

A.1.6.1 | Reasoning for a scenario which would lead to a reasonably low number of infected cell grown plants

This scenario assumes that the pest is not present in the nursery area.

A.1.6.2 | Reasoning for a scenario which would lead to a reasonably high number of infected cell grown plants

This scenario assumes high pest pressure in and around nurseries especially when in proximity with greenhouses. It also assumes, that cell grown plants may be stored nearby the greenhouses or be grown inside the greenhouses at the beginning of the cultivation, which makes it more likely that they could be infested with *B- tabaci*. Moreover, cell grown plants are exported with leaves.

A.1.6.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infected bundles of whips and seedlings (Median)

The scenario assumes low values for the central scenario because *B. tabaci* is not expected to be present outdoors and because there is uncertainty about the host status of *B. tabaci* on *Populus*. In addition, cell grown plants are smaller compared to potted plants, so they are easier to inspect. However, it has also been taken into account that the pest is repeatedly intercepted in the UK in glasshouses, that visual inspections could miss the pest and that it is possible that there could be spread to plants grown outdoors from the glasshouse.

A.1.6.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The Panel expresses the maximum uncertainty with the first quartile, and a lower uncertainty with the third quartile, mainly because plants are relatively small and easy to inspect. It is very unlikely to be present outdoors and *Populus* is not a major host. The pest is a quarantine pest in the UK and therefore more likely to be detected in the greenhouse where measures must be taken.

A.1.6.5 | Elicitation outcomes of the assessment of the pest freedom for *Bemisia tabaci* (European populations) on cell grown plants

The following Tables show the elicited and fitted values for pest infection (Table A.3) and pest freedom (Table A.4).

Percentile	1%	2.5%	5%	10 %	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5 %	99 %
Elicited values	0					9		18		35					70
EKE	0.292	0.746	1.52	3.14	5.41	8.41	11.7	19.1	28.3	34.1	41.2	48.9	57.2	63.6	70.0

Note: The EKE results is the BetaGeneral (0.98178, 2.6842, 0, 85.5) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.4.

TABLE A.4 The uncertainty distribution of plants free of Bemisia tabaci per 10,000 bundles calculated by Table A.3.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67%	75%	83%	90%	95%	97.5%	99 %
Values	9930					9965		9982		9991					10,000
EKE results	9930	9936	9943	9951	9959	9966	9972	9981	9988	9992	9995	9997	9998	9999.3	9999.7







Pestfree bundles [number out of 10,000]

46 of 120

FIGURE A.2 (Continued)



FIGURE A.2 (A) Elicited uncertainty of pest infection per 10,000 bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 bundles.

A.1.7 | Overall likelihood of pest freedom for plants in pots

A.1.7.1 | Reasoning for a scenario which would lead to a reasonably low number of infested plants in pots

This scenario assumes that the pest is not present in the nursery area.

A.1.7.2 | Reasoning for a scenario which would lead to a reasonably high number of infested plants in pots

This scenario assumes high pest pressure in and around nurseries especially when in proximity with greenhouses. It also assumes high inspection difficulty in the canopy of large trees, so there are more possibilities that the pest is unnoticed. However, *Populus* may not be a good host for *B. tabaci* as no reports of damage on *Populus* were found and there is only one record of *B. tabaci* on *Populus* without damage in Iran.

A.1.7.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infested plants in pots (Median)

The scenario assumes low values for the central scenario because *B. tabaci* is not expected to be present outdoors and because there is uncertainty about the host status of *B. tabaci* on *Populus*. However, it has also been taken into account that the pest is repeatedly intercepted in the UK in glasshouses, that visual inspections could miss the pest and that it is possible that there could be spread to plants grown outdoors from the glasshouse.

A.1.7.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The Panel expresses the maximum uncertainty with the first quartile, and a slightly lower uncertainty with the third quartile, mainly because there is relatively high distance between the greenhouse and the commodity outside. Moreover, it is very unlikely that the pest is present outdoors and *Populus* is not a major host. The pest is a quarantine one in the UK and therefore it is more likely to be detected in the greenhouse where measures must be taken.

A.1.7.5 | Elicitation outcomes of the assessment of the pest freedom for *Bemisia tabaci* (European populations) on plants in pots

The following Tables show the elicited and fitted values for pest infection (Table A.5) and pest freedom (Table A.6).

TABLE A.5 Elicited and fitted values of the uncertainty distribution of pest infestation by Bemisia tabaci per 10,000 plants.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95 %	97.5%	99 %
Elicited values	0					10		20		40					75
EKE	0.247	0.681	1.47	3.19	5.72	9.15	12.9	21.5	32.2	38.8	46.7	54.9	63.3	69.5	75.1

Note: The EKE results is the BetaGeneral (0.9073, 2.1215, 0, 85.5) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.6.

TABLE A.6 The uncertainty distribution of plants free of Bemisia tabaci per 10,000 plants calculated by Table A.5.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5 %	99 %
Values	9925					9960		9980		9990					10,000
EKE results	9925	9931	9937	9945	9953	9961	9968	9978	9987	9991	9994	9997	9998.5	9999.3	9999.8







Pestfree plants [number out of 10,000]

FIGURE A.3 (Continued)



FIGURE A.3 (A) Elicited uncertainty of pest infection per 10,000 plants (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free plants per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants.

53 of 120

A.1.8 | Reference list

Abd-Rabou, S., and Simmons, A. M. (2010). Survey of reproductive host plants of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in Egypt, including new host records. *Entomological News*, 121(5), 456–465. https://doi.org/10.3157/021.121.0507

Avidov, Z. (1956). Bionomics of the tobacco whitefly (Bemisia tabaci Cennad.) in Israel. Ktavin, 7, 25-41.

- Bradshaw, C. D., Hemming, D., Baker, R., Everatt, M., Eyre, D., and Korycinska, A. (2019). A novel approach for exploring climatic factors limiting current pest distributions: A case study of *Bemisia tabaci* in north-west Europe and assessment of potential future establishment in the United Kingdom under climate change. *PLoS One*, 14(8), e0221057. https://doi.org/10.1371/journal.pone.0221057
- Byrne, D. N. (1999). Migration and dispersal by the sweet potato whitefly, *Bemisia* tabaci. *Agricultural and Forest Meteorology*, 97(4), 309–316. https://doi. org/10.1016/s0168-1923(99)00074-x
- CABI (Centre for Agriculture and Bioscience International). (2015). *Bemisia tabaci* (tobacco whitefly). https://www.cabi.org/cpc/datasheet/8927#F8A36 FF8-D287-4CBD-A0C8-B380F2CFB753 (accessed: 2024/10/29).
- Cohen, S., Kern, J., Harpaz, I., and Ben-Joseph, R. (1988). Epidemiological studies of the tomato yellow leaf curl virus (TYLCV) in the Jordan Valley, Israel. *Phytoparasitica*, 16(3), 259. https://doi.org/10.1007/bf02979527
- Cohen, A. C., Henneberry, T. J., and Chu, C. C. (1996). Geometric relationships between whitefly feeding behavior and vascular bundle arrangements. Entomologia Experimentalis et Applicata, 78(2), 135–142. https://doi.org/10.1111/j.1570-7458.1996.tb00774.x
- Coudriet, D. L., Prabhaker, N., Kishaba, A. N., and Meyerdirk, D. E. (1985). Variation in developmental rate on different host and overwintering of the sweetpotato whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae). *Environmental Entomology*, *14*, 516–519. https://doi.org/10.1093/ee/14.4.516
- Cuthbertson, A. G. (2013). Update on the status of *Bemisia tabaci* in the UK and the use of entomopathogenic fungi within eradication programmes. *Insects*, 4(2), 198–205. https://doi.org/10.3390/insects4020198
- Cuthbertson, A. G., and Vänninen, I. (2015). The importance of maintaining protected zone status against *Bemisia tabaci. Insects*, 6(2), 432–441. https://doi.org/10.3390/insects6020432
- Davidson, E. W., Segura, B. J., Steele, T., & Hendrix, D. L. (1994). Microorganisms influence the composition of honeydew produced by the silverleaf whitefly, *Bemisia argentifolii. Journal of Insect Physiology*, 40(12), 1069–1076. https://doi.org/10.1016/0022-1910(94)90060-4
- De Barro, P. J. (2012). The Bemisia tabaci species complex: Questions to guide future research. Journal of Integrative Agriculture, 11, 187–196. https://doi. org/10.1016/s2095-3119(12)60003-3
- De Barro, P. J., Liu, S. S., Boykin, L. M., and Dinsdale, A. B. (2011). *Bemisia tabaci*: A statement of species status. *Annual Review of Entomology*, 56, 1–19. https://doi.org/10.1146/annurev-ento-112408-085504
- DEFRA (Department for Environment, Food and Rural Affairs). (2022). UK Risk Register Details for *Bemisia tabaci* European populations. https://planthealt hportal.defra.gov.uk/pests-and-diseases/uk-plant-health-risk-register/viewPestRisks.cfm?cslref=13756&riskId=27242 (accessed: 2024/10/29).
- DEFRA (Department for Environment, Food and Rural Affairs). (2023). UK Risk Register Details for *Bemisia tabaci* non-European populations. https://plant healthportal.defra.gov.uk/pests-and-diseases/uk-plant-health-risk-register/viewPestRisks.cfm?cslref=13756&riskld=13756 (accessed: 2024/10/29).
- EFSA PLH Panel (EFSA Panel on Plant Health). (2013). Scientific Opinion on the risks to plant health posed by *Bemisia tabaci* species complex and viruses it transmits for the EU territory. *EFSA Journal*, *11*(4), 3162. https://doi.org/10.2903/j.efsa.2013.3162
- EFSA PLH Panel (EFSA Panel on Plant Health), Bragard, C., Baptista, P., Chatzivassiliou, E., Di Serio, F., Jaques Miret, J. A., Justesen, A. F., MacLeod, A., Magnusson, C. S., Milonas, P., Navas- Cortes, J. A., Parnell, S., Potting, R., Reignault, P. L., Stefani, E., Thulke, H.-H., Van der Werf, W., Vicent Civera, A., Yuen, J., Zappalà, L., Battisti, A., Mas, H., Rigling, D., Faccoli, M., Mikulová, A., Mosbach-Schulz, O., Stergulc, F., Streissl, F., and Gonthier, P. (2024). Commodity risk assessment of *Ligustrum ovalifolium* and *Ligustrum vulgare* plants from the UK. *EFSA Journal*, 22(3), e8648. https://doi.org/10. 2903/j.efsa.2024.8648
- El-Helaly, M. S., El-Shazli, A. Y., & El-Gayar, F. H. (1971). Biological studies on *Bemisia tabaci* Genn. (Homopt., Aleyrodidae) in Egypt 1. Zeitschrift für Angewandte Entomologie, 69(1–4), 48–55. https://doi.org/10.1111/j.1439-0418.1971.tb03181.x
- EPPO (European and Mediterranean Plant Protection Organisation). (2004). Diagnostic protocols for regulated pests *Bemisia tabaci*, PM 7/35(1). *OEPP/* EPPO Bulletin, 34, 281–288.
- EPPO (European and Mediterranean Plant Protection Organization). (2024a). EPPO A2 List of pests recommended for regulation as quarantine pests, version 2024–09: https://www.eppo.int/ACTIVITIES/plant_quarantine/A2_list (accessed: 2024/10/29).
- EPPO (European and Mediterranean Plant Protection Organization). (2024b). *Bemisia tabaci* (BEMITA), Categorization. https://gd.eppo.int/taxon/ BEMITA/categorization (accessed: 2024/10/29).
- EPPO (European and Mediterranean Plant Protection Organization). (2024c). Bemisia tabaci (BEMITA), Distribution. https://gd.eppo.int/taxon/BEMITA/ distribution (accessed: 2024/10/29).
- EUROPHYT (European Union Notification System for Plant Health Interceptions). (2024). https://food.ec.europa.eu/plants/plant-health-and-biosecurity/ europhyt_en (accessed: 2024/12/10).
- Fiallo-Olivé, E., Pan, L. L., Liu, S. S., and Navas-Castillo, J. (2020). Transmission of begomoviruses and other whitefly-borne viruses: Dependence on the vector species. *Phytopathology*, *110*(1), 10–17. https://doi.org/10.1094/phyto-07-19-0273-fi
- Gerling, D., Horowitz, A. R., and Baumgaertner, J. (1986). Autecology of *Bemisia tabaci. Agriculture, Ecosystems and Environment*, 17(1–2), 5–19. https://doi. org/10.1016/0167-8809(86)90022-8
- Hill, B. G. (1969). A morphological comparison between two species of whitefly, *Trialeurodes vaporariorum* (Westw.) and *Bemisia tabaci* (Genn.) (Homoptera: Aleurodidae) which occur on tobacco in the Transvaal. *Phytophylactica*, 1(3–4), 127–146.
- Janssen, J. A. M., Tjallingii, W. F., and van Lenteren, J. C. (1989). Electrical recording and ultrastructure of stylet penetration by the greenhouse whitefly. Entomology Experience Apply, 52, 69–81. https://doi.org/10.1111/j.1570-7458.1989.tb01250.x
- Jiang, Y. X., Lei, H., Collar, J. L., Martin, B., Muñiz, M., and Fereres, A. (1999). Probing and feeding behavior of two distinct biotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) on tomato plants. *Journal of Economic Entomology*, *92*, 357–366. https://doi.org/10.1093/jee/92.2.357
- Khatun, M. F., Jahan, S. H., Lee, S., and Lee, K. Y. (2018). Genetic diversity and geographic distribution of the *Bemisia tabaci* species complex in Bangladesh. Acta Tropica, 187, 28–36. https://doi.org/10.1016/j.actatropica.2018.07.021
- Lei, H., Tjallingii, W. F., & van Lenteren, J. C. (1997). Effect of tethering during EPG recorded probing byadults of the greenhouse whitefly. *Journal of Applied Entomology*, 121, 211–217. https://doi.org/10.1111/j.1439-0418.1997.tb01395.x
- Lei, H., van Lenteren, J. C., & Xu, R. M. (2001). Effects of plant tissue factors on the acceptance of four greenhouse vegetable host plants by the greenhouse whitefly: An electrical penetration graph (EPG) study. European Journal of Entomology, 98, 31–36. https://doi.org/10.14411/eje.2001.005
- Li, S.-J., Xue, X., Ahmed, M. Z., Ren, S.-X., Du, Y.-Z., Wu, J.-H., Cuthbertson, A. G. S., & Qiu, B.-L. (2011). Host plants and natural enemies of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in China. *Insect Science*, *18*(1), 101–120. https://doi.org/10.1111/j.1744-7917.2010.01395.x
- Norman, J. W., Stansty, D. G., Ellsworth, P. A., & Toscano, N. C. P. C. (1995). Management of silverleaf whitefly: A comprehensive manual on the biology, economic impact and control tactics. USDA/CSREES Grant Pub. 93-EPIX-1-0102. 13 pp.
- Price, J. F., & Taborsky, D. (1992). Movement of immature *Bemisia tabaci* (Homoptera: Aleyrodidae) on poinsettia leaves. *The Florida Entomologist*, 75(1), 151–153. https://doi.org/10.2307/3495495

Ren, S.-X., Wang, Z.-Z., Qiu, B.-L., & Xiao, Y. (2001). The pest status of *Bemisia tabaci* in China and non-chemical control strategies. *Insect Science*, 8(3), 279–288. https://doi.org/10.1111/j.1744-7917.2001.tb00453.x

Samin, N., Ghahari, H., & Behnood, S. (2015). A contribution to the knowledge of whiteflies (Hemiptera: Aleyrodidae) in Khorasan and Semnan provinces, Iran. Acta Phytopathologica et Entomologica Hungarica, 50(2), 287–295. https://doi.org/10.1556/038.50.2015.2.12

Simmons, A. M., and Elsey, K. D. (1995). Overwintering and cold tolerance of *Bemisia argentifolii* (Homoptera: Aleyrodidae) in coastal South Carolina. Journal of Entomological Science, 30(4), 497–506. https://doi.org/10.18474/0749-8004-30.4.497

Summers, C. G., Newton, Jr A. S., and Estrada, D. (1996). Intraplant and interplant movement of *Bemisia argentifolii* (Homoptera: Aleyrodidae) crawlers. *Environmental Entomology*, 25(6), 1360–1364. https://doi.org/10.1093/ee/25.6.1360

TRACES-NT (TRAde Control and Expert System). (2024). https://webgate.ec.europa.eu/tracesnt (accessed: 2024/12/10).

Walker, G. P., Perring, T. M., and Freeman, T. P. (2010). Life history, functional anatomy, feeding and mating behavior. In Stansly, P. A., and Naranjo, S. E. (eds.), *Bemisia*: Bionomics and management of a global pest, Springer, Dordrecht, 109–160. https://doi.org/10.1007/978-90-481-2460-2_4

A.2 | ENTOLEUCA MAMMATA

A.2.1 | Organism information

Taxonomic information	Current valid scientific name: Entoleuca mammata Synonyms: Anthostoma blakei, Anthostoma morsei, Fuckelia morsei, Hypoxylon blakei, Hypoxylon holwayi, Hypoxylon mammatum, Hypoxylon morsei, Hypoxylon pauperatum, Hypoxylon pruinatum, Nemania mammata, Rosellinia pruinata, Sphaeria mammata, Sphaeria pruinata (according to index Fungorum) Name used in the EU legislation: Entoleuca mammata (Wahlenb.) Rogers and JU Order: Xylariales Family: Xylariaceae Common name: Hypoxylon canker of poplar, canker of poplar, canker of aspen Name used in the dossier: Entoleuca mammata
Group	Fungi
EPPO code	НҮРОМА
Regulated status	<i>Entoleuca mammata</i> is listed in Annex III of Commission Implementing Regulation (EU) 2019/2072 as protected zone quarantine pest for Ireland and the UK (Northern Ireland). The pathogen is quarantine pest in China and Israel and is on the A1 list of Türkiye (EPPO, 2024a).
Pest status in the UK	 Entoleuca mammata is present in the UK, with few occurrences in England, Wales, Channel Islands and Scotland (CABI, 2019; EPPO, 2024b; Granmo et al., 1999; Matthiassen, 1993). According to the Dossier Section 2.0 the pathogen is present in the UK: not widely distributed and not under official control.
Pest status in the EU	<i>Entoleuca mammata</i> is present in the following EU MS: Austria, Belgium, Croatia, Czechia, Denmark, Estonia, Finland, France, Germany, Greece, Italy, Latvia, Lithuania, the Netherlands, Poland, Slovakia, Slovenia, Spain and Sweden (EFSA PLH Panel, 2023; EPPO, 2024b).
Host status on Populus alba, P. nigra and P. tremula	 Populus alba, P. nigra and P. tremula are hosts of E. mammata (EFSA PLH Panel, 2023). P. tremula is considered the main host in Europe, whereas P. nigra is listed only as minor host (EFSA PLH Panel, 2017). According to Ostry (2013), the pest was found in plantations of P. nigra var. betulifolia × P. nigra 'Volga', P. nigra var. betulifolia × P. balsamifera subsp. trichocarpa and P. deltoides × P. nigra 'Incrassata'. Entoleuca mammata is also host for the hybrid P. tremula × P. tremuloides (Ostry, 2013).
PRA information	Pest Risk Assessments available: – Pest categorisation of <i>Entoleuca mammata</i> (EFSA PLH Panel, 2017); – Express Pest Risk Analysis: <i>Entoleuca mammata</i> (Klejdysz et al. 2018); – UK Risk Register Details for <i>Entoleuca mammata</i> (DEFRA, 2023).
Other relevant information	for the assessment
Biology	 Entoleuca mammata causes canker disease in Populus tremuloides and P. tremula as primary hosts, but other hardwood species can be also affected as minor hosts (EFSA PLH Panel, 2017). The fungus is also known as primary saprophyte on several Salix species (Matthiasen, 1993). E. mammata is thought to be native to North America and introduced into Europe several centuries ago (Kasanen et al., 2004). It is now largely spread in the temperate zones of the northern hemisphere in North America, Europe and Asia. Entoleuca mammata is present in Canada and in several states of the USA, mostly in the north. In Asia, it is only found in the Korea Republic on decayed wood (Lee et al., 2000). In Europe, in addition to the mentioned EU MS and the UK (see above), it is also reported from Andorra, Bosnia and Herzegovina, Montenegro, North Macedonia, Russia (Southern Russia and Western Siberia), Serbia, Switzerland, Ukraine (CABI, 2019; EPPO, 2024c) and Norway (Granmo et al., 1999; NBIC, 2021). The ascospores of <i>E. mammata</i> can infect the living wood of the hosts penetrating in the periderm and invading tissues under bark through mechanical wounds and injuries, often caused by woodpeckers and insects (Anderson et al., 1979a; Ostry & Anderson, 1983); water stress can increase host susceptibility (EFSA PLH Panel, 2017; EFSA PLH Panel, 2013). Infection usually starts from branches and twigs and then can spread to the main stem. Entoleuca mammata is most frequently found on stems about 1.5–2.5 m above the ground (Matthiasen, 1993). The cankers expand very rapidly (7–8 cm per month) in summar, and more slowly during winter; branches and stems can be girdled causing drying and breakage. E. mammata mostly develops in the range from 8 to 32°C; the optimum temperature is 28°C; toxins host-specific produced by the fungus are involved in pathogenesis (EFSA PLH Panel, 2017; EFPO, 2023; Stermer et al., 1984).

55 of 120

(Continued)							
	 The pathogen overwinters in host tissues as both mycelium and spores. Conidia are produced 5 to 14 month infection, but their role in the disease transmission is considered not relevant, and ascospores are the masource of inoculum (EFSA PLH Panel, 2017; Ostry & Anderson, 2009; Ostry, 2013). Entoleuca mammata can spread over long distances via windborne ascospores, which are produced 2–3 year infection (Anderson et al., 1979b); cankers on felled trees on the ground continue to produce ascospores months (Ostry & Anderson, 2009). Ascospores are dispersed with a temperature above –4°C and wet wea a minimum of 16°C is required for starting germination, which became rapid at 28–32°C (EFSA PLH Panel Infected wood, mostly with bark, may be a pathway for passive spread of <i>E. mammata</i> in international trahowever, also young plants may carry ascospores or mycelium of the fungus, which can survive as a later infection on living material inadvertently moved (EFSA PLH Panel, 2017; EPPO, 2024c). Entoleuca mammata is considered an important pathogen of poplars in the USA and Canada, causing econo losses of millions of dollars a year (Anderson et al., 1979b; EFSA PLH Panel, 2017; Ostry, 2013). In Europe, d on <i>Populus tremula</i> has been reported in natural stands in France and Italy and in poplar plantations in S⁴ and Estonia (EFSA PLH Panel, 2017; Lutter et al., 2019); however, the pathogen is generally known as a per low importance (EFSA PLH Panel, 2023). 						
Symptoms	Main type of symptoms	 Symptoms of <i>E. mammata</i> infection have been described especially for <i>Populus</i> species. Early symptoms of cankers on the bark appear as slightly sunken, yellowish-orange areas with an irregular border. Young cankers can be easily identified by removing the bark to expose the white mycelium in the cambial zone. The outer bark in older cankers is then lifted into blister-like patches and break away, exposing blackened areas prominently visible on green branches and trunks. Callus formation only occasionally develops because cankers spread very quickly (Anderson et al., 1979b; EPPO, 2023). Wilting of leaves may be observed when living trees are girdled by cankers, as well as sprouting of new shoots on stem and branches. Infected trees can be secondarily colonised by other fungi, accelerating the host decline (EPPO, 2023). 					
	Presence of asymptomatic plants	The disease caused by <i>E. mammata</i> has a latent period and symptoms can appear only 2 years after the ascospore infection, therefore asymptomatic plants can be found (Ostry & Anderson, 2009).					
	Confusion with other pests	Some <i>Hypoxylon</i> species present in Europe on deciduous trees (<i>H. confluens</i> and <i>H. udum</i>) show symptoms similar to those of <i>E. mammata</i> but can be easily distinguished in laboratory by the ascospore characteristics (EFSA PLH Panel, 2017).					
Host plant range	 The list of hosts of <i>E. mammata</i> includes: <i>Alnus sinuata, Betula</i> sp., <i>Fagus</i> sp., <i>Malus</i> sp., <i>Ostrya</i> sp., <i>Populus adenopoda, P. alba, P. balsamifera, P. grandidentata, P. nigra, P. tremula, P. tremuloides, P. trichocarpa, P. × wettsteini, Populus hybrids, Salix caprea, S. cinerea, S. daphnoides, S. myrisinifolia, S. pentandra, S. phylicifolia, S. triandra, Salix,</i> sp. and <i>Sorbus aucuparia</i> (EFSA PLH Panel, 2023; EPPO, 2024c, 2024e; Ostry, 2013). In North America, <i>E. mammata</i> mainly infects the quacking aspen (<i>Populus tremuloides</i>); minor damage is recorded on <i>P. alleghaniensis, P. balsamifera, P. grandidentata</i> and various <i>Populus hybrids.</i> Other secondary hosts in North America are <i>Acer, Alnus, Betula, Carpinus, Fagus, Picea, Pyrus, Salix, Sorbus</i> and <i>Ulmus</i> (Manion & Griffin, 1986). In Europe, the main hosts are poplars, mostly <i>Populus tremula</i>; other hosts are <i>Populus alba, P. nigra, P. trichocarpa</i> and the hybrid <i>P. tremula× P. tremuloides</i> (Ostry, 2013). In central and northern Scandinavia, willows seem to be the main bosts of <i>E. mammata</i> mostly. <i>Soliv, Songar, S. neutandra</i> and S. <i>myrisinifolia</i> (Matthiacon, 1903). 						
Reported evidence of impact	Entoleuca mammata is an EU protected zone quarantine pest.						
Evidence that the commodity is a pathway	Plants for planting may carry ascospores and mycelium of <i>E. mammata</i> also as asymptomatic plants (EFSA PLH Panel, 2017; EPPO 2023) therefore the commodity is a pathway. <i>E. mammata</i> is believed to have been introduced in the last century into France with plant material (flowering branches of <i>Populus tremula</i>) used for hybridisation (EPPO, 1976).						
Surveillance information	<i>Entoleuca mammata</i> is not a reg (Dossier Section 2.0).	ulated pest for the UK and it is not under official control – limited in parts of the UK					

A.2.2 | Possibility of pest presence in the nursery

A.2.2.1 | Possibility of entry from the surrounding environment

Entoleuca mammata is present in the UK in England, Wales, Channel Islands and Scotland (CABI, 2019; EPPO, 2024b; Dossier Section 2.0). In Wales the pathogen was found on *Salix* sp. (Matthiassen, 1993).

Entoleuca mammata can easily spread with ascospores dispersed by air currents also over long distance and can infect *Acer campestre, A. pseudoplatanus* and *Populus* spp., which are present within 2 km from the nurseries in woodlands and hedgerows (Dossier Sections 1.1, 1.2 and 1.3). Other possible hosts, as *Betula* spp., *Fagus* spp., *Sorbus aucuparia* and *Salix* spp. might be present in the private gardens in the same area.

Uncertainties

- The presence of the pathogen on host plants in the surrounding area.

Taking into consideration the above evidence and uncertainties, the Panel considers that it is possible for *Entoleuca* mammata to enter the nurseries from surrounding environment via ascospores transported by wind and air currents.

A.2.2.2 | Possibility of entry with new plants/seeds

For all the *Populus* species of the Dossier the starting material is a mix of seeds, seedlings and cuttings, depending on the nursery. Seeds are certified and coming from the UK. Seedlings are either from the UK or the EU (including the Netherlands, Belgium and France) (Dossier Sections 1.1, 1.2 and 1.3).

None of the nurseries have mother plants of *Populus* or mother plants of other species, since no plants from grafting are produced (Dossier Sections 1.1, 1.2 and 1.3).

In addition to *Populus alba*, *P. nigra* and *P. tremula* plants, the nurseries also produce other plants (Dossier Sections 3.1, 3.2 and 3.3). Out of them, there are suitable hosts for the pathogen such as *Alnus* spp., *Fagus* spp., *Malus* spp., *Pyrus* spp., *Salix caprea*, *S. pentandra*, *Salix* spp., *Sorbus aucuparia* and *Ulmus* spp.. However, there is no information on how and where the plants are produced. Therefore, if the plants are first produced in another nursery, the pathogen could possibly travel with them.

The nurseries are using virgin peat or peat-free compost (a mixture of coir, tree bark, wood fibre, etc.) as a growing media (Dossier Sections 1.1, 1.2 and 1.3). The growing media is certified and heat-treated by commercial suppliers during production to eliminate pests and diseases. There is no evidence that soil or growing media may be a pathway for *E. mammata*.

Uncertainties

 No information is available on the provenance of new plants other than *Populus* used for plant production in the nurseries.

Taking into consideration the above evidence and uncertainties, the Panel considers that it is possible for the pathogen to enter the nurseries via new seedlings of *Populus* and plants of other species used for plant production in the area. The entry of the pathogen with seeds and the growing media the Panel considers as not possible.

A.2.2.3 | Possibility of spread within the nursery

Populus alba, P. nigra and *P. tremula* plants are either grown in containers (cells, pots, tubes, etc.) outdoors, in the open air or in field. Cell grown trees may be grown in greenhouses, however most plants will be field-grown, or field-grown in containers; there are no mother plants present in the nurseries (Dossier Sections 1.1, 1.2 and 1.3).

The pathogen can infect other suitable plants, such as *Alnus* spp., *Fagus* spp., *Malus* spp., *Salix* spp., *Sorbus* spp., etc. present within the nurseries (Dossier Sections 3.1, 3.2 and 3.3).

Once entered, ascospores of *E. mammata* could be produced on infected plants and naturally spread within the nurseries by air currents.

Uncertainties

- Whether ascospores are produced on infected nursery plants

Taking into consideration the above evidence and uncertainties, the Panel considers that the spread of the pathogen within the nurseries is possible by air currents.

A.2.3 | Information from interceptions

In the EUROPHYT/TRACES-NT database there are no records of notification of *Populus alba, P. nigra* and *P. tremula* plants for planting neither from the UK nor from other countries due to the presence of *E. mammata* between the years 1995 and September 2024 (EUROPHYT, 2024; TRACES-NT, 2024).

A.2.4 | Evaluation of the risk mitigation measures

In the table below, all risk mitigation measures currently applied in the UK are listed and an indication of their effectiveness on *E. mammata* is provided. The description of the risk mitigation measures currently applied in the UK is provided in the Table 7.

N	Risk mitigation measure	Effect on the pest	Evaluation and uncertainties
1	Registration of production sites	Yes	The risk mitigation measure is expected to be effective in reducing the likelihood of presence of the pathogen on the commodity. <u>Uncertainties</u> : – None
2	Physical separation	No	Not applicable.
3	Certified plant material	Yes	 The risk mitigation measure is expected to be effective in reducing the likelihood of presence of the pathogen on the commodity. <u>Uncertainties</u>: None
4	Growing media	No	Not applicable.
5	Surveillance, monitoring and sampling	Yes	 This measure could have some effect. <i>Entoleuca mammata</i> is not a regulated pest for Great Britain, and no specific measures on surveillance are taken. The pest has been a protected zone quarantine pest in Northern Ireland for many years and exports to North Ireland from other areas of the UK are checked in accordance with the requirements. <u>Uncertainties</u>: Whether plants are subjected to annual surveys
6	Hygiene measures	No	Not applicable.
7	Removal of infested plant material	Yes	This measure could have some effect. <u>Uncertainties</u> : – None
8	Irrigation water	No	Not applicable.
9	Application of pest control products	Yes	 Although little information exists on the efficacy of chemical treatments against <i>E. mammata</i> (Ostry, 2013), some of the fungicides used in the nursery targeting canker pathogens (Azoxystrobin, Pyrimethanil, Triazolinthione, Tebuconazole, Propamocarb Hydrochloride) could reduce the likelihood of the infection by the pathogen. <u>Uncertainties</u>: The level of efficacy of fungicides in reducing infection of <i>E. mammata</i>
10	Measures against soil pests	No	Not applicable.
11	Inspections and management of plants before export	Yes	 This measure could have some effect, although symptoms can appear only 2 years after the infection. <u>Uncertainties</u>: None
12	Separation during transport to the destination	No	Not applicable.

A.2.5 | Overall likelihood of pest freedom for cuttings/graftwood

A.2.5.1 | Reasoning for a scenario which would lead to a reasonably low number of infected cuttings/graftwood

The scenario assumes the pathogen to be absent or with a low pressure of the pathogen in the nurseries and in the surroundings. Younger plants are exposed to the pathogen for only short period of time. The scenario also assumes that symptoms of the disease are visible and promptly detected during inspections.

A.2.5.2 Reasoning for a scenario which would lead to a reasonably high number of infected cuttings/graftwood

The scenario assumes a high pressure of the pathogen in the nurseries and in the surroundings as suitable hosts are present. Older plants are exposed to the pathogen for longer period of time. The scenario also assumes that symptoms of the disease are not easily recognizable during inspections and that infections are asymptomatic.

A.2.5.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infected bundles of cuttings/graftwood (Median)

The scenario assumes a limited presence of the pathogen in the nurseries and the surroundings and that the plants are exposed to the pathogen for a sufficient period of time to cause infection through mechanical wounds. Poplars are suitable hosts.

A.2.5.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The limited information on the occurrence of the pathogen in the UK including the nurseries and the surroundings results in high level of uncertainties for infection rates below the median. Otherwise, the pest pressure from the surroundings is expected to be low giving less uncertainties for rates above the median. The young age of plants would also leave less uncertainty for estimates above the median.

A.2.5.5 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on cuttings/graftwood of *Populus nigra*

The following Tables show the elicited and fitted values for pest infection (Table A.7) and pest freedom (Table A.8).

TABLE A.7	Elicited and fitted valu	es of the uncertainty	distribution of	pest infestation by	y Entoleuca mammata pe	r 10,000 plants/bundles
-----------	--------------------------	-----------------------	-----------------	---------------------	------------------------	-------------------------

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5 %	99%
Elicited values	0					8		16		30					70
EKE	0.405	0.912	1.70	3.20	5.19	7.72	10.4	16.5	24.4	29.5	36.1	43.8	53.0	61.0	70.2

Note: The EKE results is the BetaGeneral (1.1421, 5.5388, 0, 120) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants/bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.8.

TABLE A.8 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants/bundles calculated by Table A.7

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5%	99 %
Values	9930					9970		9984		9992					10,000
EKE results	9930	9939	9947	9956	9964	9971	9976	9983	9990	9992	9995	9997	9998	9999.1	9999.6









Pestfree plants/bundles [number out of 10,000]

FIGURE A.4 (Continued)



FIGURE A.4 (A) Elicited uncertainty of pest infection per 10,000 plants/bundles (histogram in blue– vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest free plants/bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants/bundles.

A.2.5.6 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on cuttings/graftwood of *Populus tremula*

The following Tables show the elicited and fitted values for pest infection (Table A.9) and pest freedom (Table A.10).

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95%	97.5%	99 %
Elicited values	0					16		32		60					140
EKE	0.810	1.82	3.39	6.39	10.4	15.4	20.8	33.1	48.8	59.0	72.2	87.6	106	122	140

TABLE A.9 Elicited and fitted values of the uncertainty distribution of pest infestation by Entoleuca mammata per 10,000 plants/bundles

Note: The EKE results is the BetaGeneral (1.1421, 5.5388, 0, 240) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants/bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.10.

TABLE A.10 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants/bundles calculated by Table A.9

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95 %	97.5%	99 %
Values	9860					9940		9968		9984					10000
EKE results	9860	9878	9894	9912	9928	9941	9951	9967	9979	9985	9990	9994	9997	9998	9999









Pestfree plants/bundles [number out of 10,000]

FIGURE A.5 (Continued)



FIGURE A.5 (A) Elicited uncertainty of pest infection per 10,000 plants/bundles (histogram in blue–vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest free plants/bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants/bundles.

A.2.6 | Overall likelihood of pest freedom for bare root plants

A.2.6.1 | Reasoning for a scenario which would lead to a reasonably low number of infected bare root plants

The scenario assumes the pest to be absent or with a low pressure in the nurseries and in the surroundings. Younger plants are exposed to the pathogen for only a short period of time. The scenario also assumes that symptoms of the disease are visible and promptly detected during inspections.

A.2.6.2 | Reasoning for a scenario which would lead to a reasonably high number of infected bare root plants

The scenario assumes a high pressure of the pathogen in the nurseries and in the surroundings as suitable hosts are present. Older plants are exposed to the pathogen for a longer period of time. The scenario also assumes that symptoms of the disease are not easily recognisable during inspections and that infections are asymptomatic.

A.2.6.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infected bundles of whips and seedlings (Median)

The scenario assumes a limited presence of the pathogen in the nurseries and the surroundings and that the plants are exposed to the pathogen for a sufficient period of time to cause infection through mechanical wounds. Poplars are suitable hosts.

A.2.6.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The limited information on occurrence of the pathogen in the UK including the nurseries and the surroundings results in high level of uncertainties for infection rates below the median. Otherwise, the pest pressure from the surroundings is expected to be low giving less uncertainties for rates above the median.

68 of 120

A.2.6.5 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on bare root plants of *Populus alba* and *Populus nigra*

The following Tables show the elicited and fitted values for pest infection (Table A.11) and pest freedom (Table A.12).

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5%	99 %
Elicited values	0					15		29		50					120
EKE	1.25	2.45	4.11	7.00	10.6	14.9	19.3	29.2	41.7	49.8	60.5	73.1	88.8	103	120

TABLE A.11 Elicited and fitted values of the uncertainty distribution of pest infestation by Entoleuca mammata per 10,000 plants/bundles.

Note: The EKE results is the BetaGeneral (1.3991, 10.013, 0, 290) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants/bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.12.

TABLE A.12 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants/bundles calculated by Table A.11.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5 %	99 %
Values	9880					9950		9971		9985					10,000
EKE results	9880	9897	9911	9927	9940	9950	9958	9971	9981	9985	9989	9993	9996	9998	9999







Pestfree plants/bundles [number out of 10,000]

FIGURE A.6 (Continued)



FIGURE A.6 (A) Elicited uncertainty of pest infection per 10,000 plants/bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free bare root plants/bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants/bundles.

72 of 120

A.2.6.6 | Elicitation outcomes of the assessment of the pest freedom for Entoleuca mammata on bare root plants of Populus tremula

The following Tables show the elicited and fitted values for pest infection (Table A.13) and pest freedom (Table A.14).

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95%	97.5%	99 %
Elicited values	0					30		58		100					240
EKE	2.51	4.90	8.22	14.0	21.1	29.8	38.7	58.4	83.3	99.6	121	146	178	206	241

TABLE A.13 Elicited and fitted values of the uncertainty distribution of pest infestation by Entoleuca mammata per 10,000 plants/bundles.

Note: The EKE results is the BetaGeneral (1.3991, 10.013, 0, 580) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants/bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.14.

TABLE A.14 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants/bundles calculated by Table A.13.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95 %	97.5 %	99 %
Values	9760					9900		9942		9970					10,000
EKE results	9759	9794	9822	9854	9879	9900	9917	9942	9961	9970	9979	9986	9992	9995	9997


FIGURE A.7 (Continued)



Pestfree plants/bundles [number out of 10,000]

FIGURE A.7 (Continued)



FIGURE A.7 (A) Elicited uncertainty of pest infection per 10,000 plants/bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free bare root plants/bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants/bundles.

A.2.7 | Overall likelihood of pest freedom for cell grown plants

A.2.7.1 | Reasoning for a scenario which would lead to a reasonably low number of infected cell grown plants

The scenario assumes the pest to be absent or with a low pressure in the nurseries and in the surroundings. Younger plants are exposed to the pathogen for only a short period of time. The scenario also assumes that symptoms of the disease are visible and promptly detected during inspections.

A.2.7.2 | Reasoning for a scenario which would lead to a reasonably high number of infected cell grown plants

The scenario assumes a high pressure of the pathogen in the nurseries and in the surroundings as suitable hosts are present. Older plants are exposed to the pathogen for a longer period of time. Cell grown plants are in close proximity to each other, which increases the humidity and hence provides good growth conditions for *E. mammata*. The scenario also assumes that symptoms of the disease are not easily recognisable during inspections and that infections are asymptomatic.

A.2.7.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infected bundles of whips and seedlings (Median)

The scenario assumes a limited presence of the pathogen in the nurseries and the surroundings and that the plants are exposed to the pathogen for a sufficient period of time to cause infection through mechanical wounds. Plants are very young and therefore they display a limited susceptibility to the pathogen. Poplars are suitable hosts.

A.2.7.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The limited information on occurrence of the pathogen in the UK including the nurseries and the surroundings results in high level of uncertainties for infection rates below the median. Otherwise, the pest pressure from the surroundings is expected to be low giving less uncertainties for rates above the median. The young age of plants would also leave less uncertainty for estimates above the median.

A.2.7.5 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on cell grown plants of *Populus alba* and *Populus nigra*

The following Tables show the elicited and fitted values for pest infection (Table A.15) and pest freedom (Table A.16).

TABLE A.15	Elicited and fitted values of the uncertaint	y distribution of pest infestation l	by <i>Entoleuca mammata</i> per 10,000 bundles.
------------	--	--------------------------------------	---

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95 %	97.5%	99 %
Elicited values	0					10		20		40					90
EKE	0.351	0.873	1.75	3.54	6.04	9.34	12.9	21.2	32.0	38.9	47.8	57.9	69.6	79.4	90.0

Note: The EKE results is the BetaGeneral (1.0126, 3.9819, 0, 131) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.16.

TABLE A.16 The uncertainty distribution of plants free of *Entoleuca mammata* per 10,000 bundles calculated by Table A.15.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95 %	97.5%	99 %
Values	9910					9960		9980		9990					10,000
EKE results	9910	9921	9930	9942	9952	9961	9968	9979	9987	9991	9994	9996	9998	9999.1	9999.6

Note: The EKE results are the fitted values.







Pestfree bundles [number out of 10,000]

FIGURE A.8 (Continued)



FIGURE A.8 (A) Elicited uncertainty of pest infection per 10,000 bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 bundles.

A.2.7.6 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on cell grown plants of *Populus tremula*

The following Tables show the elicited and fitted values for pest infection (Table A.17) and pest freedom (Table A.18).

TABLE A.17	Elicited and fitted values of the uncertain	y distribution of pest infestation b	y Entoleuca mammata per 10,000 bundles.
------------	---	--------------------------------------	---

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90%	95%	97.5%	99 %
Elicited values	0					20		40		80					180
EKE	0.702	1.75	3.50	7.07	12.1	18.7	25.8	42.5	63.9	77.7	95.6	116	139	159	180

Note: The EKE results is the BetaGeneral (1.0126, 3.9819, 0, 262) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected bundles per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.18.

TABLE A.18 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 bundles calculated by Table A.17.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67%	75%	83%	90%	95%	97.5 %	99 %
Values	9820					9920		9960		9980					10,000
EKE results	9820	9841	9861	9884	9904	9922	9936	9958	9974	9981	9988	9993	9997	9998	9999

Note: The EKE results are the fitted values.







Pestfree bundles [number out of 10,000]

FIGURE A.9 (Continued)



FIGURE A.9 (A) Elicited uncertainty of pest infection per 10,000 bundles (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest free bundles per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 bundles.

A.2.8 | Overall likelihood of pest freedom for plants in pots

A.2.8.1 | Reasoning for a scenario which would lead to a reasonably low number of infected plants in pots

The scenario assumes the pest to be absent or with a low pressure in the nurseries and in the surroundings. Younger plants are exposed to the pathogen for only a short period of time. The scenario also assumes that symptoms of the disease are visible and promptly detected during inspections.

A.2.8.2 | Reasoning for a scenario which would lead to a reasonably high number of infected plants in pots

The scenario assumes a high pressure of the pathogen in the nurseries and in the surroundings as suitable hosts are present. Older plants are exposed to the pathogen for a longer period of time. The scenario also assumes that symptoms of the disease are not easily recognisable during inspections and that infections are asymptomatic.

A.2.8.3 | Reasoning for a central scenario equally likely to over- or underestimate the number of infected plants in pots (Median)

The scenario assumes a limited presence of the pathogen in the nurseries and the surroundings and that the plants are exposed to the pathogen for a sufficient period of time to cause infection through mechanical wounds. Poplars are suitable hosts.

A.2.8.4 | Reasoning for the precision of the judgement describing the remaining uncertainties (1st and 3rd quartile/interquartile range)

The limited information on occurrence of the pathogen in the UK including the nurseries and the surroundings results in high level of uncertainties for infection rates below the median. Otherwise, the pest pressure from the surroundings is expected to be low giving less uncertainties for rates above the median.

A.2.8.5 | Elicitation outcomes of the assessment of the pest freedom for *Entoleuca mammata* on plants in pots of *Populus alba* and *Populus nigra*

The following Tables show the elicited and fitted values for pest infection (Table A.19) and pest freedom (Table A.20).

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67%	75%	83%	90 %	95%	97.5%	99 %
Elicited values	0					20		39		80					170
EKE	0.604	1.56	3.22	6.69	11.6	18.2	25.4	42.2	63.6	77.3	94.6	114	135	153	171

TABLE A.19 Elicited and fitted values of the uncertainty distribution of pest infestation by Entoleuca mammata per 10,000 plants.

Note: The EKE results is the BetaGeneral (0.96971, 3.2104, 0, 225) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.20.

TABLE A.20 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants calculated by Table A.19.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67%	75%	83%	90%	95 %	97.5 %	99 %
Values	9830					9920		9961		9980					10,000
EKE results	9829	9847	9865	9886	9905	9923	9936	9958	9975	9982	9988	9993	9997	9998	9999

Note: The EKE results are the fitted values.





87 of 120



Pestfree plants [number out of 10,000]



FIGURE A.10 (Continued)



FIGURE A.10 (A) Elicited uncertainty of pest infection per 10,000 plants (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free plants per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants.

90 of 120

A.2.8.6 | Elicitation outcomes of the assessment of the pest freedom for Entoleuca mammata on plants in pots of Populus tremula

The following Tables show the elicited and fitted values for pest infection (Table A.21) and pest freedom (Table A.22).

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67%	75%	83%	90%	95%	97.5%	99 %
Elicited values	0					40		78		160					340
EKE	1.21	3.12	6.43	13.4	23.3	36.5	50.9	84.4	127	155	189	227	270	305	341

TABLE A.21 Elicited and fitted values of the uncertainty distribution of pest infestation by Entoleuca mammata per 10,000 plants.

Note: The EKE results is the BetaGeneral (0.96971, 3.2104, 0, 450) distribution fitted with @Risk version 7.6.

Based on the numbers of estimated infected bundles the pest freedom was calculated (i.e. = 10,000 – number of infected plants per 10,000). The fitted values of the uncertainty distribution of the pest freedom are shown in Table A.22.

TABLE A.22 The uncertainty distribution of plants free of Entoleuca mammata per 10,000 plants calculated by Table A.21.

Percentile	1%	2.5%	5%	10%	17%	25%	33%	50%	67 %	75%	83%	90 %	95 %	97.5%	99 %
Values	9660					9840		9922		9960					10,000
EKE results	9659	9695	9730	9773	9811	9845	9873	9916	9949	9964	9977	9987	9994	9997	9999

Note: The EKE results are the fitted values.







Pestfree plants [number out of 10,000]

FIGURE A.11 (Continued)



FIGURE A.11 (A) Elicited uncertainty of pest infection per 10,000 plants (histogram in blue – vertical blue line indicates the elicited percentile in the following order: 1%, 25%, 50%, 75%, 99%) and distributional fit (red line); (B) uncertainty of the proportion of pest-free plants per 10,000 (i.e. = 1 – pest infection proportion expressed as percentage); (C) descending uncertainty distribution function of pest infection per 10,000 plants.

A.2.9 | References

- Anderson, N. A., Ostry, M. E., and Anderson, G. W. (1979a). Insect wounds as infection sites for *Hypoxylon mammatum* on trembling aspen. *Phytopathology*, 69, 476–479. https://doi.org/10.1094/phyto-69-476
- Anderson, R. L., Anderson, G. W., and Schipper, A. L. Jr. (1979b). Hypoxylon canker of aspen. USDA Forest Insect and Disease Leaflet, 6, 6.

CABI (Centre for Agriculture and Bioscience International). (2019). *Hypoxylon mammatum* (poplar canker). https://www.cabi.org/cpc/datasheet/28323 (accessed: 2024/10/20).

- DEFRA (Department for Environment, Food and Rural Affairs). (2023). UK risk register details for *Entoleuca mammata*. https://planthealthportal.defra.gov.uk/pests-and-diseases/uk-plant-health-risk-register/viewPestRisks.cfm?cslref=11840 (accessed: 2024/10/20).
- EFSA PLH Panel (EFSA Panel on Plant Health), Jeger, M., Bragard, C., Caffier, D., Candresse, T., Chatzivassiliou, E., Dehnen Schmutz, K., Gilioli, G., Gregoire, J.-C., Jaques Miret, J. A., MacLeod, A., Navajas Navarro, M., Niere, B., Parnell, S., Potting, R., Rafoss, T., Rossi, V., Urek, G., Van Bruggen, A., Van der Werf, W., West, J., Winter, S., Boberg, J., Gonthier, P., and Pautasso, M. (2017). Scientific Opinion on the pest categorisation of *Entoleuca mammata*. *EFSA Journal*, 15(7), 4925. https://doi.org/10.2903/j.efsa.2017.4925
- EFSA PLH Panel (EFSA Panel on Plant Health), Bragard, C., Baptista, P., Chatzivassiliou, E., Di Serio, F., Jaques Miret, J. A., Justesen, A. F., MacLeod, A., Magnusson, C. S., Milonas, P., Navas-Cortes, J. A., Parnell, S., Potting, R., Reignault, P. L., Stefani, E., Thulke, H.-H., Van der Werf, W., Vicent Civera, A., Yuen, J., Zappalà, L., Battisti, A., Mas, H., Rigling, D., Faccoli, M., Gardi, C., Iacopetti, G., Mikulova, A., Mosbach-Schulz, O., Stergulc, F., Streissl, F., & Gonthier, P. (2023). Commodity risk assessment of *Acer campestre* plants from the UK. *EFSA Journal*, *21*(7), 8071. https://doi.org/10.2903/j.efsa.2023. 8071
- EPPO (European and Mediterranean Plant Protection Organization). (1976). Outbreak in France of Aspen Canker caused by *Hypoxylon mammatum* (Wahl) Mill. (= *H. pruinatum* (Klotz) Cke). https://gd.eppo.int/reporting/article-5822 (accessed: 2024/10/22).
- EPPO (European and Mediterranean Plant Protection Organization). (2023). Datasheets on Quarantine Pests: Entoleuca mammata (HYPOMA), Datasheet. https://gd.eppo.int/taxon/HYPOMA/datasheet (accessed: 2024/10/21).
- EPPO (European and Mediterranean Plant Protection Organization). (2024a). Entoleuca mammata (HYPOMA), Categorization. https://gd.eppo.int/taxon/ HYPOMA/categorization (accessed: 2024/10/20).
- EPPO (European and Mediterranean Plant Protection Organization). (2024b). Entoleuca mammata (HYPOMA), Distribution. https://gd.eppo.int/taxon/ HYPOMA/distribution (accessed: 2024/10/20).
- EPPO (European and Mediterranean Plant Protection Organization). (2024c). Entoleuca mammata (HYPOMA), Hosts. https://gd.eppo.int/taxon/ HYPOMA/hosts (accessed: 2024/10/20).
- EUROPHYT (European Union Notification System for Plant Health Interceptions). (2024). http://ec.europa.eu/food/plant/plant_health_biosecurity/ europhyt/index_en.htm (accessed: 2024-10-24).
- Granmo, A., Laessoe, T., and Schumacher, T. (1999). The genus Nemania s.l. (Xylariaceae) in Norden. Sommerfeltia, 27, 1–96. https://doi.org/10.2478/ som-1999-0002
- Kasanen, R., Hantula, J., Ostry, M. E., Pinon, J., and Kurkela, T. (2004). North American populations of *Entoleuca mammata* are genetically more variable than populations in Europe. *Mycological Research*, *108*, 766–774. https://doi.org/10.1017/s0953756204000334
- Klejdysz, T., Kubasik, W., Strażyński, P., Gawlak, M., Pruciak, A., Rzepecka, D., and Kałuski, T. (2018). Express pest risk analysis for *Hypoxylon mammatum*. https://www.plantguarantine.pl/pl/?node_id=1683&literka=E (accessed: 2024/10/20).
- Lee, Y. S., Han, S. S., Shin, J. H., Lee, Y. M., and Song, B. K. (2000). Germ tube formation of ascospores of two terrestrial higher ascomycetes, *Hypoxylon* mammatum and *H. truncatum. Journal of Korean Wood Science and Technology*, 28, 10–16.
- Lutter, R., Drenkhan, R., Tullus, A., Jürimaa, K., Tullus, T., and Tullus, H. (2019). First record of *Entoleuca mammata* in hybrid aspen plantations in hemiboreal Estonia and stand-environmental factors affecting its prevalence. *European Journal of Forest Research*, 138(2), 263–274. https://doi.org/10.1007/ s10342-019-01165-7
- Manion, P. D., and Griffin, D. H. (1986). Sixty-five years of research on Hypoxylon canker of aspen. *Plant Disease*, 70(8), 803–808. https://doi.org/10.1094/pd-70-803
- Mathiassen, G. (1993). Corticolous and lignicolous Pyrenomycetes s. lat. (Ascomycetes) on Salix along a mid-Scandinavian transect. Sommerfeltia, 20, 1–180. https://doi.org/10.2478/som-1993-0006
- NBIC (Norwegian Biodiversity Information Center). (2021). Entoleuca mammata. https://artsdatabanken.no/Taxon/Entoleuca%20mammata/82864 (accessed: 2024/10/21).
- Ostry, M. E. (2013). Hypoxylon canker. In: P. Gonthier, and G. Nicolotti (eds.), Infectious Forest diseases (pp. 407–419). CABI International, Wallingford. Ostry, M. E., and Anderson, N. A. (2009). Genetics and ecology of the *Entoleuca mammata–Populus* pathosystem: Implications for aspen improvement
- and management. Forest Ecology and Management, 257, 390–400. https://doi.org/10.1016/j.foreco.2008.09.053
- Stermer, B. A., Scheffer, R. P., and Hart, J. H. (1984). Isolation of toxins from *Hypoxylon mammatum* and demonstration of some toxin effects on selected clones of *Populus tremuloides*. *Phytopathology*, *74*, 654–658. https://doi.org/10.1094/phyto-74-654
- TRACES-NT (TRAde Control and Expert System). (2024). https://webgate.ec.europa.eu/tracesnt (accessed: 2024-10-24).

APPENDIX B

Web of Science All Databases Search String

In the Table B.1, the search string for Populus alba used in Web of Science is reported. Totally, 653 papers were retrieved. Titles and abstracts were screened, and 64 pests were added to the list of pests (see Appendix F).

In the Table B.2, the search string for Populus nigra used in Web of Science is reported. Totally, 1230 papers were retrieved. Titles and abstracts were screened, and 60 pests were added to the list of pests (see Appendix F).

In the Table B.3, the search string for Populus tremula used in Web of Science is reported. Totally, 707 papers were retrieved. Titles and abstracts were screened, and 145 pests were added to the list of pests (see Appendix F).

TABLE B.1 String for Populus alba.

Web of Science All databases	TOPIC: "Populus alba" OR "P. alba" OR "abele" OR "silver-leaved poplar" OR "white poplar" OR "Bolle's poplar" OR "Leuce alba" OR "Populus bolleana" OR "Populus nivea"
	AND TOPIC: pathogen* OR pathogenic bacteria OR fung* OR oomycet* OR myce* OR bacteri* OR virus* OR viroid* OR insect\$ OR mite\$ OR phytoplasm* OR arthropod* OR nematod* OR disease\$ OR infecti* OR damag* OR symptom* OR pest\$ OR vector OR hostplant\$ OR "host plant\$" OR host OR "root lesion\$" OR decline\$ OR infestation\$ OR damage\$ OR symptom\$ OR dieback* OR "die back*" OR "malaise" OR aphid\$ OR curculio OR thrip\$ OR cicad\$ OR miner\$ OR borer\$ OR weevil\$ OR "plant bug\$" OR spittlebug\$ OR moth\$ OR mealybug\$ OR cutworm\$ OR pillbug\$ OR "root feeder\$" OR caterpillar\$ OR "foliar feeder\$" OR virosis OR viroses OR blight\$ OR wilt\$ OR wiltd OR canker OR scab\$ OR rot OR rots OR rotten OR "damping off" OR "damping-off" OR blister\$ OR "smut" OR mould OR mold OR "damping syndrome\$" OR mildew OR scald\$ OR "root knot" OR "root feeding" OR "root\$feeding" NOT
	 NOT TOPIC: "winged seeds" OR metabolites OR "tannins OR climate OR "maple syrup" OR syrup OR mycorrhiz" OR "carbon loss" OR pollut* OR weather OR propert* OR probes OR spectr* OR antioxidant\$ OR transformation OR RNA OR DNA OR "Secondary plant metabolite\$" OR metabol* OR "Phenolic compounds" OR Quality OR Abiotic OR Storage OR Pollen* OR fertil* OR Mulching OR Nutrient* OR Pruning OR drought OR "human virus" OR "animal disease*" OR "plant extracts" OR immunological OR "purified fraction" OR "traditional medicine" OR medicine OR mammal* OR bird* OR "human disease*" OR biomarker\$ OR "health education" OR bat\$ OR "seedling\$ survival" OR "anthropogenic disturbance" OR "cold resistance" OR "salt stress" OR salinity OR "aCER method" OR "adaptive cognitive emotion regulation" OR nitrogen OR hygien* OR "cognitive function\$" OR fossil\$ OR *toxicity OR Miocene OR postglacial OR "weed control" OR landscape NOT TOPIC: 'Agrilus horni" OR "Anoplophora chinensis" OR "Apriona cinerea" OR "Apriona germari" OR "Chondrostereum purpureum" OR "Choristoneura conflictana" OR "Orgyia leucostigma" OR "Entoleuca mammata" OR "Euwallacea fornicatus sensu lato" OR "Euwallacea fornicatus sensu stricto" OR "Lopholeucaspis japonica" OR "Lycorma delicatula" OR "Megaplatypus mutatus" OR "Oemona hirta" OR "Orgyia leucostigma" OR "Chondrostereum purpureum" OR "Chorabereum purpureum" OR "Karias vermana" OR "Gypsonoma aceriana" OR "Lymantria dispar asiatica" OR "Lymantria obfuscata" OR "Melampsora populnea" OR "Paranthrene tabaniformis" OR "Phloeomyzus passerinii" OR "Rhizobium radiobacter" OR "Rhizobium rhizogenes" OR "Saperda populnea" OR "Tremex fusciornis" OR "Triachys sartus" OR "Melampsora populnea" OR "Paperda populnea" OR "Chordosportera littoralis" OR "Longidorus euonymus" OR "Cyptorhynchus lapathi" OR "Diplodia seriata" OR "Armillaria gallica" OR "Armillaria mellea" OR "Armillaria tabescens" OR "Chyptorhynchus cactorum" OR "Saperda cacharias" OR "Spodoptera littoralis" OR "Longidorus euo
	warshavensis" OR "Pemphigus bursarius" OR "Pemphigus immunis" OR "Pemphigus protospirae" OR "Pemphigus vesicarius" OR "Phloeomyzus passerinii" OR "Phylloxerina populi" OR "Pterocomma anyangense" OR "Pterocomma dubium" OR "Pterocomma populeum" OR "Pterocomma sinipopulifoliae" OR "Pterocomma smithiae" OR "Pterocomma yezoense" OR "Stomaphis longirostris" OR "Tuberolachnus salignus" OR "Eriophyes populi" OR "Byctiscus betulae" OR "Byctiscus populi" OR "Saperda carcharias" OR "Saperda populnea" OR "Saperda scalaris" OR "Zeugophora subspinosa" OR "Rutidosoma globulus" OR "Cryptorhynchus lapathi" OR "Dorytomus dejeani" OR "Dorytomus filirostris" OR "Dorytomus longimanus" OR "Dorytomus tremulae" OR "Ellescus scanicus" OR "Ramphus pulicarius" OR "Ahynchaenus stigma" OR "Phyllobius pyri" OR "Agromyza albitarsis" OR "Hexomyza schineri" OR "Contarinia petioli" OR "Dasineura populeti" OR "Harmandia loewi" OR "Harmandiola cavernosa" OR "Harmandiola globuli" OR "Harmandiola tremulae" OR "Helicomyia saliciperda" OR "Lasioptera populnea" OR "Rabdophaga giraudiana" OR "Rabdophaga saliciperda" OR "Edwardsiana candidula" OR "Idiocerus albicans" OR "Idiocerus confusus" OR "Idiocerus distinguendus" OR "Idiocerus populi" OR "Idiocerus tremulae" OR "Kybos populi" OR "Kybos strigilifer" OR "Lygocoris populi" OR "Sthenarus rotermundi" OR "Choinaspis salicis" OR "Lepidosaphes ulmi" OR "Pseudaulacaspis pentagona" OR "Chaitophorus albus" OR "Chaitophorus populeti" OR "Phloeomyzus passerinii" OR "Phylloxerina populi" OR "Camarotoscena speciosa" OR "Messa glaucopis" OR "Nymphalis polychloros" OR "Lobophora halterata" OR "Peerostoma palpina" OR "Leucoma salicis" OR "Acronicta leporina" OR "Acronicta megacephala" OR "Catocala fraxini" OR "Pterostoma palpina" OR "Laothoe populi" OR "Tethea ocularis" OR "Gelechia nigra" OR "Phyllonorycter comparella" OR "Batrachedra praeangusta" OR "Ectoedemia turbidella" OR "Stigmella assinilella" OR "Sciota adelphella"
	comparella" OR "Batrachedra praeangusta" OR "Ectoedemia turbidella" OR "Stigmella assimilella" OR "Sciota adelphella" (Continues)

OR "Gypsonoma oppressana" OR "Acleris rufana" OR "Acleris hastiana" OR "Trypophloeus granulatus" OR "Aonidiella orientalis" OR "Aspidiotus nerii" OR "Ceroplastes rusci" OR "Chionaspis salicis" OR "Chrysomphalus dictyospermi" OR "Coccus hesperidum hesperidum" OR "Comstockaspis perniciosa" OR "Diaspidiotus ancylus" OR "Diaspidiotus armenicus" OR "Diaspidiotus caucasicus" OR "Diaspidiotus gigas" OR "Diaspidiotus kaussarii" OR "Diaspidiotus lenticularis" OR "Diaspidiotus ostreaeformis" OR "Diaspidiotus slavonicus" OR "Diaspidiotus transcaspiensis" OR "Eulecanium ciliatum" OR "Eulecanium douglasi" OR "Eulecanium tiliae" OR "Hemiberlesia lataniae" OR "Hemiberlesia rapax" OR "Lepidosaphes ulmi" OR "Lopholeucaspis japonica" OR "Newsteadia floccosa" OR "Oceanaspidiotus spinosus" OR "Pseudaulacaspis pentagona" OR "Pulvinaria costata" OR "Pulvinaria loralaiensis" OR "Pulvinaria occidentalis" OR "Salicicola kermanensis" OR "Heterodera salixophila" OR "Trichodorus primitivus" OR "Meloidogyne javanica" OR "Paranthrene tabaniformis" OR "Perisomena caecigena" OR "Phalanta phalantha" OR "Phyllonorycter chiclanella" OR "Phyllonorycter comparella" OR "Phyllonorycter populiella" OR "Phyllonorycter scudderella" OR "Pyralis pictalis" OR "Sciota adelphella" OR "Sesia flavicollis" OR "Sesia tibialis" OR "Sparganothis pettitana" OR "Spiramater lutra" OR "Stigmella assimilella" OR "Stigmella trimaculella" OR "Synanthedon melliniformis" OR "Synaxis jubararia" OR "Thyridopteryx ephemeraeformis" OR "Yponomeuta gigas" OR "Yponomeuta rorrella" OR "Anacampsis innocuella" OR "Anacampsis niveopulvella" OR "Gelechia nigra" OR "Odontopera bidentata" OR "Caloptilia stigmatella" OR "Cameraria obliquifascia" OR "Malacosoma disstria" OR "Malacosoma californica" OR "Malacosoma incurva" OR "Hyphantria cunea" OR "Leucoma salicis" OR "Lymantria dispar" OR "Orgyia leucostigma" OR "Paraleucoptera albella" OR "Paraleucoptera sinuella" OR "Ectoedemia klimeschi" OR "Ectoedemia turbidella" OR "Acronicta leporina" OR "Acronicta megacephala" OR "Catocala relicta" OR "Earias vernana" OR "Cerura vinula" OR "Clostera curtula" OR "Clostera pigra" OR "Furcula furcula" OR "Notodonta ziczac" OR "Apatura ilia" OR "Basilarchia arthemis" OR "Basilarchia archippus" OR "Limenitis populi" OR "Nymphalis polychloros" OR "Nymphalis antiopa" OR "Paranthrene diaphana" OR "Laothoe populi" OR "Paonias excaecata" OR "Acleris hastiana" OR "Acleris fuscana" OR "Acleris rufana" OR "Choristoneura conflictana" OR "Evora hemidesma" OR "Gypsonoma oppressana" OR "Pandemis limitata" OR "Amphitetranychus viennensis" OR "Eotetranychus albus" OR "Eotetranychus edi" OR "Eotetranychus populi" OR "Tetranychus urticae" OR "Agromyza albitarsis" OR "Caloptilia stigmatella" OR "Fenusella glaucopis" OR "Gypsonoma oppressana" OR "Phyllonorycter comparella" OR "Phyllonorycter salictella" OR "Stigmella assimilella" OR "Stigmella trimaculella" OR "Drepanopeziza populi-albae" OR "Chondrostereum purpureum" OR "Marssonina brunnea" OR "Marssonina castagnei" OR "Phoma exigua" OR "Rigidoporus vinctus" OR "Pseudomonas syringae" OR "Melampsora rostrupii" OR "Melampsora laricispopulina" OR "Strossmayeria basitricha" OR "Cytospora paratranslucens" OR "Drepanopeziza tremulae" OR "Antrodia malicola" OR "Trametes zonata" OR "Oemona hirta" OR "Agrocybe aegerita" OR "Alatospora acuminata" OR "Alternaria alternata" OR "Alternaria scrophulariae" OR "Apioplagiostoma populi" OR "Aporpium caryae" OR "Armillaria mellea" OR "Asteroma frondicola" OR "Asteromella osteospora" OR "Botrytis cinerea" OR "Capnodium salicinum" OR "Cercospora populina" OR "Chondrostereum purpureum" OR "Ciboria poronioides" OR "Cladosporium aphidis" OR "Cladosporium brunneum" OR "Cladosporium epiphyllum" OR "Cladosporium fumago" OR "Cladosporium herbarum" OR "Cladosporium herbarum f. hormodendroides" OR "Cladosporium martianoffianum" OR "Clavariopsis aquatica" OR "Colletotrichum gloeosporioides" OR "Coniothecium applanatum" OR "Coryneum populinum" OR "Cryptosphaeria multicontinentalis" OR "Cryptosphaeria pullmanensis" OR "Cytospora chrysosperma" OR "Cytospora fugax" OR "Cytospora nivea" OR "Cytospora paratranslucens" OR "Cytospora translucens" OR "Diaporthe medusaea" OR "Dicoccum populinum" OR "Didymosphaeria populina" OR "Diplodia gongrogena" OR "Diplodia malorum" OR "Diplodia mutila" OR "Diplodia populina" OR "Discella populina" OR "Discosporium populeum" OR "Dothichiza populea" OR "Dothiorella populina" OR "Drepanopeziza castagnei" OR "Drepanopeziza populi-albae" OR "Drepanopeziza populorum" OR "Drepanopeziza tremulae" OR "Erysiphe adunca" OR "Eutypa leptoplaca" OR "Eutypa populina" OR "Eutypa sparsa" OR "Flammula argentina" OR "Fomes applanatus" OR "Fomes connatus" OR "Fomes fomentarius" OR "Fomes fomentarius subsp. fomentarius" OR "Fomes igniarius" OR "Fusarium oxysporum f. sp. tracheiphilum" OR "Fusicladium martianoffianum" OR "Fusicladium radiosum" OR "Fusicladium radiosum var. lethiferum" OR "Fusicladium radiosum var. populi-albae" OR "Fusicladium radiosum var. radiosum" OR "Fusicladium romellianum" OR "Ganoderma applanatum" OR "Ganoderma lucidum" OR "Gloeosporium castagnei" OR "Gloeosporium populi-albae" OR "Gloeosporium tremulae" OR "Glomus fasciculatum" OR "Glomus geosporum" OR "Helicobasidium mompa" OR "Helicobasidium purpureum" OR "Helicoma perelegans" OR "Helicosporium griseum" OR "Helvella pityophila" OR "Hendersonula toruloidea" OR "Hyphodiscus gemmarum" OR "Hyphodontia sambuci" OR "Hypoxylon laschii" OR "Irpex zonatus" OR "Laetiporus sulphureus" OR "Leptosphaeria aegira" OR "Leptosphaeria alcides" OR "Leucostoma niveum" OR "Lophiostoma myriocarpum" OR "Marssonia piriformis" OR "Marssonina brunnea" OR "Marssonina castagnei" OR "Marssonina piriformis" OR "Marssonina populi" OR "Melampsora abietis-canadensis" OR "Melampsora aecidioides" OR "Melampsora allii-populina" OR "Melampsora castellana" OR "Melampsora larici-tremulae" OR "Melampsora laricis" OR "Melampsora laricis-populina" OR "Melampsora magnusiana" OR "Melampsora magnusiana f. tomentosea" OR "Melampsora mercurialis-tremulae" OR "Melampsora occidentalis" OR "Melampsora pinitorqua" OR "Melampsora populina" OR "Melampsora populina subsp. populina" OR "Melampsora populnea" OR "Melampsora populnea f. sp. laricis" OR "Melampsora populnea f. sp. rostrupii" OR "Melampsora pruinosae" OR "Melampsora pulcherrima" OR "Melampsora rostrupii" OR "Melampsora tremulae" OR "Mycosphaerella maculiformis" OR "Mycosphaerella togashiana" OR "Myxosporium ellisii" OR "Nectria ditissima" OR "Neofusicoccum australe" OR "Neofusicoccum luteum" OR "Nigrospora oryzae" OR "Ocellaria ocellata" OR "Olpidium brassicae" OR "Osmoporus proteus" OR "Peyronellaea glomerata" OR "Pezicula populi" OR "Phellinus populicola" OR "Phoma exigua" OR "Phoma glomerata" OR "Phomopsis tirrenica" OR "Phyllactinia corylea" OR "Phyllactinia guttata" OR "Phyllactinia populi" OR "Phyllosticta alcides" OR "Phyllosticta brunnea" OR "Phyllosticta cinerea" OR "Phyllosticta intermixta" OR "Phyllosticta longispora" OR "Phymatotrichum omnivorum" OR "Physalospora obtusa" OR "Phytophthora cactorum" OR "Pleurotus fuscosquamulosus" OR "Pollaccia elegans" OR "Pollaccia radiosa" OR "Polyporus hirsutus" OR "Polyporus sulphureus" OR "Polyporus zonatus" OR "Pseudocercospora salicina" OR "Pseudomonas tumefaciens" OR "Pseudopeziza populi-albae" OR "Rosellinia necatrix" OR "Schizophyllum amplum" OR "Schizophyllum commune" OR "Septoria candida" OR "Septoria musiva" OR "Septoria populi" OR "Septotis populiperda" OR "Sirothecium minor" OR "Stereum purpureum" OR "Synchytrium aureum" OR "Taphrina aurea" OR "Taphrina johansonii" OR "Taphrina populina" OR "Taphrina rhizophora" OR "Trametes hirsuta" OR "Trametes trogii" OR "Trametes zonata" OR "Trichocladium angelicum" OR "Triscelophorus monosporus" OR "Truncatella hartigii" OR "Tryblidaria azarae" OR "Uncinula adunca var. adunca" OR "Uncinula salicis" OR "Valsa ambiens" OR "Valsa nivea" OR "Valsa pauperata" OR "Valsa salicina" OR "Valsa sordida" OR "Vargamyces aquaticus" OR "Venturia macularis" OR "Venturia populina" OR "Venturia tremulae" OR "Venturia tremulae var. populi-albae" OR "Verticillium albo-atrum" OR "Xanthoria parietina" OR "Perrisia populnea" OR "Lasioptera populnea" OR "Aceria populi" OR "Aculops reticulatus" OR "Aculus mogeri" OR "Aculus reticulatus" OR "Megaplatypus mutatus" OR "Longidorus aetnaeus" OR "Xiphinema simile" OR "Scolytus kirschi" OR "Trypophlocus granulatus" OR "Anisandrus dispar" OR "Euwallacea fornicatus" OR "Phyllocnistis xenia" OR "Stigmella assimilella" OR "Stigmella trimaculella" OR

"Ectoedemia turbidella" OR "Pilophorus gallicus" OR "Prionus coriarius" OR "Xylotrechus rusticus" OR "Leiopus nebulosus" OR "Aegomorphus clavipes" OR "Morimus asper" OR "Leiopus punctulatus" OR "Anoplodera rufipes" OR "Saperda perforata" OR "Aegosoma scabricorne" OR "Obrium cantharinum" OR "Rhamnusium bicolor" OR "Poecilium fasciatum" OR "Rhaesus serricollis" OR "Thrips albopilosus" OR "Xanthia icteritia" OR "Epinotia nisella" OR "Batrachedra praeangusta" OR "Dorytomus schoenherri" OR "Dorytomus filirostris" OR "Dorytomus tremulae" OR "Dorytomus minutus" OR "Dorytomus nebulosus" OR "Dorytomus puberulus" OR "Dorytomus villosulus" OR "Dorytomus dejeani" OR "Dorytomus longimanus" OR "Egle concomitans" OR "Ellescus scanicus" OR "Taphrina rhizophora" OR "Eotetranychus populi" OR "Amphitetranychus viennensis" OR "Aegyptobia salisicola" OR "Phratora laticollis" OR "Pemphiqus immunis" OR "Euura fuscomaculata" OR "Pilophorus simulans" OR "Phyllactinia populina" OR "Fusicladium romellianum" OR "Fusicladium radiosum var. populialbae" OR "Venturia radiosa" OR "Drepanopeziza castagnei" OR "Sphaerulina frondicola" OR "Phyllosticta osteospora" OR "Agnocoris rubicundus" OR "Ectagela guttata" OR "Neolygus zebei" OR "Agnocoris rubicundus" OR "Neolygus populi" OR "Melampsora allii-populina" OR "Hyphantria cunea" OR "Noctua janthina" OR "Orthosia incerta" OR "Orthosia populeti" OR "Apterogenum ypsillon" OR "Agrochola macilenta" OR "Scoliopteryx libatrix" OR "Euproctis chrysorrhoea" OR "Lymantria dispar" OR "Macrothylacia rubi" OR "Cerura vinula" OR "Notodonta torva" OR "Notodonta ziczac" OR "Clostera pigra" OR "Tethea ocularis" OR "Lomaspilis marginata" OR "Stegania trimaculata" OR "Apatura ilia" OR "Boudinotiana puella" OR "Earias vernana" OR "Smerinthus ocellata" OR "Laothoe populi" OR "Stauronematus platycerus" OR "Monosteira unicostata" OR "Zygina tithide" OR "Tremulicerus distinguendus" OR "Zygina nivea" OR "Macropsis vicina" OR "Edwardsiana candidula" OR "Populicerus albicans" OR "Kybos populi" OR "Chrysomela tremulae" OR "Apterygothrips neolongiceps" OR "Chrysomela populi" OR "Pamphilius betulae" OR "Sciota rhenella" OR "Sciota elegiella" OR "Gelechia nigra" OR "Sthenarus rotermundi" OR "Acleris hastiana" OR "Acleris rufana" OR "Cenopalpus cumanicus" OR "Aculops knowltoni" OR "Aceria populicanescentis" OR "Phytoptus albae" OR "Byctiscus populi" OR "Chaitophorus populeti sensoriatus" OR "Pemphigus populinigrae" OR "Pemphigus protospirae" OR "Pemphigus spyrothecae" OR "Chaitophorus longisetosus" OR "Myzus persicae" OR "Gypsonoma dealbana" OR "Viridicerus ustulatus" OR "Erysiphe adunca" OR "Phyllactinia populi" OR "Chaitophorus populeti" OR "Gootiella alba" OR "Pachypappa tremulae" OR "Pachypappa vesicalis" OR "Pachypappa warshavensis" OR "Contarinia petioli" OR "Contarinia populi" OR "Dasineura populeti" OR "Harmandiola cavernosa" OR "Harmandiola globuli" OR "Harmandiola populi" OR "Harmandiola pustulans" OR "Harmandiola tremulae" OR "Lasioptera populnea" OR "Aceria populi" OR "Phyllocoptes populi" OR "Camarotoscena speciosa" OR "Taphrina populina" OR "Agromyza albitarsis" OR "Aulagromyza populi" OR "Japanagromyza salicifolii" OR "Zeugophora subspinosa" OR "Rhamphus pulicarius" OR "Tachyerges rufitarsis" OR "Caloptilia stigmatella" OR "Phyllocnistis labyrinthella" OR "Phyllocnistis xenia" OR "Phyllonorycter chiclanella" OR "Phyllonorycter comparella" OR "Phyllonorycter connexella" OR "Phyllonorycter pastorella" OR "Leucoptera sinuella" OR "Ectoedemia klimeschi" OR "Ectoedemia turbidella" OR "Stigmella assimilella" OR "Fenusella glaucopis" OR "Heterarthrus ochropoda" OR "Gypsonoma minutana" OR "Gypsonoma oppressana" OR "Cladius grandis" OR "Synchytrium aureum" OR "Melampsora aecidioides" OR "Melampsora castellana" OR "Melampsora laricis-tremulae" OR "Melampsora magnusiana" OR "Melampsora pinitorqua" OR "Melampsora populnea" OR "Melampsora pulcherrima" OR "Melampsora rostrupii" OR "Aculus mogeri" OR "Aphis fabae" OR "Macrosiphum euphorbiae" OR "Chaitophorus nassonowi" OR "Gypsonoma sociana" OR "Batrachedra praeangusta" OR "Gypsonoma dealbana" OR "Dasineura populnea" OR "Aceria dispar" OR "Crepidodera aurea" OR "Crepidodera aurata" OR "Crepidodera pluta" OR "Crepidodera lamina" OR "Crepidodera nitidula" OR "Agrilus viridis" OR "Saperda populnea" OR "Saperda carcharias" OR "Diaspidiotus armenicus" OR "Didymella macrostoma" OR "Cossus cossus" OR "Agrilus massanensis" OR "Agrilus ater" OR "Eurythyrea micans" OR "Poecilonota variolosa" OR "Eurythyrea aurata" OR "Trachypteris picta" OR "Anthaxia manca" OR "Agrilus suvorovi" OR "Agrilus pratensis" OR "Dicerca aenea" OR "Xyleborus cryptographus" OR "Trypophloeus binodulus" OR "Trypophloeus tremulae" OR "Trypophloeus granulatus" OR "Magdalis nitidipennis" OR "Gypsonoma sociana" OR "Diaspidiotus ostreaeformis" OR "Diaspidiotus gigas" OR "Lepidosaphes ulmi" OR "Chionaspis salicis" OR "Gomezmenoraspis pinicola" OR "Sesia pimplaeformis" OR "Paranthrene diaphana" OR "Synanthedon vespiformis" OR "Synanthedon melliniformis" OR "Sesia apiformis" OR "Chyliza leptogaster" OR "Phylloxerina populi" OR "Phloeomyzus passerinii" OR "Chaitophorus populialbae" OR "Hexomyza schineri" OR "Rabdophaga giraudiana" OR "Viscum album" OR "Paranthrene tabaniformis" OR "Gypsonoma aceriana" OR "Pterocomma populeum" OR "Pulvinaria vitis" OR "Phytophthora cactorum" OR "Salicicola kermanensis" OR "Psylliodes algirica" OR "Rutidosoma globulus"

TABLE B.2 String for *Populus nigra*.

Web of Science All databases	TOPIC: "Populus nigra" OR "P. nigra" OR "black poplar" OR "golden Lombardy poplar" OR "Aigiros nigra" AND
	TOPIC: pathogen* OR pathogenic bacteria OR fung* OR oomycet* OR myce* OR bacteri* OR virus* OR viroid* OR insect\$ OR mite\$ OR phytoplasm* OR arthropod* OR nematod* OR disease\$ OR infecti* OR damag* OR symptom* OR pest\$ OR vector OR hostplant\$ OR "host plant\$" OR host OR "root lesion\$" OR decline\$ OR infestation\$ OR damage\$ OR symptom\$ OR dieback* OR "die back*" OR "malaise" OR aphid\$ OR curculio OR thrip\$ OR cicad\$ OR miner\$ OR borer\$ OR weevil\$ OR "plant bug\$" OR spittlebug\$ OR moth\$ OR mealybug\$ OR curvorm\$ OR pillbug\$ OR "root feeder\$" OR caterpillar\$ OR "foliar feeder\$" OR virosis OR viroses OR blight\$ OR wilt\$ OR wilt@ OR canker OR scab\$ OR rot OR rots OR rotten OR "damping off" OR "damping-off" OR blister\$ OR "smut" OR mould OR mold OR "damping syndrome\$" OR mildew OR scald\$ OR "root knot" OR "root-knot" OR root\$ Not Cyst\$ OR "dagger" OR "plant parasitic" OR "parasitic plant" OR "plant\$ parasitic" OR "root feeding" OR "root\$ feeding"
	 TOPIC: "winged seeds" OR metabolites OR *tannins OR climate OR "maple syrup" OR syrup OR mycorrhiz* OR "carbon loss" OR pollut* OR weather OR propert* OR probes OR spectr* OR antioxidant\$ OR transformation OR RNA OR DNA OR "Secondary plant metabolite\$" OR metabol* OR "Phenolic compounds" OR Quality OR Abiotic OR Storage OR Pollen* OR fertil* OR Mulching OR Nutrient* OR Pruning OR drought OR "human virus" OR "animal disease*" OR "plant extracts" OR immunological OR "purified fraction" OR "traditional medicine" OR medicine OR mammal* OR bird* OR "human disease*" OR "health education" OR bat\$ OR "seedling\$ survival" OR "anthropogenic disturbance" OR "cold resistance" OR "salt stress" OR salinity OR "aCER method" OR "adaptive cognitive emotion regulation" OR landscape NOT

TOPIC: "Acanthosoma haemorrhoidale" OR "Acanthothrips nodicornis" OR "Aceria dispar" OR "Aceria populi" OR "Acleris hastiana" OR "Acleris issikii" OR "Acmaeodera crinita" OR "Acmaeodera pulchra" OR "Acronicta americana" OR "Acronicta leporina" OR "Acronicta lepusculina" OR "Acronicta megacephala" OR "Acronicta oblinita" OR "Acrostalagmus cinnabarinu" OR "Aculus aegerinus" OR "Aculus mogeri" OR "Aegeria apiformis" OR "Aegomorphus clavipes" OR "Aegosoma scabricorne" OR "Aeolesthes sarta" OR "Agrilus ater" OR "Agrilus massanensis" OR "Agrilus pratensis" OR "Agrilus suvorovi" OR "Agrobacterium tumefaciens" OR "Agrochola circellaris" OR "Agrochola macilenta" OR "Agrocybe aegerita" OR "Agrocybe cylindracea" OR "Agrocybe parasitica" OR "Agromyza albitarsis" OR "Aguriahana stellulata" OR "Alebra wahlbergi" OR "Alocerus moesiacus" OR "Alternaria alternata" OR "Amphipyra perflua" OR "Amphipyra pyramidea" OR "Amphipyra pyramidoides" OR "Anacampsis innocuella" OR "Anacampsis populella" OR "Ancylis laetana" OR "Anisandrus dispar" OR "Anisarthron barbipes" OR "Anoplophora chinensis" OR "Anoplophora glabripennis" OR "Anoplus plantaris" OR "Antaeotricha leucillana" OR "Anthaxia manca" OR "Anthocoris nemoralis" OR "Apatura ilia" OR "Apatura iris" OR "Aphis galiae" OR "Aphis maculatae" OR "Apocheima cinerarium" OR "Apotomis dextrana" OR "Apriona cinerea" OR "Apterogenum ypsillon" OR "Archiearis notha" OR "Archips breviplicanus" OR "Armillaria mellea" OR "Aromia moschata" OR "Ascochyta bacteriiformis" OR "Ascochyta populina" OR "Ascochyta populorum" OR "Ascochyta tremulae" OR "Aspidiotus nerii" OR "Asteromella bacteriiformis" OR "Asteromella osteospora" OR "Asteromella populina" OR "Aulacorthum solani" OR "Aulagromyza populi" OR "Aulagromyza populicola" OR "Aulagromyza tremulae" OR "Auriculariopsis ampla" OR "Auriscalpium villipes" OR "Barrmaelia macrospora" OR "Basilarchia archippus" OR "Basilarchia arthemis" OR "Bemisia tabaci" OR "Bjerkandera adusta" OR "Boeremia exigua" OR "Boeremia populi" OR "Botryodiplodia penzigii" OR "Botryosphaeria dothidea" OR "Botryosphaeria parva" OR "Botryosphaeria populi" OR "Botryosphaeria ribis" OR "Boudinotiana notha" OR "Brachylomia viminalis" OR "Byctiscus populi" OR "Cabera exanthemata" OR "Caloptilia chrysolampra" OR "Caloptilia stigmatella" OR "Camarosporidiella populina" OR "Camarosporium propinguum" OR "Camarotoscena fulgidipennis" OR "Camarotoscena speciosa" OR "Camarotoscena subrubescens" OR "Campaea honoraria" OR "Campylomma ribesi" OR "Candidatus Phytoplasma asteris" OR "Candidatus Phytoplasma fraxini" OR "Capnodium citri" OR "Capnodium footii" OR "Capnodium salicinum" OR "Carystoterpa fingens" OR "Catocala amatrix" OR "Catocala concumbens" OR "Catocala elocata" OR "Catocala fraxini" OR "Catocala meskei" OR "Catocala nupta" OR "Catocala oberthuri" OR "Catocala relicta" OR "Catocala unijuga" OR "Ceratocystis microcarpa" OR "Ceratostomella microcarpa" OR "Cercospora jamuensis" OR "Cercospora populina" OR "Cerioporus squamosus" OR "Ceriporiopsis aneirina" OR "Cerura erminea" OR "Cerura iberica" OR "Cerura vinula" OR "Chaitophorus euphraticus" OR "Chaitophorus floris" OR "Chaitophorus indicus" OR "Chaitophorus leucomelas" OR "Chaitophorus longisetosus" OR "Chaitophorus nassonowi" OR "Chaitophorus neglectus" OR "Chaitophorus populeti" OR "Chaitophorus populialbae" OR "Chaitophorus populicola" OR "Chaitophorus pruinosae" OR "Chaitophorus tremulae" OR "Chaitophorus versicolor" OR "Chalcoides aurata" OR "Chalcoides aurea" OR "Chionaspis salicis" OR "Chondroplea populea" OR "Chondrostereum purpureum" OR "Choristoneura diversana" OR "Chrysolina oricalcia" OR "Chrysomela aenea" OR "Chrysomela populi" OR "Chrysomphalus dictyospermi" OR "Ciborinia bifrons" OR "Ciborinia seaveri" OR "Ciborinia whetzelii" OR "Cirrhia icteritia" OR "Cladius grandis" OR "Cladosporium cladosporioides" OR "Cladosporium herbarum" OR "Clostera anachoreta" OR "Clostera anastomosis" OR "Clostera curtula" OR "Clostera inclusa" OR "Clostera pigra" OR "Colletotrichum populi" OR "Colotois pennaria" OR "Comstockaspis perniciosa" OR "Coprinellus micaceus" OR "Coriolopsis gallica" OR "Coryneum populicola" OR "Coryneum populinum" OR "Cossonus linearis" OR "Cossonus parallelepipedus" OR "Cossus cossus" OR "Creosphaeria sassafras" OR "Crepidodera aurata" OR "Crepidodera aurea" OR "Crepidodera fulvicornis" OR "Crepidodera nitidula" OR "Crepidotus crocophyllus" OR "Cryptodiaporthe populea" OR "Cryptodiaporthe salicina" OR "Cryptorhynchus lapathi" OR "Cryptosphaeria ligniota" OR "Cryptosphaeria pullmanensis" OR "Cryptosporium populi" OR "Cryptothrips nigripes" OR "Cryptovalsa ampelina" OR "Curvularia fallax" OR "Cyclocybe aegerita" OR "Cyclocybe cylindracea" OR "Cyclocybe parasitica" OR "Cyptotrama costesii" OR "Cytospora ambiens" OR "Cytospora atrocirrhata" OR "Cytospora chrysosperma" OR "Cytospora melnikii" OR "Cytospora nivea" OR "Cytospora populina" OR "Cytospora salicacearum" OR "Dasineura populicola" OR "Dasyneura populicola" OR "Dematophora necatrix" OR "Dendrothele tetracornis" OR "Dennisiella babingtonii " OR "Deraeocoris lutescens" OR "Diaporthe eres" OR "Diaporthe forabilis" OR "Diaporthe medusaea" OR "Diaporthe putator" OR "Diaporthe rudis" OR "Diaporthe santonensis" OR "Diaspidiotus armenicus" OR "Diaspidiotus caucasicus" OR "Diaspidiotus gigas" OR "Diaspidiotus ostreaeformis" OR "Diaspidiotus slavonicus" OR "Diatrype bullata" OR "Diatrypella populi" OR "Dicerca aenea" OR "Didymella glomerata" OR "Didymosphaeria populina" OR "Diplodia gongrogena" OR "Diplodia mutila" OR "Diplodia seriata" OR "Diplodia tumefaciens" OR "Discella populina" OR "Discohainesia oenotherae" OR "Discosporium hyalinum" OR "Discosporium populeum" OR "Discula tremulae" OR "Dorytomus affinis" OR "Dorytomus dejeani" OR "Dorytomus edoughensis" OR "Dorytomus filirostris" OR "Dorytomus ictor" OR "Dorytomus longimanus" OR "Dorytomus minutus" OR "Dorytomus nebulosus" OR "Dorytomus puberulus" OR "Dorytomus schoenherri" OR "Dorytomus suratus" OR "Dorytomus tremulae" OR "Dorytomus validirostris" OR "Dorytomus villosulus" OR "Dothichiza populea" OR "Dothiorella gregaria" OR "Dothiorella sarmentorum" OR "Drepanopeziza brunnea" OR "Drepanopeziza castagnei" OR "Drepanopeziza populi" OR "Drepanopeziza populorum" OR "Drepanopeziza punctiformis" OR "Drepanopeziza tremulae" OR "Ectoedemia hannoverella" OR "Ectoedemia turbidella" OR "Edwardsiana lethierryi" OR "Egira dolosa" OR "Ellescus scanicus" OR "Elsinoe populi" OR "Elsinoë populi" OR "Elsinoe populi" OR "Enargia abluta" OR "Enargia paleacea" OR "Eotetranychus populi" OR "Epinotia nisella" OR "Epione repandaria" OR "Epipemphigus imaicus" OR "Epipemphigus marginalis" OR "Epiphyas postvittana" OR "Erannis defoliaria" OR "Erannis golda" OR "Erannis tiliaria" OR "Eriophyes populi" OR "Erthesina fullo" OR "Erynnis icelus" OR "Erysiphe adunca" OR "Erysiphe adunca var. adunca" OR "Erysiphe horridula" OR "Erysiphe populicola" OR "Erysiphe salicis" OR "Eucosma hapalosarca" OR "Eudarluca caricis" OR "Eulecanium tiliae" OR "Euproctis chrysorrhoea" OR "Eurythyrea micans" OR "Eutypa acharii" OR "Eutypa lata" OR "Eutypa leptoplaca" OR "Eutypa maura" OR "Eutypa populina" OR "Euura amerinae" OR "Euura caeruleocarpa" OR "Euura pavida" OR "Euwallacea fornicatus" OR "Euwallacea fornicatus sensu lato" OR "Euwallacea fornicatus sensu stricto" OR "Euwallacea kuroshio" OR "Exomias pellucidus" OR "Fenusella glaucopis" OR "Fenusella hortulana" OR "Fomes applanatus" OR "Fomes connatus" OR "Fomes fomentarius" OR "Fomes igniarius" OR "Fomes ulmarius" OR "Fomitopsis pinicola" OR "Framinghamia helvalis" OR "Funalia gallica" OR "Funalia hispida" OR "Funalia trogii" OR "Furcula bifida" OR "Fusarium aquaeductuum" OR "Fusarium silvicola" OR "Fusarium solani-melongenae" OR "Fusarium sporotrichioides" OR "Fusicladium elegans" OR "Fusicladium martianoffianum" OR "Fusicladium radiosum" OR "Fusicladium romellianum" OR "Fusicoccum aesculi" OR "Fusicoccum parvum" OR "Fusicoccum populi" OR

"Fusicolla aquaeductuum" OR "Galerucella lineola" OR "Ganoderma applanatum" OR "Ganoderma lucidum" OR "Gastropacha populifolia" OR "Gelechia nigra" OR "Gelechia rhombelliformis" OR "Gelechia turpella" OR "Glena cribrataria" OR "Gluphisia crenata" OR "Gluphisia septentrionis" OR "Gnomonia gnomon" OR "Gnophomyia viridipennis" OR "Golovinomyces cynoglossi" OR "Gonioctena decemnotata" OR "Graphium penicillioides" OR "Grifola gargal" OR "Gypsonoma aceriana" OR "Gypsonoma dealbana" OR "Gypsonoma minutana" OR "Gypsonoma oppressana" OR "Gypsonoma sociana" OR "Hadrobregmus magnus" OR "Hadrotrichum populi" OR "Haplothrips subtilissimus" OR "Harmonia axyridis" OR "Hedya salicella" OR "Helicobasidium mompa" OR "Helicobasidium purpureum" OR "Hemiberlesia lataniae" OR "Hemiberlesia rapax" OR "Hemicycliophora iberica" OR "Hemicycliophora theinemanni" OR "Heterarthrus ochropoda" OR "Heterarthrus ochropodus" OR "Heterobasidion annosum" OR "Hexomyza schineri" OR "Hoplandrothrips bidens" OR "Hoplandrothrips ellisi" OR "Hoplothrips fungi" OR "Hoplothrips ulmi" OR "Hyalopeziza millepunctata" OR "Hydnum platense" OR "Hyphantria cunea" OR "Hypocrea atrata" OR "Hypocrea contorta" OR "Hypoxylon bifrons" OR "Hypoxylon fuscum" OR "Hypoxylon rubiginosum" OR "Hypoxylon rubiginosum var. rubiginosum" OR "Hysterium pulicare" OR "Hysterobrevium mori" OR "Hysterographium mori" OR "Idiocerus distinguendus" OR "Idiocerus fulgidus" OR "Idiocerus poecilus" OR "Idiocerus tremulae" OR "Idiocerus vitreus" OR "Inonotus farlowii" OR "Inonotus hispidus" OR "Ipimorpha nanaimo" OR "Ipimorpha pleonectusa" OR "Ipimorpha retusa" OR "Ipimorpha subtus" OR "Isochnus sequensi" OR "Jalapriya toruloides" OR "Japanagromyza salicifolii" OR "Junghuhnia vincta" OR "Kalotermes brouni" OR "Kastanostachys aterrima" OR "Kleidocerys resedae" OR "Kybos abstrusus" OR "Kybos populi" OR "Lamia textor" OR "Laothoe populi" OR "Laothoë populi" OR "Leiopus nebulosus" OR "Lentinellus vulpinus" OR "Lepidosaphes malicola" OR "Lepidosaphes salicina" OR "Lepidosaphes ulmi" OR "Leptosphaeria salicinearum" OR "Leptura quadrifasciata" OR "Leucoma salicis" OR "Leucoptera sinuella" OR "Leucostoma niveum" OR "Limenitis populi" OR "Lispothrips crassipes" OR "Lobophora halterata" OR "Lomaspilis marginata" OR "Longidorus apuloides" OR "Longidorus iranicus" OR "Lophiostoma nuculoides" OR "Lophocampa maculata" OR "Lucanus cervus" OR "Luperus xanthopoda" OR "Lycia hirtaria" OR "Lycia ursaria" OR "Lyctus brunneus" OR "Lymantria dispar" OR "Lymantria monacha" OR "Lymantria obfuscata" OR "Macrophoma gongrogena" OR "Macropsis graminea" OR "Macrothylacia rubi" OR "Magdalis nitidipennis" OR "Malacosoma californica" OR "Malacosoma neustria" OR "Marasmius favrei" OR "Marssonia populina" OR "Marssonia rhabdospora" OR "Marssonina brunnea" OR "Marssonina castagnei" OR "Marssonina populi" OR "Marssonina rhabdospora" OR "Megaplatypus mutatus" OR "Melampsora abietis-canadensis" OR "Melampsora abietis-populi" OR "Melampsora allii-populina" OR "Melampsora ciliata" OR "Melampsora laricis-populina" OR "Melampsora magnusiana" OR "Melampsora medusae" OR "Melampsora medusae f.sp. deltoidis" OR "Melampsora mercurialis-tremulae" OR "Melampsora mercuriali-tremulae" OR "Melampsora microspora" OR "Melampsora occidentalis" OR "Melampsora populina" OR "Melampsora populnea" OR "Melampsora pruinosae" OR "Melampsora pulcherrima" OR "Melampsora rostrupii" OR "Melanaspis inopinata" OR "Melanconium hyalinum" OR "Menesia bipunctata" OR "Messa glaucopis" OR "Messa hortulana" OR "Mikiola populicola" OR "Mollisina flava" OR "Monosteira discoidalis" OR "Monosteira unicostata" OR "Mordwilkoja vagabunda" OR "Morganella longispina" OR "Morimus asper" OR "Mycosphaerella maculiformis" OR "Mycosphaerella populi" OR "Mycosphaerella populorum" OR "Mycosphaerella togashiana" OR "Myxosporium tremulae" OR "Myzus persicae" OR "Myzus varians" OR "Narycia monilifera" OR "Naupactus xanthographus" OR "Nectria cinnabarina" OR "Nectria coccinea" OR "Nectria galligena" OR "Neocosmospora ipomoeae" OR "Neocosmospora silvicola" OR "Neofusicoccum luteum" OR "Neofusicoccum parvum" OR "Neofusicoccum ribis" OR "Neolygus zebei" OR "Neonectria coccinea" OR "Neonectria ditissima" OR "Neoscytalidium dimidiatum" OR "Neoscytalidium hyalinum" OR "Neoscytalidium novaehollandiae" OR "Nesothrips propinquus" OR "Notodonta torva" OR "Notodonta tritophus" OR "Notodonta ziczac" OR "Nycteola asiatica" OR "Nycteola cinereana" OR "Nymphalis antiopa" OR "Nymphalis polychloros" OR "Obrium cantharinum" OR "Oceanaspidiotus spinosus" OR "Odontopera bidentata" OR "Oemona hirta" OR "Oligocentria semirufescens" OR "Ophiostoma fusiforme" OR "Orgyia antiqua" OR "Orgyia leucostigma" OR "Orientus ishidae" OR "Orius minutus" OR "Orthosia cerasi" OR "Orthosia gracilis" OR "Orthosia incerta" OR "Orthosia populeti" OR "Orthotylus prasinus" OR "Otiorhynchus ovalipennis" OR "Oxyporus populinus" OR "Pachyderris nigricans" OR "Pachypappa marsupialis" OR "Pachypappa vesicalis" OR "Pachysphinx modesta" OR "Paecilomyces variotii" OR "Pamphilius betulae" OR "Pamphilius histrio" OR "Pandemis chlorograpta" OR "Papilio cresphontes" OR "Paraleucoptera albella" OR "Paraleucoptera sinuella" OR "Paralongidorus serbicus" OR "Paranthrene tabaniformis" OR "Paraphytomyza populi" OR "Paraphytomyza populicola" OR "Parastichtis suspecta" OR "Parastichtis ypsillon" OR "Parlatoria oleae" OR "Parthenolecanium corni" OR "Pemphigus borealis" OR "Pemphigus bursarius" OR "Pemphigus dorocola" OR "Pemphigus filaginis" OR "Pemphigus gairi" OR "Pemphigus immunis" OR "Pemphigus lysimachiae" OR "Pemphigus matsumurai" OR "Pemphigus mordvilkoi" OR "Pemphigus napaeus" OR "Pemphigus passeki" OR "Pemphigus phenax" OR "Pemphigus plicatus" OR "Pemphigus populi" OR "Pemphigus populinigrae" OR "Pemphigus protospirae" OR "Pemphigus spirothecae" OR "Pemphigus spyrothecae" OR "Pemphigus trehernei" OR "Pemphigus vesicarius" OR "Periphoba hircia" OR "Perisomena caecigena" OR "Pestalotia populi-nigrae" OR "Pestalotia populi-nigrae var. italica" OR "Pestalotiopsis populi-nigrae" OR "Petriella asymmetrica" OR "Petriella asymmetrica var. cypria" OR "Petriella sordida" OR "Peyronellaea glomerata" OR "Pezicula populi" OR "Pezizella oenotherae" OR "Phaeoacremonium parasiticum" OR "Phaeoramularia maculicola" OR "Phaiogramma etruscaria" OR "Phalera bucephala" OR "Phellinus igniarius" OR "Pheosia gnoma" OR "Pheosia portlandia" OR "Pheosia tremula" OR "Phigalia pilosaria" OR "Phigalia sinuosaria" OR "Phlaeothrips coriaceus" OR "Phloeomyzus passerinii" OR "Phloeomyzus redelei" OR "Phloeophagosoma thoracicum" OR "Phobetron hipparchia" OR "Pholiota crassivela" OR "Pholiota destruens" OR "Pholiota edulis" OR "Pholiota populnea" OR "Phoma exigua" OR "Phoma exigua var. populi" OR "Phoma exigua var. exigua" OR "Phoma glomerata" OR "Phoma populicola" OR "Phoma populi-nigrae" OR "Phoma exigua f.sp. exigua" OR "Phratora laticollis" OR "Phratora tibialis" OR "Phratora vitellinae" OR "Phtheochroa micana" OR "Phtheochroa schreibersiana" OR "Phyllactinia populi" OR "Phyllactinia populina" OR "Phyllobius pyri" OR "Phyllocnistis extrematrix" OR "Phyllocnistis populiella" OR "Phyllocnistis unipunctella" OR "Phyllocoptes didelphis" OR "Phyllocoptes populi" OR "Phyllonorycter comparella" OR "Phyllonorycter connexella" OR "Phyllonorycter pastorella" OR "Phyllonorycter populiella" OR "Phyllonorycter populifoliella" OR "Phyllosticta alcides" OR "Phyllosticta intermixta" OR "Phyllosticta osteospora" OR "Phyllosticta populea" OR "Phyllosticta populina" OR "Phyllosticta populorum" OR "Phylloxerina populi" OR "Phymatotrichopsis omnivora" OR "Phymatotrichum omnivorum" OR "Phytobia cambii" OR "Phytocoris longipennis" OR "Phytophthora citricola" OR "Plagiodera versicolora" OR "Plagiostoma apiculatum" OR "Planotortrix excessana" OR "Plectosphaerella populi" OR "Pleurotus ostreatus" OR "Poecilocampa populi" OR

"Poecilonota variolosa" OR "Poecilothrips albopictus" OR "Pollaccia elegans" OR "Pollaccia radiosa" OR "Polyozellus tristis" OR "Polyporus adustus" OR "Polyporus farlowii" OR "Polyporus hispidus" OR "Polyporus squamosus" OR "Popillia japonica" OR "Poplar mosaic virus" OR "Populicerus nitidissimus" OR "Pristiphora conjugata" OR "Pseudocamarosporium propinquum" OR "Pseudocercospora salicina" OR "Pseudocercospora togashiana" OR "Pseudoclavellaria amerinae" OR "Pseudomonas syringae pv. syringae" OR "Pseudopeziza populi-albae" OR "Pseudotomentella tristis" OR "Pterocomma bhutanense" OR "Pterocomma bicolor" OR "Pterocomma pilosum" OR "Pterocomma populeum" OR "Pterocomma sinipopulifoliae" OR "Pterocomma tremulae" OR "Pterocomma yezoense" OR "Pterocomma anyangense" OR "Pterocomma atuberculatum" OR "Pterostoma palpina" OR "Ptilodon capucina" OR "Pulvinaria loralaiensis" OR "Pulvinaria vitis" OR "Pygaera anastomosis" OR "Quadraspidiotus zonatus" OR "Raduliporus aneirinus" OR "Ramphus pulicarius" OR "Ramularia uredinis" OR "Raphia frater" OR "Rhabdospora longispora" OR "Rhamnusium bicolor" OR "Rhamnusium graecum" OR "Rhamphus pulicarius" OR "Rhynchaenus decoratus" OR "Rhynchaenus rusci" OR "Rhynchaenus salicis" OR "Rhynchaenus stigma" OR "Rhytidodus decimusquartus" OR "Rhytidodus wagneri" OR "Ribautiana ulmi" OR "Rigidoporus ulmarius" OR "Rigidoporus vinctus" OR "Ropalopus macropus" OR "Rosellinia corticalis" OR "Rosellinia necatrix" OR "Rosellinia novae-zelandiae" OR "Salicicola kermanensis" OR "Saliciphaga acharis" OR "Saperda carcharias" OR "Saperda perforata" OR "Saperda populnea" OR "Saperda scalaris" OR "Schizophyllum amplum" OR "Sciota rhenella" OR "Scleroderma bovista" OR "Scoliopteryx libatrix" OR "Scolytus intricatus" OR "Scolytus multistriatus" OR "Scolytus scolytus" OR "Selenia tetralunaria" OR "Septoria populi" OR "Sesia apiformis" OR "Sesia flavicollis" OR "Sesia siningensis" OR "Sesia yezoensis" OR "Sillia ferruginea" OR "Smerinthus cerisyi" OR "Smerinthus ocellata" OR "Speira toruloides" OR "Sphaceloma populi" OR "Sphaerellopsis filum" OR "Sphaerulina frondicola" OR "Sphaerulina musiva" OR "Sphinx luscitiosa" OR "Spongipellis spumea" OR "Spongipellis spumeus" OR "Sporocadus populinus" OR "Sporothrix fusiformis" OR "Stauronematus compressicornis" OR "Stauronematus platycerus" OR "Stegania trimaculata" OR "Stenidiocerus poecilus" OR "Stereum purpureum" OR "Stictochorella populi-nigrae" OR "Stigmella trimaculella" OR "Stomaphis longirostris" OR "Stomaphis sp. nr graffii" OR "Subacronicta megacephala" OR "Synanthedon melliniformis" OR "Synanthedon vespiformis" OR "Tachyerges decoratus" OR "Tachyerges pseudostigma" OR "Tachyerges rufitarsis" OR "Tachyerges salicis" OR "Tachyerges stigma" OR "Taphrina aurea" OR "Taphrina populina" OR "Teichospora macrosperma" OR "Teichospora pruniformis" OR "Tethea ocularis" OR "Tethea or" OR "Tetra populi" OR "Tetranychus ludeni" OR "Tetranychus urticae" OR "Thecabius affinis" OR "Thecabius lysimachiae" OR "Thyridaria macrostomoides" OR "Thyridopteryx ephemeraeformis" OR "Trachypteris picta" OR "Trachysmia schreibersiana" OR "Trametes hirsuta" OR "Trametes hispida" OR "Trametes trogii" OR "Trematosphaeria pertusa" OR "Tremex fuscicornis" OR "Tremulicerus fulgidus" OR "Tremulicerus vitreus" OR "Trichoderma citrinoviride" OR "Trichoferus fasciculatus" OR "Trirachys sartus" OR "Tritophia tritophus" OR "Trochila populorum" OR "Trypophloeus asperatus" OR "Trypophloeus binodulus" OR "Trypophloeus granulatus" OR "Trypophloeus rybinskii" OR "Trypophloeus tremulae" OR "Tubercularia vulgaris" OR "Tuberolachnus salignus" OR "Typhula setipes" OR "Uncinula adunca" OR "Uncinula adunca var. adunca" OR "Uncinula populi" OR "Uncinula salicis" OR "Uncinula tenuitunicata" OR "Uraba lugens" OR "Uredo tholopsora" OR "Valsa ambiens" OR "Valsa nivea" OR "Valsa sordida" OR "Venturia macularis" OR "Venturia martianoffiana' OR "Venturia populina" OR "Venturia radiosa" OR "Venturia tremulae" OR "Verbasciola petioli" OR "Verticillium tenerum" OR "Viscum album" OR "Xanthia icteritia" OR "Xanthia ocellaris" OR "Xanthomonas arboricola pv. populi" OR "Xanthomonas populi" OR "Xyleborinus attenuatus" OR "Xyleborinus saxesenii" OR "Xyleborus cryptographus" OR "Xyleborus dispar" OR "Xylotoles griseus" OR "Xylotrechus namanganensis" OR "Xylotrechus rusticus" OR "Ypsolopha parenthesella" OR "Zeugophora flavicollis" OR "Zeugophora scutellaris" OR "Zeugophora subspinosa" OR "Zignoella aterrima" OR "Zygina nivea"

TABLE B.3 String for Populus tremula.

Web of Science All databases databases **TOPIC:** "Populus tremula" OR "P. tremula" OR "European aspen" OR "trembling poplar" OR "Populus australis" OR "Populus bonatii" OR "Populus duclouxiana" OR "Populus microcarpa" OR "Populus pseudotremula" OR "Populus repanda" OR "Populus rotundifolia" OR "Populus villosa" OR "Tremula vulgaris"

AND

TOPIC: pathogen* OR pathogenic bacteria OR fung* OR oomycet* OR myce* OR bacteri* OR virus* OR viroid* OR insect\$ OR mite\$ OR phytoplasm* OR arthropod* OR nematod* OR disease\$ OR infecti* OR damag* OR symptom* OR pest\$ OR vector OR hostplant\$ OR "host plant\$" OR host OR "root lesion\$" OR decline\$ OR infestation\$ OR damage\$ OR symptom\$ OR dieback* OR "die back*" OR "malaise" OR aphid\$ OR curculio OR thrip\$ OR cicad\$ OR miner\$ OR borer\$ OR weevil\$ OR "plant bug\$" OR spittlebug\$ OR moth\$ OR mealybug\$ OR cutworm\$ OR pillbug\$ OR "root feeder\$" OR caterpillar\$ OR "foliar feeder\$" OR virosis OR viroses OR blight\$ OR wilt\$ OR wilted OR canker OR scab\$ OR rot OR rots OR rotten OR "damping off" OR "damping-off" OR blister\$ OR "smut" OR mould OR mold OR "damping syndrome\$" OR mildew OR scald\$ OR "root knot" OR "root feeding" OR "root\$feeding"

NOT

TOPIC: "winged seeds" OR metabolites OR *tannins OR climate OR "maple syrup" OR syrup OR mycorrhiz* OR "carbon loss" OR pollut* OR weather OR propert* OR probes OR spectr* OR antioxidant\$ OR transformation OR RNA OR DNA OR "Secondary plant metabolite\$" OR metabol* OR "Phenolic compounds" OR Quality OR Abiotic OR Storage OR Pollen* OR fertil* OR Mulching OR Nutrient* OR Pruning OR drought OR "human virus" OR "animal disease*" OR "plant extracts" OR immunological OR "purified fraction" OR "traditional medicine" OR medicine OR mammal* OR bird* OR "human disease*" OR biomarker\$ OR "health education" OR bat\$ OR "seedling\$ survival" OR "anthropogenic disturbance" OR "cold resistance" OR "salt stress" OR salinity OR "aCER method" OR Miocene OR postglacial OR "weed control" OR landscape

TOPIC: "Acalyptus carpini" OR "Acanthonitschkea tristis" OR "Acanthosoma haemorrhoidale" OR "Aceria dispar" OR "Aceria populi" OR "Aceria varia" OR "Aceria varius" OR "Acleris emargana" OR "Acleris roscidana" OR "Acleris variegana" OR "Accossus terebra" OR "Acremonium murorum" OR "Acrobeles elaboratus" OR "Acronicta alni" OR "Acronicta leporina" OR "Acronicta megacephala" OR "Acronicta psi" OR "Actinonema populorum" OR "Aculops granulatus" OR "Aculus aegerinus" OR "Aculus aegirinus" OR "Aculus dispar" OR "Aculus mogeri" OR "Aculus reticulatus" OR "Aculus unctus" OR "Aegomorphus clavipes" OR "Aegosoma scabricorne" OR "Aegyptobia salisicola" OR "Agonopterix ocellana" OR "Agrilus ater" OR "Agrilus fleischeri" OR "Agrilus horni" OR "Agrilus pratensis" OR "Agrilus pseudocyaneus" OR "Agrilus subauratus" OR "Agrilus suvorovi" OR "Agrilus suvorovi populneus" OR "Agrilus viridis" OR "Agriopis marginaria" OR "Agrochola circellaris" OR "Agrochola macilenta" OR "Agromyza albitarsis" OR "Alcis repandata" OR "Aleurodiscus polygonius" OR "Aleurodiscus roseus" OR "Allygidius commutatus" OR "Alosterna tabacicolor" OR "Alsophila aescularia" OR "Alternaria alternata" OR "Alternaria tenuis" OR "Amauronematus krausi" OR "Amauronematus puniceus" OR "Ametastegia albipes" OR "Amphipyra berbera" OR "Amphipyra perflua" OR "Amphisphaerella amphisphaerioides" OR "Amphisphaerella dispersella" OR "Amphisphaeria millepunctata" OR "Anacampsis populella" OR "Ancylis laetana" OR "Ancylis tineana" OR "Angerona prunaria" OR "Anisandrus dispar" OR "Anisandrus maiche" OR "Anoplophora chinensis" OR "Anoplophora glabripennis" OR "Anorthoa munda" OR "Anthaxia manca" OR "Anthostomella anserina" OR "Antrodia macra" OR "Antrodia mellita" OR "Antrodia pulvinascens" OR "Aonidiella citrina" OR "Apatura ilia" OR "Apatura iris" OR "Aphis fabae" OR "Aphis fabae fabae" OR "Apiognomonia errabunda" OR "Apocheima pilosaria" OR "Apoda limacodes" OR "Apoderus coryli" OR "Aporcelaimellus obscurus" OR "Apotomis inundana" OR "Apriona cinerea" OR "Apriona germari" OR "Apriona rugicollis" OR "Arboridia spathulata" OR "Archiearis notha" OR "Archips betulana" OR "Archips crataegana" OR "Archips xylosteana" OR "Archips xylosteanus" OR "Arctia caja" OR "Armillaria borealis" OR "Armillaria mellea" OR "Arrhenia tenella" OR "Arrhenodes minutus" OR "Arthopyrenia cinereopruinosa" OR "Ascochyta tremulae" OR "Asiphum tremulae" OR "Aspen mosaic-associated virus" OR "Aspergillus fumigatus" OR "Aspidiotus juglandis" OR "Asterodon ferruginosum" OR "Asterodon ferruginosus" OR "Asteroma frondicola" OR "Asteroscopus sphinx" OR "Athelicium hallenbergii" OR "Aulagromyza populi" OR "Aulagromyza populicola" OR "Aulagromyza tremulae" OR "Auriculariopsis ampla" OR "Bacidia fraxinea" OR "Baltazaria galactina" OR "Barrmaelia oxyacanthae" OR "Batrachedra praeangusta" OR "Biscogniauxia mediterranea" OR "Biston betularia" OR "Biston strataria" OR "Botryobasidium laeve" OR "Botryosphaeria berengeriana" OR "Botryosphaeria dothidea" OR "Botryosphaeria populi" OR "Botrytis asperula" OR "Boudinotiana notha" OR "Brachionycha nubeculosa" OR "Brachyarthrum limitatum" OR "Brachylomia viminalis" OR "Brachyopa pilosa" OR "Brachyopa scutellaris" OR "Brachysporium fusiforme" OR "Bryobia rubrioculus" OR "Bursaphelenchus populi" OR "Bursaphelenchus trypophloei" OR "Byctiscus betulae" OR "Byctiscus populi" OR "Cabera exanthemata" OR "Cabera pusaria" OR "Caliciopsis calicioides" OR "Caliroa annulipes" OR "Caliroa tremulae' OR "Calliteara pudibunda" OR "Caloptilia stigmatella" OR "Camarosporium propinquum" OR "Camarotoscena speciosa" OR "Candelabrochaete septocystidia" OR "Capnodium citri" OR "Capnodium elongatum" OR "Capronia mansonii" OR "Capronia pulcherrima" OR "Carestiella socia" OR "Carlavirus populi" OR "Catocala elocata" OR "Catocala fraxini" OR "Catocala nupta" OR "Centrotus cornutus" OR "Cerambyx scopolii" OR "Ceratocystiopsis synnemata" OR "Ceratosphaeria lampadophora" OR "Cerioporus leptocephalus" OR "Cerioporus squamosus" OR "Ceriporia septocystidia" OR "Ceriporiopsis aneirina" OR "Ceriporiopsis niger" OR "Ceriporiopsis nigra" OR "Ceroplastes ceriferus" OR "Ceroplastes rusci" OR "Cerrena unicolor" OR "Cerura erminea" OR "Cerura vinula" OR "Ceuthospora pulvinata" OR "Chaetospermum carneum" OR "Chaetospermum chaetosporum" OR "Chaetospermum tubercularioides" OR "Chaetosphaeria pulviscula" OR "Chaitophorus albus" OR "Chaitophorus leucomelas" OR "Chaitophorus longisetosus" OR "Chaitophorus nassonowi" OR "Chaitophorus populeti" OR "Chaitophorus populeti sensoriatus" OR "Chaitophorus populialbae" OR "Chaitophorus tremulae" OR "Chaitophorus tremulae ssp. Sorini" OR "Chalara cylindrosperma" OR "Chalcoides aurea" OR "Chalcoides nitidula" OR "Chionaspis salicis" OR "Chloroclysta miata" OR "Chloroclysta siterata" OR "Chlorophorus varius" OR "Chondrostereum purpureum" OR "Choristoneura diversana" OR "Chrysobothris affinis" OR "Chrysomela cuprea" OR "Chrysomela populi" OR "Chrysomela tremula" OR "Chyliza leptogaster" OR "Cimbex luteus" OR "Cladius grandis" OR "Cladobotryum mycophilum" OR "Cladosporium cladosporioides" OR "Cladosporium epiphyllum" OR "Cladosporium fumago" OR "Cladosporium herbarum" OR "Cladosporium nigrellum" OR "Cladosporium populicola" OR "Clostera anachoreta" OR "Clostera anastomosis" OR "Clostera curtula" OR "Clostera pigra" OR "Coccomyces tumidus" OR "Coleophora lusciniaepennella" OR "Colobochyla salicalis" OR "Colocasia coryli" OR "Colotois pennaria" OR "Comstockaspis perniciosa" OR "Coniella populina" OR "Coniochaeta dakotensis" OR "Conioscinella gallarum" OR "Coniothyrium fuckelii" OR "Conistra vaccinii" OR "Contarinia petioli" OR "Contarinia populi" OR "Contarinia tremulae" OR "Coronophora ovipara" OR "Corticium roseum" OR "Cortinarius cumatilis" OR "Coryneum populinum" OR "Cosmia trapezina" OR "Cossus cossus" OR "Crepidodera aurata" OR "Crepidodera aurea" OR "Crepidodera fulvicornis" OR "Crepidodera lamina" OR "Crepidodera nitidula" OR "Crepidodera pluta" OR "Cresporhaphis wienkampii" OR "Crocallis elinguaria" OR "Cryptadelphia fusiformis" OR "Cryptocephalus frontalis" OR "Cryptocephalus parvulus" OR "Cryptocephalus sexpunctatus" OR "Cryptocline dubia" OR "Cryptocoryneum condensatum" OR "Cryptodiaporthe populea" OR "Cryptorhynchus lapathi" OR "Cryptosphaeria ligniota" OR "Cryptosphaeria populina" OR "Cryptosporiopsis fasciculata" OR "Cydia corollana" OR "Cylindrosporium populinum" OR "Cyrtidula hippocastani" OR "Cytospora ambiens" OR "Cytospora chrysosperma" OR "Cytospora leucostoma" OR "Cytospora nivea" OR "Cytospora populina" OR "Daedalea unicolor" OR "Daldinia concentrica" OR "Daldinia pyrenaica" OR "Daruvedia bacillata" OR "Dasineura populeti" OR "Dasineura populnea" OR "Descarpentriesina variolosa" OR "Diaphora mendica" OR "Diaspidiotus gigas" OR "Diaspidiotus lenticularis" OR "Diaspidiotus ostreaeformis" OR "Diaspidiotus perniciosus" OR "Diaspidiotus pyri" OR "Diatrype bullata" OR "Dicallomera fascelina" OR "Dicerca aenea" OR "Dictyotrichiella mansonii" OR "Didymella barbieri" OR "Didymosphaeria congruella" OR "Dinemasporium strigosum" OR "Dinoptera collaris" OR "Diplococcium spicatum" OR "Diplodia gongrogena" OR "Diplodia mutila" OR "Diplodia tumefaciens" OR "Discosia artocreas" OR "Discosia julia" OR "Diurnea fagella" OR "Doraphis populi" OR "Doraphis populi ssp. Tremulae" OR "Dorytomus affinis" OR "Dorytomus dejeani" OR "Dorytomus edoughensis" OR "Dorytomus ictor" OR "Dorytomus longimanus" OR "Dorytomus nordenskioldi" OR "Dorytomus rubrirostris" OR "Dorytomus suratus" OR "Dorytomus taeniatus" OR "Dorytomus tortrix" OR "Dorytomus tremulae" OR "Dothichiza populina" OR "Dothiopsis tremulae" OR "Dothiora sphaeroides" OR "Dothiorella crepinii" OR "Drepana falcataria" OR "Drepanopeziza brunnea" OR "Drepanopeziza castagnei" OR "Drepanopeziza populi" OR "Drepanopeziza punctiformis" OR "Drepanopeziza tremulae" OR "Echinosphaeria canescens" OR "Ectoedemia argyropeza" OR "Ectropis crepuscularia" OR "Egle ciliata" OR

"Egle muscaria" OR "Elasmostethus interstinctus" OR "Elasmucha grisea" OR "Eligmodonta ziczac" OR "Ellescus scanicus" OR "Enargia paleacea" OR "Encoelia fascicularis" OR "Endoxyla populi" OR "Ennomos erosaria" OR "Ennomos quercinaria" OR "Entoleuca mammata" OR "Eotetranychus populi" OR "Epinotia cinereana" OR "Epinotia maculana" OR "Epinotia nisella" OR "Epinotia solandriana" OR "Epione paralellaria" OR "Epione repandaria" OR "Epione vespertaria" OR "Epirranthis diversata" OR "Epirrita autumnata" OR "Epirrita christyi" OR "Epirrita dilutata" OR "Erannis defoliaria" OR "Eriogaster lanestris" OR "Eriophyes diversipunctatus" OR "Eriophyes populi" OR "Erostella minima" OR "Erysiphe adunca" OR "Erysiphe penicillata" OR "Erysiphe salicis" OR "Eulecanium douglasi" OR "Eulecanium tiliae" OR "Eulecanium transvittatum" OR "Eulithis populata" OR "Eulithis testata" OR "Euphydryas maturna" OR "Eupithecia subfuscata" OR "Euproctis chrysorrhoea" OR "Eupsilia transversa" OR "Eutypa sparsa" OR "Eutypella stellulata" OR "Euura amerinae" OR "Euura atra" OR "Euura cadderensis" OR "Euura fahraei" OR "Euura fuscomaculata" OR "Euura krausi" OR "Euura miliaris" OR "Euura nigricornis" OR "Euura papillosa" OR "Euura pavida" OR "Euura ranini" OR "Euura sylvestris" OR "Euwallacea kuroshio" OR "Exophiala calicioides" OR "Favolus pseudobetulinus" OR "Fenusella glaucopis" OR "Flavidoporia mellita" OR "Flavidoporia pulvinascens" OR "Fomes annosus" OR "Fomes connatus" OR "Fomes fomentarius" OR "Fomes igniarius" OR "Fomitopsis pinicola" OR "Furcula bifida" OR "Furcula furcula" OR "Fusarium sambucinum" OR "Fusicladium asteroma" OR "Fusicladium martianoffianum" OR "Fusicladium radiosum" OR "Fusicladium radiosum var. radiosum" OR "Fusicladium romellianum" OR "Fusidium griseum" OR "Ganoderma lipsiense" OR "Garella musculana" OR "Gelechia muscosella" OR "Gelechia nigra" OR "Gibberifera simplana" OR "Gliomastix cerealis" OR "Gliomastix convoluta" OR "Gloeosporium dubium" OR "Gloeosporium naevioides" OR "Gloeosporium tremulae" OR "Gloniopsis curvata" OR "Gluphisia crenata" OR "Glyphium elatum" OR "Gnomonia cerastis" OR "Gnomonia fahrendorffii" OR "Gonioctena decemnotata" OR "Gonioctena viminalis" OR "Gootiella tremulae" OR "Gypsonoma aceriana" OR "Gypsonoma imparana" OR "Gypsonoma minutana" OR "Gypsonoma nitidulana" OR "Gypsonoma sociana" OR "Hammerschmidtia ferruginea" OR "Harmandia loewi" OR "Harmandiola cavernosa" OR "Harmandiola cavinosa" OR "Harmandiola globuli" OR "Harmandiola populi" OR "Harmandiola pustulans" OR "Harmandiola tremulae" OR "Harzia acremonioides" OR "Hedya salicella" OR "Helicogloea aquilonia" OR "Helicogloea pellucida" OR "Helicogloea septifera" OR "Helicogloea sputum" OR "Helicoma fumosum" OR "Helicomyia saliciperda" OR "Heliococcus bohemicus" OR "Hericium erinaceus" OR "Heterarthrus ochropoda" OR "Heterobasidion annosum" OR "Heterobasidion parviporum" OR "Heteroradulum deglubens" OR "Hexomyza schineri" OR "Hilberina caudata" OR "Hyalinia rubella" OR "Hydria undulata" OR "Hydriomena furcata" OR "Hydropisphaera peziza" OR "Hylecoetus dermestoides" OR "Hymenoscyphus phyllogenus" OR "Hyphantria cunea" OR "Hyphoderma lapponicum" OR "Hypochnicium bombycinum" OR "Hypomyces pseudopolyporinus" OR "Hypoxylon macrocarpum" OR "Hypoxylon mammatum" OR "Hypoxylon mediterraneum" OR "Hypoxylon morsei" OR "Hypoxylon rubiginosum" OR "Hypoxylon rubiginosum var. rubiginosum" OR "Hysterobrevium curvatum" OR "Idiocerus distinguendus" OR "Idiocerus fulgidus" OR "Idiocerus laminatus" OR "Idiocerus populi" OR "Idiocerus tremulae" OR "Inocutis dryophila" OR "Inocutis rheades" OR "Inonotus hispidus" OR "Ipimorpha contusa" OR "Ipimorpha subtusa" OR "Janus luteipes" OR "Jikradia olitoria" OR "Jodis lactearia" OR "Kalmusia coniothyrium" OR "Kalmusia ebuli" OR "Kretzschmaria deusta" OR "Kybos populi" OR "Lacanobia thalassina" OR "Lachnella karstenii" OR "Lachnum corticale" OR "Laetiporus sulphureus" OR "Lahmia kunzei" OR "Lamia textor" OR "Laothoe populi" OR "Lasiobelonium corticale" OR "Lasiocampa quercus" OR "Lasiocampa trifolii" OR "Lasioptera populnea" OR "Lasiosphaeria caudata" OR "Lasiosphaeria crinita" OR "Lasiosphaeria ovina" OR "Lasiosphaeria pyramidata" OR "Lecanidion clavisporum" OR "Ledra aurita" OR "Leiopus linnei" OR "Leiopus nebulosus" OR "Leiopus punctulatus" OR "Lentinus brumalis" OR "Lentinus substrictus" OR "Lepidosaphes malicola" OR "Lepidosaphes ulmi" OR "Lepidosaphes ussuriensis" OR "Leptographium alneum" OR "Leptographium alni" OR "Leptosillia wienkampii" OR "Leptosphaeria immunda" OR "Leptothyrium populi" OR "Leptura annularis" OR "Leptura quadrifasciata" OR "Lepturalia nigripes" OR "Leucoma salicis" OR "Leucoptera sinuella" OR "Leucostoma niveum" OR "Leucostoma persoonii" OR "Limenitis populi" OR "Lindbergina aurovittata" OR "Linospora ceuthocarpa" OR "Linospora populina" OR "Lispothrips crassipes" OR "Lithophane ornitopus" OR "Lobophora halterata" OR "Lochmaea caprea" OR "Lochmaea capreae" OR "Lomaspilis bithynica" OR "Lomaspilis marginata" OR "Lomographa temerata" OR "Longidorus attenuatus" OR "Lophiostoma compressum" OR "Lophiostoma macrostomoides" OR "Lophiostoma nucula" OR "Lophiotrema nucula" OR "Luperus longicornis" OR "Lycia graecarius" OR "Lycia hirtaria" OR "Lygaeonematus compressicornis" OR "Lygocoris pabulinus" OR "Lygocoris populi" OR "Lymantria dispar asiatica" OR "Lymantria monacha" OR "Macrolabis bedeguariformis" OR "Macroleptura thoracica" OR "Macrophoma gongrogena" OR "Macrophoma tumefaciens" OR "Macropsis fuscinervis" OR "Magdalis nitidipennis" OR "Malacosoma disstria" OR "Malacosoma neustria" OR "Malacosoma parallela" OR "Marasmius favrei" OR "Marssonina castagnei" OR "Marssonina populi" OR "Megaplatypus mutatus" OR "Melampsora aecidioides" OR "Melampsora allii-populina" OR "Melampsora caprearum" OR "Melampsora farinosa" OR "Melampsora laricis-populina" OR "Melampsora laricis-tremulae" OR "Melampsora larici-tremulae" OR "Melampsora magnusiana" OR "Melampsora medusae" OR "Melampsora medusae f. sp. deltoidis" OR "Melampsora mercurialis-tremulae" OR "Melampsora pinitorqua" OR "Melampsora populnea" OR "Melampsora populnea f. sp. laricis" OR "Melampsora populnea f. sp. magnusiana" OR "Melampsora populnea f. sp. pinitorqua" OR "Melampsora populnea f. sp. rostrupii" OR "Melampsora pulcherrima" OR "Melampsora rostrupii" OR "Melampsora tremulae" OR "Melanophila picta" OR "Melanospora fimbriata" OR "Melittosporiella pulchella" OR "Melomastia mastoidea" OR "Membranomyces spurius" OR "Menesia bipunctata" OR "Menispora caesia" OR "Menispora glauca" OR "Menispora libertiana" OR "Menispora tortuosa" OR "Messa glaucopis" OR "Microsphaera penicillata" OR "Microthecium fimbriatum" OR "Monodictys melanopa" OR "Monosteira unicostata" OR "Morimus asper" OR "Mycoporum hippocastani" OR "Mycosphaerella populi" OR "Mycosphaerella punctiformis" OR "Mycosphaerella togashiana" OR "Mycosphaerella tremulicola" OR "Mycterothrips salicis" OR "Mytilinidion gemmigenum" OR "Naeviopsis carneopallida" OR "Napicladium asteroma" OR "Nectria cinnabarina" OR "Nectria dematiosa" OR "Nectria ditissima" OR "Nectria peziza" OR "Necydalis major" OR "Nemania serpens" OR "Nemania serpens var. serpens" OR "Nematus fahraei" OR "Nematus fuscomaculatus" OR "Nematus incompletus" OR "Nematus melanaspis" OR "Nematus nigricornis" OR "Nematus pavidus" OR "Neofusicoccum luteum" OR "Neolygus zebei" OR "Neomecomma bilineatus" OR "Neonectria ditissima" OR "Neta patuxentica" OR "Niesslia exilis" OR "Nivellia sanguinosa" OR "Noctua comes" OR "Noctua fimbriata" OR "Notodonta dromedarius" OR "Notodonta torva" OR "Notodonta tritophus" OR "Notodonta ziczac' OR "Nymphalis antiopa" OR "Nymphalis polychloros" OR "Nymphalis vaualbum" OR "Obrium cantharinum" OR "Oligoporus cerifluus" OR "Operophtera brumata" OR "Operophtera fagata" OR "Ophiostoma tremuloaureum" OR

"Ophiostoma tremulo-aureum" OR "Orbilia rubella" OR "Orchestes jota" OR "Orgya antigua" OR "Orgya recens" OR "Orgyia antiqua" OR "Orgyia leucostigma" OR "Orgyia recens" OR "Orthosia cerasi" OR "Orthosia gracilis" OR "Orthosia incerta" OR "Orthosia munda" OR "Orthosia populeti" OR "Orthotylus bilineatus" OR "Oxyporus corticola" OR "Oxyporus populinus" OR "Pachypappa marsupialis" OR "Pachypappa populi" OR "Pachypappa rosettei" OR "Pachypappa tremulae" OR "Pachypappella lactea" OR "Palaeolecanium bituberculatum" OR "Pamphilius betulae" OR "Pamphilius brevicornis" OR "Pamphilius festivus" OR "Pamphilius histrio" OR "Pamphilius latifrons" OR "Pamphilius maculosus" OR "Pamphilius silvaticus" OR "Pamphilius sylvaticus" OR "Panonychus ulmi" OR "Pappia fissilis" OR "Paraleucoptera sinuella" OR "Paranthrene tabaniformis" OR "Paraphytomyza tremulae" OR "Parastichtis suspecta" OR "Parlatoria oleae" OR "Parthenolecanium corni" OR "Patellariopsis clavispora" OR "Patinellaria sanguinea" OR "Pedostrangalia revestita" OR "Penicillium canescens" OR "Penicillium citrinum" OR "Penicillium purpurogenum var. rubri" OR "Peniophora polygonia" OR "Peniophora rufa" OR "Periconia hispidula" OR "Pestalotiopsis populi-nigrae" OR "Pezicula populi" OR "Phaeoacremonium minimum" OR "Phaeocalicium praecedens" OR "Phaeoramularia maculicola" OR "Phalera bucephala" OR "Phellinus igniarius" OR "Phellinus populicola" OR "Phellinus tremulae" OR "Pheosia tremula" OR "Phigalia pilosaria" OR "Phlebia rufa" OR "Phloeomyzus passerinii" OR "Phloeospora tremulae" OR "Phlyctis argena" OR "Phlyctis erythrosora" OR "Phoma cinerea" OR "Phoma crepini" OR "Phoma tremulae" OR "Phoma urens" OR "Phomatospora dinemasporium" OR "Phomopsis pallida" OR "Phratora atrovirens" OR "Phratora laticollis" OR "Phratora vitellinae" OR "Phratora vulgatissima" OR "Phyllactinia guttata" OR "Phyllactinia populi" OR "Phyllactinia populina" OR "Phyllactinia suffulta" OR "Phyllobius calcaratus" OR "Phyllobius glaucus" OR "Phyllobius maculicornis" OR "Phyllobius pyri" OR "Phyllobius viridiaeris" OR "Phyllocnistis labyrinthella" OR "Phyllocnistis ramulicola" OR "Phyllocnistis xenia" OR "Phyllocoptes didelphis" OR "Phyllocoptes populi" OR "Phyllocoptes populinus" OR "Phyllodecta laticollis" OR "Phyllodecta vitellinae" OR "Phyllodecta vulgatissima" OR "Phyllodesma ilicifolia" OR "Phyllodesma tremulifolia" OR "Phyllonorycter apparella" OR "Phyllonorycter sagitella" OR "Phyllonorycter salicicolella" OR "Phyllonorycter salictella" OR "Phyllosticta alcides" OR "Phyllosticta cinerea" OR "Phyllosticta populea" OR "Phyllosticta populina" OR "Phyllosticta populi-nigrae" OR "Phylloxerina populi" OR "Physatocheila dumetorum" OR "Phytobia cambii" OR "Phytocoris tiliae" OR "Phytodecta decemnotata" OR "Phytodecta viminalis" OR "Picipes tubaeformis" OR "Plagodis dolabraria" OR "Platystomum populinae" OR "Pleurophomopsis salicina" OR "Pleurotheciopsis bramleyi" OR "Pleurotus calyptratus" OR "Pleurotus dryinus" OR "Pleurotus ostreatus" OR "Poecilocampa populi" OR "Poecilonota variolosa" OR "Pollaccia radiosa" OR "Polydrusus cervinus" OR "Polydrusus flavipes" OR "Polydrusus pterygomalis" OR "Polydrusus tereticollis" OR "Polydrusus undatus" OR "Polyporus brumalis" OR "Polyporus dryadeus" OR "Polyporus dryophilus" OR "Polyporus hispidus" OR "Polyporus leptocephalus" OR "Polyporus lipsiensis" OR "Polyporus pseudobetulinus" OR "Polyporus squamosus" OR "Polyporus sulphureus" OR "Polyporus tubaeformis" OR "Polyporus zonatus" OR "Populicerus laminatus" OR "Populicerus populi" OR "Postia ceriflua" OR "Pristiphora conjugata" OR "Psallus confusus" OR "Psallus perrisi" OR "Psallus variabilis" OR "Psallus wagneri" OR "Pseudaulacaspis pentagona" OR "Pseudocamarosporium propinguum" OR "Pseudocercospora togashiana" OR "Pseudochermes fraxini" OR "Pseudoclavellaria amerinae" OR "Pseudococcus comstocki" OR "Pseudoinonotus dryadeus" OR "Pseudoips fagana" OR "Pseudoips prasinana" OR "Pseudomonas syringae pv. syringae" OR "Pseudosciaphila branderiana" OR "Pseudotrichia mutabilis" OR "Pterocomma populeum" OR "Pterocomma tremulae" OR "Pterostoma palpina" OR "Ptilodon capucina" OR "Ptycholoma lecheana" OR "Pulvinaria tremulae" OR "Pulvinaria vitis" OR "Pyrenopeziza petiolaris" OR "Pyrenophora buddleiae" OR "Pyrenophora buddlejae" OR "Rabdophaga giraudiana" OR "Rabdophaga saliciperda" OR "Raduliporus aneirinus" OR "Radulodon erikssonii" OR "Ramphus pulicarius" OR "Ramularia jaczevskii" OR "Ramularia rosea" OR "Resseliella quercivora" OR "Rhagium bifasciatum" OR "Rhagium mordax" OR "Rhamphus pulicarius" OR "Rheumaptera undulata" OR "Rhogogaster chlorosoma" OR "Rhogogaster dryas" OR "Rhogogaster punctulata" OR "Rhogogaster viridis" OR "Rhynchaenus salicis" OR "Rhynchites longiceps" OR "Rhynchites tomentosus" OR "Rhynchostoma minutum" OR "Rhytidiella moriformis" OR "Ropalopus femoratus" OR "Rosellinia subsimilis" OR "Rutidosoma globulus" OR "Rutidosoma graminosum" OR "Saccosoma farinaceum" OR "Saperda carcharias" OR "Saperda perforata" OR "Saperda populnea" OR Saperda scalaris" OR "Saturnia pavonia" OR "Schizophyllum amplum" OR "Schizophyllum commune" OR "Schizotetranychus garmani" OR "Schizotetranychus schizopus" OR "Schizoxylon albescens" OR "Sciota hostilis" OR "Sciota rhenella" OR "Sclerencoelia fascicularis" OR "Scoliopteryx libatrix" OR "Scytinostroma galactinum" OR "Semioscopis strigulana" OR "Septogloeum populiperdum" OR "Septoria marmorata" OR "Septoria populi" OR "Septotinia populiperda" OR "Septotis populiperda" OR "Sesia apiformis" OR "Sesia melanocephala" OR "Smerinthus ocellata" OR "Smerinthus ocellatus" OR "Sphaerulina frondicola" OR "Spilonota ocellana" OR "Sporocadus populinus" OR "Stachybotrys alternans" OR "Stauronematus compressicornis" OR "Stauronematus platycerus" OR "Stauropus fagi" OR "Stegania cararia" OR "Stegania trimaculata" OR "Stegonsporium taphrinum" OR "Stenocorus meridianus" OR "Stenostola dubia" OR "Stenostola ferrea" OR "Sthenarus rotermundi" OR "Stictis brunnescens" OR "Stictis confusa" OR "Stictis populorum" OR "Stictochorella populinigrae" OR "Stigmella assimilella" OR "Stigmella trimaculella" OR "Stomaphis longirostris" OR "Strangalia attenuata" OR "Strangalia aurulenta" OR "Strangalia maculata" OR "Subacronicta megacephala" OR "Sympodiella acicola" OR "Synanthedon formicaeformis" OR "Synanthedon melliniformis" OR "Synanthedon spuleri" OR "Syndemis musculana" OR "Tachyerges rufitarsis" OR "Tachyerges salicis" OR "Talaromyces purpureogenus" OR "Tapesia cinerella" OR "Taphrina johansonii" OR "Taphrina populina" OR "Taphrina rhizophora" OR "Taphrorychus bicolor" OR "Tectella calyptrata" OR "Teichospora abducens" OR "Teichospora pruniformis" OR "Temnocerus coeruleus" OR "Temnocerus longiceps" OR "Tethea ocularis" OR "Tethea or" OR "Tetheella fluctuosa" OR "Tetranychus turkestani" OR "Tetranychus urticae" OR "Tetropium castaneum" OR "Thecabius affinis" OR "Thyridaria macrostomoides" OR "Titaeosporina tremulae" OR "Tomentella asperula" OR "Trachypteris picta" OR "Trachypteris picta decostigma" OR "Trachys minutus" OR "Tracylla julia" OR "Trametes cervina" OR "Trametes gibbosa" OR "Trametes ochracea" OR "Trametes pubescens" OR "Trametes trogii" OR "Trametes versicolor" OR "Trametopsis cervina" OR "Trematosphaeria pertusa" OR "Tremex fuscicornis" OR "Tremulicerus fulgidus" OR "Tremulicerus tremulae" OR "Trichiocampus grandis" OR "Trichiosoma pusillum" OR "Trichiura crataegi" OR "Trichoderma lignorum" OR "Trichoderma strictipile" OR "Trichoderma viride" OR "Trichoferus campestris" OR "Trichopeziza karstenii" OR "Trichopteryx carpinata" OR "Trichothecium roseum" OR "Triposporium elegans" OR "Tritophia tritophus" OR "Troposporella fumosa" OR "Trypodendron domesticum" OR "Trypophloeus asperatus" OR "Trypophloeus bispinulus" OR "Trypophloeus granulatus" OR "Trypophloeus tremulae" OR "Tympanis alpina" OR "Tympanis spermatiospora" OR "Typhula ochraceosclerotiata" OR "Tyromyces fissilis" OR "Tyromyces fumidiceps" OR "Tyromyces vivii" OR "Uncinula adunca" OR "Uncinula adunca var. adunca" OR

"Uncinula salicis" OR "Ustulina vulgaris" OR "Valsa ambiens" OR "Valsa nivea" OR "Valsa sordida" OR "Valsaria anserina" OR "Valsella nigroannulata" OR "Venturia macularis" OR "Venturia maculiformis" OR "Venturia martianoffiana" OR "Venturia populina" OR "Venturia radiosa" OR "Venturia tremulae" OR "Venturia tremulae var. tremulae" OR "Venturia viennotii" OR "Verticillium alboatrum" OR "Verticillium albo-atrum" OR "Viridicerus ustulatus" OR "Vuilleminia comedens" OR "Xanthia icteritia" OR "Xanthia ocellaris" OR "Xanthomonas populi" OR "Xenasma rimicola" OR "Xenosporium pleurococcum" OR "Xestia castanea" OR "Xiphydria camelus" OR "Xylaria hypoxylon" OR "Xyleborinus attenuatus" OR "Xyleborus cryptographus" OR "Xyleborus dispar" OR "Xyleborus pfeili" OR "Xylella fastidiosa" OR "Xylosandrus crassiusculus" OR "Xylosandrus germanus" OR "Xylotrechus rusticus" OR "Ypsolopha parenthesella" OR "Ypsolopha ustella" OR "Zeugophora flavicollis" OR "Zeugophora frontalis" OR "Zeugophora scutellaris" OR "Zeugophora subspinosa" OR "Zeugophora turneri" OR "Zeuzera pyrina" OR "Zignoella ovoidea" OR "Zygina nivea"

APPENDIX C

Plant taxa reported to be present in the nurseries of Populus alba, P. nigra and P. tremula

TABLE C.1 Plant taxa reported in the Dossier Sections 3.1, 3.2 and 3.3 to be present in the nurseries of *Populus alba*, *P. nigra* and *P. tremula*.

Number	Plant taxa	Number	Plant taxa
1	Abelia	610	Malus 'Scotch Dumpling'
2	Abies alba	611	Malus 'Scrumptious'
3	Abies fraserii	612	Malus 'Somerset Redstreak'
4	Abies grandis	613	Malus 'Spartan'
5	Abies nobilis	614	Malus 'St Edmund's Russet'
6	Abies nordmanniana	615	Malus 'Stirling Castle'
7	Acacia	616	Malus 'Stoke Red'
8	Acanthus	617	Malus 'Sun Rival'
9	Acer	618	Malus 'Sunset'
10	Acer campestre	619	Malus 'Surprize'
11	Acer macrocarpa	620	Malus sylvestris
12	Acer palmatum 'Pixie'	621	Malus 'Three Counties'
13	Acer palmatum 'Sango kaku'	622	Malus 'TICKLED PINK Baya Marisa'
14	Acer palmatum 'Seiryu'	623	Malus 'Tom Putt'
15	Acer palmatum 'Shaina'	624	Malus toringo subsp. sargentii 'Tina'
16	Acer palmatum 'Suminagashi'	625	Malus transitoria
17	Acer palmatum 'Tamukeyama'	626	Malus transitoria 'Thornhayes Tansy'
18	Acer palmatum 'Trompenburg'	627	Malus 'Tremlett's Bitter'
19	Acer palmatum 'Villa Taranto'	628	Malus trilobata 'Guardsman'
20	Acer platanoides	629	Malus 'Trinity'
21	Acer platanoides 'Crimson King'	630	Malus tschonoskii
22	Acer platanoides 'Crimson Sentry'	631	Malus tschonoskii 'Belmonte'
23	Acer platanoides 'Drummondii'	632	Malus 'Van Eseltine'
24	Acer platanoides 'Princeton Gold'	633	Malus 'Vicky'
25	Acer pseudoplatanus	634	Malus 'Warner's King'
26	Acer pseudoplatanus 'Brilliantissimum'	635	Malus 'William Crump'
27	Acer pseudoplatanus 'Esk Sunset'	636	Malus 'Winter Gem'
28	Acer pseudoplatanus 'Leopoldii'	637	Malus 'Worcester Pearmain'
29	Acer pseudoplatanus 'Prinz Handjery'	638	<i>Malus×moerlandsii</i> 'Profusion Improved'
30	Acer rubrum	639	Malus 'Yarlington Mill'
31	Acer rubrum 'Autumn Flame'	640	Matteuccia
32	Acer rubrum 'Brandywine'	641	Meconopsis
33	Acer rubrum 'October Glory'	642	Mespilus 'Nottingham'
34	Acer rubrum 'Red Sunset'	643	Metasequoia glyptostroboides
35	Acer rubrum 'Scanlon'	644	Miscanthus
36	Acer rubrum 'Sun Valley'	645	Molinia
37	Acer saccharum	646	Monarda
38	Acer shirasawanum 'Autumn Moon'	647	Morus 'Carman'
39	Acer×freemanii 'Autumn Blaze'	648	Morus 'Chelsea'
40	Acer×freemanii 'Morgan'	649	Morus 'Giant Fruit'
41	Achillea	650	Morus 'Mojo Berry'
42	Acorus	651	Morus 'Pendula'
43	Actaea	652	Myrtus
44	Aesculus × carnea 'Briotii'	653	Nandina
45	Aesculus parviflora	654	Nemesia
46	Agapanthus	655	Nepeta

(Continues)

TABLE C.1 (Continued)				
Number	Plant taxa	Number	Plant taxa	
47	Agastache	656	Nothofagus	
48	Ajuga	657	Nothofagus antarctica	
49	Akebia	658	Nyssa sylvatica	
50	Albizia julibrissin 'Chocolate Fountain'	659	Nyssa sylvatica 'Red Rage'	
51	Albizia julibrissin 'Evys Pride"	660	Nyssa sylvatica 'Wisley Bonfire'	
52	Albizia julibrissin 'Ombrella'	661	Olearia	
53	Albizia julibrissin 'Shidare'	662	Ophiopogon	
54	Albizia julibrissin 'Summer Chocolate'	663	Osmanthus	
55	Alchemilla	664	Osmunda	
56	Allium	665	Pachysandra	
57	Alnus	666	Pachystegia	
58	Alnus cordata	667	Paeonia	
59	Alnus glutinosa	668	Panicum	
60	Alnus glutinosa 'Imperialis'	669	Parrotia persica	
61	Alnus incana	670	Parrotia persica 'Bella'	
62	Alnus incana 'Aurea'	671	Parrotia persica 'Persian Spire'	
63	Alnus rubra	672	Parrotia persica 'Vanessa'	
64	Alnus spaethii	673	Paulownia tomentosa	
65	Alstroemeria	674	Pennisetum	
66	Amelanchier	675	Penstemon	
67	Amelanchier alnifolia 'Northline'	676	Perovskia	
68	Amelanchier alnifolia 'Obelisk'	677	Persicaria	
69	Amelanchier canadensis 'Rainbow Pillar'	678	Philadelphus	
70	Amelanchier 'Edelweiss'	679	Phlomis	
71	Amelanchier 'La Paloma'	680	Phlox	
72	Amelanchier laevis 'R.J. Hilton'	681	Phormium	
73	Amelanchier laevis 'Snowflakes'	682	Photinia	
74	Amelanchier lamarckii	683	<i>Photinia×fraseri</i> 'Red Robin'	
75	Amelanchier×grandiflora 'Ballerina'	684	Phygelius	
76	Amelanchier×grandiflora 'Robin Hill'	685	Physocarpus	
77	Ammonophylla	686	Physocarpus opulifolius 'Diablo'	
78	Anemanthele	687	Physocarpus opulifolius 'Lady in Red'	
79	Anemone	688	Physostegia	
80	Aquilegia	689	Picea abies	
81	Araucaria araucana	690	Picea orientalis	
82	Arbutus	691	Picea ormorika	
83	Arbutus unedo	692	Picea pungens 'Erich Frahm'	
84	Armeria	693	Picea pungens 'Iseli Fastigiate'	
85	Artemisia	694	Picea sitchensis	
86	Arum	695	Picea smithiana 'Aurea'	
87	Aruncus	696	Pinus	
88	Asplenium	697	Pinus densiflora 'Umbraculifera'	
89	Astelia	698	Pinus flexilis 'Vanderwolf's Pyramid'	
90	Aster	699	Pinus mugo 'Winter Sun'	
91	Astilbe	700	Pinus nigra 'Bright Eyes'	
92	Astrantia	701	Pinus nigra 'Obelisk'	
93	Athyrium	702	Pinus peuce	
94	Aucuba	703	Pinus pinaster	
95	Baptisia	704	Pinus pungens glauca	

TABLE C.1 (Continued)					
Number	Plant taxa	Number	Plant taxa		
96	Berberis	705	Pinus radiata 'Aurea'		
97	Bergenia	706	Pinus strobus 'Minima'		
98	Betula	707	Pinus strobus 'Tiny Kurls'		
99	Betula alba pendula	708	Pinus sylvestris		
100	Betula albosinensis 'Chinese Ruby'	709	Pinus sylvestris 'Chantry Blue'		
101	Betula costata 'Daleside'	710	Pinus sylvestris 'Gold Medal'		
102	Betula ermanii 'Mount Zao Purple'	711	Pinus sylvestris 'Westonbirt'		
103	Betula ermanii 'Polar Bear'	712	Pinus thunbergii 'Banshosho'		
104	Betula ermanii 'White Chocolate'	713	Pinus wallichiana		
105	Betula 'Fascination'	714	Pinus×holdfordiana		
106	Betula 'Fetisowii'	715	Pittosporum		
107	Betula nigra 'Shiloh Splash'	716	Platanus		
108	Betula pendula	717	Platanus×hispanica		
109	Betula pendula 'Dalecarlica'	718	Polemonium		
110	Betula pendula 'Fastigiata Joes'	719	Polygonatum		
111	Betula pendula 'Royal Frost'	720	Polypodium		
112	Betula pendula 'Spider Alley'	721	Polystichum		
113	Betula pendula 'Tristis'	722	Populus		
114	Betula pendula 'Youngii'	723	Populus nigra		
115	Betula pubescens	724	Populus tremula		
116	Betula utilis 'Cinnamon'	725	Potentilla		
117	Betula utilis 'Dark-Ness'	726	Primula		
118	Betula utilis 'Edinburgh'	727	Prunus		
119	Betula utilis 'Melony Sanders'	728	Prunus 'Accolade'		
120	Betula utilis 'Moonbeam'	729	Prunus 'Amanogawa'		
121	Betula utilis 'Mount Luoji'	730	Prunus 'Amber Heart'		
122	Betula utilis 'Snow Queen'	731	Prunus 'Amsden June'		
123	Betula utilis ssp. Jacquemontii	732	Prunus 'Aprikyra'		
124	Betula utilis ssp. jacquemontii 'Grayswood Ghost'	733	Prunus 'Aprimira'		
125	Betula utilis ssp. jacquemontii 'Jermyns'	734	Prunus 'Aprisali'		
126	Betula utilis ssp. Jacquemontii 'Silver Shadow'	735	Prunus 'Areko'		
127	Betula utilis ssp. jacquemontii 'Trinity College'	736	Prunus 'Asano'		
128	Betula utilis subsp. albosinensis 'Cacao'	737	Prunus 'Athos'		
129	Betula utilis subsp. albosinensis 'China Rose'	738	Prunus 'Avalon'		
130	Betula utilis subsp. albosinensis 'Hergest'	739	Prunus 'Avalon Pride'		
131	Betula utilis subsp. albosinensis 'Kansu'	740	Prunus avium		
132	Betula utilis subsp. albosinensis 'Pink Champagne'	741	Prunus avium 'Plena'		
133	Betula utilis subsp. albosinensis 'Red Panda'	742	Prunus 'Aylesbury Prune'		
134	Betula utilis subsp. jacquemontii 'McBeath'	743	Prunus 'Belle de Louvain'		
135	Betula utilis 'Wakehurst Place Chocolate'	744	Prunus 'Beni-yutaka'		
136	Blechnum	745	Prunus 'Bergeron'		
137	Brachyglottis	746	Prunus 'Bergeval'		
138	Brunnera	747	Prunus 'Black Oliver'		
139	Buddleja	748	Prunus 'Blaisdon Red'		
140	Buxus	749	Prunus 'Blue Tit'		
141	Buxus sempervirens	750	Prunus 'Blushing Bride'		
142	Calamagrostis	751	Prunus 'Burcombe'		
143	Callicarpa bodinieri var. giraldii 'Profusion'	752	Prunus 'Cambridge'		
144	Calluna	753	Prunus 'Candy Floss'		

TABLE C.1	(Continued)		
Number	Plant taxa	Number	Plant taxa
145	Calycanthus 'Aphrodite'	754	Prunus 'Catherine'
146	Campanula	755	Prunus 'Celeste'
147	Carex	756	Prunus cera
148	Carpinus	757	Prunus cera 'Crimson Pointe'
149	Carpinus betulus	758	Prunus cera 'Nigra'
150	Carpinus betulus 'Chartreuse'	759	Prunus cerasifera myrobalan
151	Carpinus betulus 'Frans Fontaine'	760	Prunus 'Chocolate Ice'
152	Carpinus betulus 'Lucas'	761	Prunus 'Coes Golden Drop'
153	Carpinus betulus 'Rockhampton Red'	762	Prunus 'Collingwood Ingram'
154	Caryopteris	763	Prunus 'Compacta'
155	Castanea	764	Prunus 'Countess'
156	Castanea sativa	765	Prunus 'Czar'
157	Catalpa bignonioides 'Aurea'	766	Prunus 'Daikoku'
158	Catalpa×erubescens 'Purpurea'	767	Prunus 'de Nancy'
159	Ceanothus	768	Prunus 'Denniston's Superb'
160	Ceanothus arboreus 'Trewithen Blue'	769	Prunus 'Early Red Maraly'
161	Cedrus atlantica	770	Prunus 'Early Transparent'
162	Cedrus atlantica 'Glauca'	771	Prunus 'Edda'
163	Cedrus atlantica 'Glauca Pendula'	772	Prunus 'Excalibur'
164	Cedrus deodara 'Karl Fuchs'	773	Prunus 'Farleigh'
165	Cedrus deodara 'Klondyke'	774	Prunus 'Ferbleue'
166	Cedrus libani	775	Prunus 'Fertile'
167	Centaurea	776	Prunus 'Fice'
168	Centranthus	777	Prunus 'Flavor King'
169	Ceratostigma	778	Prunus 'Folfer'
170	Cercidiphyllum japonicum	779	Prunus 'Fragrant Cloud'
171	Cercidiphyllum japonicum 'Pendulum'	780	Prunus 'Frilly Frock'
172	Cercis canadensis 'Alley Cat'	781	Prunus 'Fugenzo'
173	Cercis canadensis 'Carolina Sweetheart'	782	Prunus 'Garden Aprigold'
174	Cercis canadensis 'Eternal Flame'	783	Prunus 'Garden Beauty'
175	Cercis canadensis 'Forest Pansy'	784	Prunus 'Garden Lady'
176	Cercis canadensis 'Golden Falls'	785	Prunus 'Goldcot'
177	Cercis canadensis 'Hearts of Gold'	786	Prunus 'Golden Glow'
178	Cercis canadensis 'Lavender Twist'	787	Prunus 'Golden Sphere'
179	Cercis canadensis 'Merlot'	788	Prunus 'Gordon Castle'
180	Cercis canadensis 'Pink Pom Pom'	789	Prunus 'Gorgeous'
181	Cercis canadensis 'Rising Sun'	790	Prunus 'Guinevere'
182	Cercis canadensis 'Ruby Falls'	791	Prunus 'Gyoiko'
183	Cercis canadensis 'Vanilla Twist'	792	Prunus 'Gypsy'
184	Cercis chinensis 'Avondale'	793	Prunus 'Haganta'
185	Cercis chinensis 'Diane'	794	Prunus 'Hales Early'
186	Cercis reniformis 'Oklahoma'	795	Prunus 'Hally Jolivette'
187	Cercis reniformis 'Texan White'	796	Prunus 'HELENA DU ROUSSILLON Aviera'
188	Cercis siliquastrum 'Bodnant'	797	Prunus 'Henriette'
189	Chaenomeles	798	Prunus 'Herman'
190	Chamaecyparis	799	Prunus 'Hertford'
191	Choisya	800	Prunus 'Hokusai'
192	Cistus	801	Prunus 'Horinji'
193	Cladrastis kentuckea	802	Prunus 'Ichiyo'
TABLE C.1 (Continued)

Number	Plant taxa	Number	Plant taxa		
194	Clematis	803	Prunus incisa 'Kojo-no-mai'		
195	Convolvulus	804	Prunus incisa 'Mikinori'		
196	Coprosma	805	Prunus incisa 'Oshidori PRINCESSE'		
197	Coreopsis	806	Prunus incisa 'Pendula'		
198	Cornus	807	Prunus incisa 'Praecox'		
199	Cornus sanguinea	808	Prunus incisa 'Yamadei'		
200	Cortaderia	809	Prunus 'Ingrid'		
201	Corydalis	810	Prunus 'Jacqueline'		
202	Corylus	811	Prunus 'Jefferson'		
203	Corylus avellana	812	Prunus 'Jubilee'		
204	Corylus avellana 'Contorta'	813	Prunus 'Kanzan'		
205	Corylus 'Cosford'	814	Prunus 'Katinka'		
206	Corylus 'Gunslebert'	815	Prunus 'Ki 2004 R11 B93'		
207	Corylus 'Hall's Giant'	816	Prunus 'Ki 2004 R14 B56'		
208	Corylus 'Lang Tidlig Zeller'	817	Prunus 'Kiku-shidare-zakura'		
209	Corylus 'Nottingham'	818	Prunus 'King of the Damsons'		
210	Corylus 'Red Filbert'	819	Prunus 'Kioto'		
211	Corylus 'Te-Terra Red'	820	Prunus 'KIR LAMOUR'		
212	Corylus 'Tonda Di Giffoni'	821	Prunus 'KIR ROSSO'		
213	Corylus 'Tonda Gentile de le Romana'	822	Prunus 'KIR VULCANO'		
214	Corylus 'Tonda Gentile Trilobata'	823	Prunus 'Knights Early Black'		
215	Corylus 'Webbs Prize Cob'	824	Prunus 'Kobuku-zakura POWDER PUFF'		
216	Cosmos	825	Prunus 'Kofugen'		
217	Cotinus	826	Prunus 'Kordia'		
218	Cotoneaster	827	Prunus 'Kursar'		
219	Cotoneaster frigidus 'Cornubia'	828	Prunus 'Lapins Cherokee'		
220	Cotoneaster 'Hybridus Pendulus'	829	Prunus laurocerasus		
221	Cotoneaster lacteus	830	Prunus 'Lindsey Gage'		
222	Cotoneaster salicifolius 'Exburiensis'	831	Prunus litigiosa		
223	Cotoneaster salicifolius 'Repens'	832	Prunus 'Little Pink Perfection'		
224	Cotoneaster × suecicus 'Coral Beauty'	833	Prunus 'Lord Napier'		
225	Cotoneaster × suecicus 'Juliette'	834	Prunus lusitanica		
226	Crataegus	835	Prunus 'Malling Elizabeth'		
227	Crataegus azarolus	836	Prunus 'Marjorie's Seedling'		
228	Crataegus laevigata 'Crimson Cloud'	837	Prunus 'Merchant'		
229	Crataegus laevigata 'Pauls Scarlet'	838	Prunus 'Meritare'		
230	Crataegus laevigata 'Plena'	839	Prunus 'Merryweather'		
231	Crataegus laevigata 'Rosea Flore Pleno'	840	Prunus 'Merton Glory'		
232	Crataegus monogyna	841	Prunus 'Mesembrine'		
233	Crataegus monogyna 'Stricta'	842	Prunus 'Mikurama-gaeshi'		
234	Crataegus persimilis 'Prunifolia Splendens'	843	Prunus 'Morello'		
235	Crataegus pinnatifida var. major 'Big Golden Star'	844	Prunus 'Nabella'		
236	Crataegus schraderiana	845	Prunus 'Napoleon Bigarreau'		
237	Crataegus succulenta 'Jubilee'	846	Prunus 'Nectarella'		
238	Crataegus×dippeliana	847	Prunus 'Nimba'		
239	Crataegus×lavallei 'Carrierei'	848	Prunus 'Okame'		
240	Crocosmia	849	Prunus 'Old Green Gage'		
241	Cryptomeria japonica	850	Prunus 'Opal'		
242	Cryptomeria japonica 'Gracilis'	851	Prunus 'Oullins Golden'		

TABLE C.1	(Continued)		
Number	Plant taxa	Number	Plant taxa
243	Cryptomeria japonica 'Sekkan-sugi'	852	Prunus padus
244	Cupressocyparis	853	Prunus padus 'Le Thoureil'
245	Cupressocyparis leylandii	854	Prunus 'Pandora'
246	Cupressus	855	Prunus 'Papillon'
247	Cupressus arizonica var. glabra 'Blue Ice'	856	Prunus pendula 'Ascendens Rosea'
248	Cupressus macrocarpa 'Wilma'	857	Prunus pendula 'Pendula Rubra'
249	Cupressus sempervirens 'Totem'	858	Prunus pendula 'Stellata'
250	Cydonia 'Aromatnaya'	859	Prunus 'Penny'
251	Cydonia 'Bereczki'	860	Prunus 'Peregrine'
252	Cydonia 'Isfahan'	861	Prunus 'Petit Noir'
253	Cydonia 'Meech's Prolific'	862	Prunus 'Pineapple'
254	Cydonia 'Serbian Gold'	863	Prunus 'Pink Marry'
255	Cydonia 'Vranja'	864	Prunus 'Pink Parasol'
256	Cynoglossum	865	Prunus 'Pink Perfection'
257	Cytisus	866	Prunus 'Pink Shell'
258	Dahlia	867	Prunus 'Purple Pershore'
259	Daphne	868	Prunus 'Queen's Crown'
260	Davidia involucrata	869	Prunus 'Red Haven'
261	Davidia involucrata 'Sonoma'	870	Prunus 'Reeves'
262	Delosperma	871	Prunus 'Regina'
263	Delphinium	872	Prunus 'Reine Claude de Bavay'
264	Deschampsia	873	Prunus 'River's Early Prolific'
265	Deutzia	874	Prunus 'Robada'
266	Dicentra	875	Prunus 'Robijn'
267	Diervilla	876	Prunus 'Rochester'
268	Digitalis	877	Prunus 'Roundel Heart'
269	Doronicum	878	Prunus 'Royal Burgundy'
270	Dryopteris	879	Prunus 'Royal Flame'
271	Echinacea	880	Prunus 'Ruby COLUMNAR'
272	Echinops	881	Prunus rufa
273	Elaeagnus	882	Prunus 'Sanctus Hubertus'
274	Elaeagnus angustifolia 'Quicksilver'	883	Prunus sargentii
275	Epimedium	884	Prunus 'Saturn'
276	Eremurus	885	Prunus 'Seneca'
277	Erigeron	886	Prunus serrula
278	Eriophorum	887	Prunus serrula 'Branklyn'
279	Eriostemon	888	Prunus 'Shepherds Bullace'
280	Eryngium	889	Prunus 'Shirotae'
281	Erysimum	890	Prunus 'Shosar'
282	Escallonia	891	Prunus 'Shropshire Prune'
283	Eucalyptus	892	Prunus 'Skeena'
284	Eucalyptus 'Azura'	893	Prunus 'Snow Goose'
285	Eucalyptus glaucescens	894	Prunus 'Snow Showers'
286	Eucalyptus gunnii	895	Prunus spinosa
287	Euonymus	896	Prunus 'Spire'
288	Euonymus alatus 'Compactus'	897	Prunus 'Spring Snow'
289	Euonymus clivicola	898	Prunus 'STARDUST COVEU'
290	Euonymus europaeus	899	Prunus 'Stella'
291	Euonymus europaeus 'Brilliant'	900	Prunus 'Stella's Star'

TABLE C.1	(Continued)						
Number	Plant taxa	Number	Plant taxa				
292	Euonymus europaeus 'Red Cascade'	901	Prunus 'Summer Sun'				
293	Euonymus hamiltonianus 'Indian Summer'	902	Prunus 'Sunburst'				
294	Euonymus hamiltonianus 'Koi Boy'	903	Prunus 'Sunset Boulevard'				
295	Euonymus phellomanus	904	Prunus 'Swan'				
296	Euonymus planipes	905	Prunus 'Sweet Prune'				
297	Euonymus planipes 'Sancho'	906	Prunus 'Sweetheart'				
298	Euphorbia	907	Prunus 'Sylvia'				
299	Exochorda	908	Prunus 'Tai-haku'				
300	Exochorda×macrantha 'The Bride'	909	Prunus 'Taoyame'				
301	Fagus	910	Prunus 'Terrace Amber'				
302	Fagus sylvatica	911	Prunus 'The Bride'				
303	Fagus sylvatica 'Black Swan'	912	Prunus 'Tiltstone Hellfire'				
304	Fagus sylvatica 'Dawyck'	913	Prunus 'Tomcot'				
305	Fagus sylvatica 'Dawyck Gold'	914	Prunus 'Topend Plus'				
306	Fagus sylvatica 'Dawyck Purple'	915	Prunus 'Topfive'				
307	Fagus sylvatica 'Midnight Feather'	916	Prunus 'Tophit Plus'				
308	Fagus sylvatica 'Pendula'	917	Prunus 'Toptaste Kulinaria'				
309	Fagus sylvatica 'Purple Fountain'	918	Prunus 'Trailblazer'				
310	Fagus sylvatica 'Purpurea'	919	Prunus 'Ukon'				
311	Fagus sylvatica 'Purpurea Pendula'	920	Prunus 'Vanda'				
312	Fagus sylvatica 'Purpurea Tricolor'	921	Prunus 'Victoria'				
313	Fagus sylvatica 'Riversii'	922	Prunus 'Violet'				
314	Fagus sylvatica var. heterophylla 'Aspleniifolia'	923	Prunus 'Walter'				
315	Fagus sylvestris 'Atropurpurea'	924	Prunus 'Warwickshire Drooper'				
316	Fargesia	925	Prunus 'Waterloo'				
317	Fatsia	926	Prunus 'Weeping Yoshino'				
318	Festuca	927	Prunus 'Willingham'				
319	Ficus 'Brown Turkey'	928	Prunus×persicoides 'Spring Glow'				
320	Ficus 'Dalmatie'	929	Prunus×subhirtella 'Autumnalis'				
321	Ficus 'Ice Crystal'	930	Prunus×subhirtella 'Autumnalis Rosea'				
322	Ficus 'Little Miss Figgy'	931	Prunus×subhirtella 'Pendula Plena Rosea'				
323	Ficus 'Panache'	932	Prunus×yedoensis				
324	Filipendula	933	Prunus 'Yellow Pershore'				
325	Foeniculum	934	Pseudotsuga menziesii				
326	Forsythia	935	Pulmonaria				
327	Forsythia × intermedia 'Lynwood Variety'	936	Pyracantha				
328	Forsythia suspensa 'Nymans'	937	Pyrus				
329	Fraxinus ornus 'Obelisk'	938	Pyrus 'Barnet'				
330	Fuchsia	939	Pyrus 'Benita Rafzas'				
331	Galium	940	Pyrus 'Beth'				
332	Garrya	941	Pyrus 'Beurre Hardy'				
333	Gaura	942	Pyrus 'Beurre Superfin'				
334	Genista	943	Pyrus 'Black Worcester'				
335	Geranium	944	Pyrus 'Blakeney Red'				
336	Geum	945	Pyrus 'Brandy'				
337	Ginkgo biloba	946	Pyrus calleryana 'Chanticleer'				
338	<i>Ginkgo biloba</i> 'Blagon'	947	Pyrus 'Catillac'				
339	Ginkgo biloba 'Menhir'	948	Pyrus 'Celebration NUVAR'				
340	Gleditsia triacanthos 'Sunburst'	949	Pyrus 'Christie'				

Number	Plant taxa	Number	Plant taxa		
341	Griselinia	950	Pyrus communis		
342	Hakonechloa	951	Pyrus 'Concorde'		
343	Halesia carolina	952	Pyrus 'Concorde'/'Conference'/'Comice'		
344	Halimium	953	Pvrus 'Conference'		
345	Hamamelis×intermedia 'Arnold Promise'	954	Pyrus 'Conference Moors Giant'		
346	Hamamelis × intermedia 'Diane'	955	Pyrus 'Conference'/'Comice'/'Williams'		
347	Hamamelis × intermedia 'Jelena'	956	Pyrus 'Dovenne du Comice'		
348	Hamamelis × intermedia 'Pallida'	957	Pyrus elaegarifolia 'Silver Sails'		
349	Нере	958	Pyrus 'Fondante d'Automne'		
350	Hedera	959	Pvrus 'Gin'		
351	Helenium	960	Pyrus 'Glou Morceau'		
352	Helichrysum	961	Pvrus 'Gorham'		
353	Helleborus	962	Pyrus 'Green Horse'		
354	Hemerocallis	963	Pyrus 'Hellens Farly'		
355	Hentacodium miconioides	964	Pyrus 'Hendre Huffcan'		
356	Heychera	965	Pyrus 'Humbua'		
357	Heucherella	966	Pyrus 'Invincible delwinor fertilia'		
358	Hinnonhae	967			
359	Hoheria sexstylosa 'Spow White'	968	Pyrus 'Josephine de Malines'		
360	Hosta	969	Pyrus 'Judge Amphlet'		
361	Houttuvnia	970			
362	Hydranaea	970	Pyrus 'Louise Bonne of Jersey'		
363	Hypericum	971	Pyrus 'Merton Pride'		
364	Iberic	972			
265	llex	975			
303	llex	974			
267		975	Pyrus Olympic		
369	llev aquifolium (Alaska)	970			
360	llex aquifolium (Argentes Marginate)	977			
270	lex aquifolium Argentea Marginata	970	Pyrus Pitmaston Dutchess		
370		979			
371	llex aquionum J.C. Van Tol	980	Pyrus Sensation		
372		981	Pyrus Sensation		
3/3	Imperata	982			
374		983			
3/5	Jasminum	984	Pyrus Inorn		
376		985	Pyrus Williams Bon Chretien		
3//	Jugians Broadview	986	Pyrus Winnal's Longdon		
3/8	Jugians Buccaneer	987	Pyrus Winter Nells		
379	Jugians Chandler	988			
380	Juglans 'Fernette'	989	Quercus		
381	Juglans 'Fernor'	990	Quercus ilex		
382	Jugians Franquette	991	Quercus myrsinifolia		
383	Juglans 'Mars'	992	Quercus palustris 'Pringreen'		
384	Juglans nigra	993	Quercus petraea		
385	Juglans regia	994	Quercus robur		
386	Juniperus	995	Quercus rubra		
387	Juniperus communis	996	Quercus texana 'New Madrid'		
388	Juniperus scopulorum 'Blue Arrow'	997	Quercus×warei 'Regal Prince'		
389	Knautia	998	Rhamnus		

TABLE C.1	(Continued)						
Number	Plant taxa	Number	Plant taxa				
390	Kniphofia	999	Rheum 'Strawberry Surprise'				
391	Koelreuteria paniculata 'Coral Sun'	1000	Rheum 'Timperley Early'				
392	Laburnum	1001	Rheum 'Victoria'				
393	Laburnum anagyroides 'Yellow Rocket'	1002	Rhus				
394	Lamium	1003	Ribes				
395	Larix	1004	Ribes 'Ben Connan'				
396	Larix×decidua	1005	Ribes 'Ben Sarek'				
397	Larix×eurolepsis	1006	Ribes 'Black 'n' Red Premiere'				
398	Lavandula	1007 <i>Ribes</i> 'Blackbells'					
399	Lavatera	1008	Ribes 'Blanka'				
400	Leucanthemum	1009	Ribes 'Captivator'				
401	Leucothoe	1010	<i>Ribes</i> 'Hinnonmaki Red'				
402	Leycesteria	1011	Ribes 'Hinnonmaki Yellow'				
403	Leymus	1012	Ribes 'Invicta'				
404	Liatris	1013	Ribes 'Jonkheer van Tets'				
405	Ligularia	1014	Ribes 'Junifer'				
406	Ligustrum	1015	Ribes 'Lowberry Little Black Sugar'				
407	Ligustrum ovalifolium	1016	Ribes 'Mucurines'				
408	Ligustrum vulgare	1017	Ribes 'Ojebyn'				
409	Liquidambar	1018	Ribes 'Rovada'				
410	Liquidambar styraciflua	1019	Ribes 'Titania'				
411	Liquidambar styraciflua 'Lane Roberts'	1020	Robinia				
412	Liquidambar styraciflua 'Palo Alto'	1021	<i>Robinia</i> × <i>margaretta</i> 'Pink Cascade'				
413	Liquidambar styraciflua 'Slender Silhouette'	1022	Robinia pseudoacacia 'Frisia'				
414	Liquidambar styraciflua 'Stared'	1023	<i>Robinia pseudoacacia '</i> Lace Lady Twisty Babe'				
415	Liquidambar styraciflua 'Worplesdon'	1024	Rosa				
416	Liriodendron tulipifera	1025	Rosa canina				
417	Liriodendron tulipifera 'Snow Bird'	1026	Rosa rugosa				
418	Liriope	1027	Rosmarinus				
419	Lithodora	1028	Rubus 'Allgold'				
420	Lobelia	1029	Rubus 'Arapaho'				
421	Lonicera	1030	Rubus 'Autumn Bliss'				
422	Lonicera nitida	1031	Rubus 'Buckingham'				
423	Lonicera periclymenum	1032	Rubus 'Cascade Delight'				
424	Lupinus	1033	Rubus 'Glen Ample'				
425	Luzula	1034	Rubus 'Glen Carron'				
426	Lycium barbarum 'Lubera Instant Success'	1035	Rubus 'Golden Everest'				
427	Lysimachia	1036	Rubus 'Joan J'				
428	Magnolia Magnolia	1037	Rubus Loch Ness				
429	Magnolla Aphrodite	1038	Rubus Lowberry Goodasgoid				
430	Magnolia 'Blue One'	1039	Rubus 'Lowberry Little Black Prince'				
431		1040	Rubus (Malling Lung)				
432	Magnolia (Doobno)	1041	Rubus (Navaha Summarland)				
455	Magnolia (Daybroak)	1042	Rubus (Octavia)				
435	Magnolia 'Eckimo'	1044	Rubus (Oregon Thornless)				
436	Magnolia 'Eairy Blush'	1045	Rubus 'Thornfree'				
437	Magnolia 'Fairy Cream'	1046	Rubus 'Tulameen'				
-							

(Continues)

```
  TABLE C.1 (Continued)
```

Number	Plant taxa	Number	Plant taxa		
438	Magnolia 'Eairy White'	1047	Rudbeckia		
439	Magnolia 'Felix Jury'	1048	Salix		
440	Magnolia 'Galaxy'	1049	Salix aurita		
441	Maanolia 'Genie'	1050	Salix caprea		
442	Maanolia 'Golden Pond'	1051	Salix caprea 'Pendula'		
443	Magnolia arandiflora 'Alta'	1052	Salix cinerea		
444	Magnolia grandiflora 'Kay Parris'	1053	Salix erythroflexuosa 'Golden Curls'		
445	Magnolia 'Heaven Scent'	1054	Salix 'Hakuro Nishiki'		
446	Maanolia 'Honey Tulip'	1055	Salix pentandra		
447	Magnolia 'Hot Flash'	1056	Salix viminalis		
448	Maanolia 'Joli Pompom'	1057	Salvia		
449	Maanolia 'Livingstone'	1058	Sambucus		
450	Magnolia 'March-Till-Frost'	1059	Sambucus nigra 'Black Beauty'		
451	Maanolia 'Peachy'	1060	Sambucus nigra 'Black Lace'		
452	Magnolia 'Red as Red'	1061	Sambucus niara 'Black Tower Eiffel'		
453	Magnolia 'Satisfaction'	1062	Sambucus 'Sampo'		
454	Maanolia 'Shirazz'	1063	Sanauisorba		
455	Maanolia 'Spectrum'	1064	Santolina		
456	Maanolia 'Sunsation'	1065	Scabiosa		
457	Maanolia 'Susan'	1066	Schizostylis		
458	Magnolia 'Watermelon'	1067	Sedum		
459	Magnolia wilsonii 'Eileen Baines'	1068	Senecio		
460	Maanolia × brooklynensis 'Yellow Bird'	1069	Seauoia sempervirens		
461	Mahonia	1070	Seauoiadendron aiaanteum		
462	Malus	1071	Seauoiadendron 'Pendulum'		
463	Malus \times purpurea 'Crimson Cascade'	1072	Sesleria		
464	Malus \times robusta 'Red Sentinel'	1073	Sophora japonica 'Gold Standard'		
465	Malus 'Adam's Pearmain'	1074	Sorbaria		
466	Malus 'Admiration'	1075	Sorbaronia 'Likjormaja Liguorice'		
467	Malus 'Angela'	1076	Sorbus		
468	Malus 'Annie Elizabeth'	1077	Sorbus alnifolia 'Red Bird'		
469	Malus 'Aros'	1078	Sorbus 'Amber Light'		
470	Malus 'Arthur Turner'	1079	Sorbus aria		
471	Malus 'Ashmead's Kernel'	1080	Sorbus aria 'Lutescens'		
472	Malus baccata	1081	Sorbus arranensis		
473	Malus 'Ballerina Flamenco'	1082	Sorbus aucuparia		
474	Malus 'Ballerina Samba'	1083	Sorbus aucuparia 'Aspleniifolia'		
475	Malus 'Bardsey'	1084	Sorbus aucuparia 'Beissneri'		
476	Malus 'Beauty of Bath'	1085	Sorbus aucuparia 'Croft Coral'		
477	Malus 'Black Dabinett'	1086	Sorbus aucuparia 'Fingerprint'		
478	Malus 'Bladon Pippin'	1087	Sorbus 'Autumn Spire'		
479	Malus 'Blenheim Orange'	1088	Sorbus bissetii 'Pearls'		
480	Malus 'Bloody Ploughman'	1089	Sorbus 'Cardinal Royal'		
481	Malus 'Bountiful'	1090	Sorbus carmesina 'Emberglow'		
482	Malus 'Braeburn'	1091	Sorbus cashmiriana		
483	Malus 'Braeburn Mariri Red'	1092	Sorbus 'Chinese Lace'		
484	Malus 'Bramley 20'	1093	Sorbus 'Copper Kettle'		
485	Malus 'Bramley 20'/'Christmas P'/'Scrumptious'	1094	Sorbus discolor		
486	Malus 'Bramley Original'	1095	Sorbus 'Eastern Promise'		

Number	Plant taxa	Number	Plant taxa		
487	Malus 'Bramley's Seedling'	1096	Sorbus 'Ghose'		
488	Malus brevipes 'Wedding Bouquet'	1097	Sorbus 'Glendoick Spire'		
489	Malus 'Browns'	1098	Sorbus 'Glendoick White Baby'		
490	Malus 'Butterball'	1099	Sorbus gonggashanica 'Snow Balls'		
491	Malus 'Candymint'	1100	Sorbus hemsleyi 'John Bond'		
492	Malus 'Cardinal'	1101	Sorbus hupehensis		
493	Malus 'Charles Ross'	1102	Sorbus hupehensis 'Pink Pagoda'		
494	Malus 'Chivers Delight'	1103	Sorbus hybrida 'Gibbsii'		
495	Malus 'Christmas Pippin'	1104	Sorbus japonica		
496	Malus 'Cinderella'	1105	Sorbus 'Joseph Rock'		
497	Malus 'Cobra'	1106	Sorbus 'Leonard Messel'		
498	Malus 'Comtesse de Paris'	1107	Sorbus 'Matthew Ridley'		
499	Malus 'Coralburst'	1108	Sorbus 'Pink Ness'		
500	Malus 'Core Blimey'	1109	Sorbus 'Pink Pearl'		
501	Malus 'Cornish Aromatic'	1110	Sorbus pseudovilmorinii		
502	Malus coronaria 'Elk River'	1111	Sorbus 'Ravensbill'		
503	Malus 'Coul Blush'	1112	Sorbus 'Rose Queen'		
504	Malus 'Cox Lavera'	1113	Sorbus sargentiana		
505	Malus 'Cox Self Fertile'	1114	Sorbus scalaris		
506	Malus 'Cox SF'/'James Grieve'/'Katy'	1115	Sorbus 'Splendens'		
507	Malus 'Cox'/'Fiesta'/'Herefordshire Russet'	1116	Sorbus 'Sunshine'		
508	Malus 'Cox's Orange Pippin'	1117	Sorbus thibetica 'John Mitchell'		
509	Malus 'Dabinett'	1118	Sorbus torminalis		
510	Malus 'Devonshire Quarrenden'	1119	Sorbus ulleungensis 'Olympic Flame'		
511	Malus 'Discovery'	1120	Sorbus vilmorinii		
512	Malus 'Discovery NFT'	1121	Sorbus vilmorinii 'Pink Charm'		
513	Malus 'Donald Wyman'	1122	Sorbus wardii		
514	Malus 'Dr Campbells'	1123	Sorbus 'Wisley Gold'		
515	Malus 'Eden'	1124	Spiraea		
516	Malus 'Egremont Russet'	1125	Stachys		
517	Malus 'Ellison's Orange'	1126	Stachyurus		
518	Malus 'Evereste'	1127	Stipa		
519	Malus 'Fiesta'	1128	Styrax japonicus 'Fragrant Fountain'		
520	Malus florentina	1129	Styrax japonicus 'June Snow'		
521	Malus floribunda	1130	Styrax japonicus 'Pink Snowbell'		
522	Malus 'Fortune'	1131	Symphiocarpus		
523	Malus 'Gala'	1132	Symphoricarpos		
524	Malus 'Gala Brookfield'	1133	Symphytum		
525	Malus 'Galloway Pippin'	1134	Syringa		
526	Malus 'Gilly'	1135	Syringa 'Pink Perfume'		
527	Malus 'Golden Delicious'	1136	Syringa vulgaris 'Beauty of Moscow'		
528	Malus 'Golden Gem'	1137	Syringa vulgaris 'Charles Joly'		
529	Malus 'Golden Glory'	1138	Syringa vulgaris 'Katherine Havemeyer'		
530	Malus 'Golden Hornet'	1139	Syringa vulgaris 'Madame Lemoine'		
531	Malus 'Gorgeous'	1140	Syringa vulgaris 'Mrs Edward Harding'		
532	Malus 'Granny Smith'	1141	Syringa vulgaris 'Primrose'		
533	Malus 'Greensleeves'	1142	Syringa vulgaris 'Sensation'		
534	Malus 'Grenadier'	1143	Syringa vulgaris 'Souvenir de Louis Spaeth'		
535	Malus 'Halloween'	1144	Taxodium distichum		

(Continues)

TABLE C.1 (Continued)	
-------------------------------	--

Number	Plant taxa	Number Plant taxa			
536	Malus 'Harry Baker'	1145	Taxodium distichum 'Shawnee Brave'		
537	Malus 'Harry M Jersey'	1146	Taxodium distichum var. imbricarium 'Nutans'		
538	Malus 'Hastings'	1147	Taxus		
539	Malus 'Herefordshire Russet'	1148	Taxus baccata		
540	Malus 'Hidden Rose'	1149	Taxus baccata 'Fastigiata Robusta'		
541	Malus 'Honeycrisp'	1150	Taxus baccata 'Standishii'		
542	Malus 'Howgate Wonder'	1151	Tellima		
543	Malus hupehensis	1152	Tetradium daniellii		
544	Malus 'Indian Magic'	1153	Thalictrum		
545	Malus ioensis 'Fimbriata'	1154	Thuja		
546	Malus ioensis 'Purpurea EVELYN'	1155	Thuja plicata		
547	Malus 'Irish Peach'	1156	Thymus		
548	Malus 'Isaac Newton'	1157	Tiarella		
549	Malus 'James Grieve'	1158	Tilia		
550	Malus 'Jelly King'	1159	<i>Tilia</i> × <i>europaea</i> 'Golden Sunset'		
551	Malus 'John Downie'	1160	<i>Tilia</i> × <i>europaea</i> 'Wratislaviensis'		
552	Malus 'Julia's Late Golden'	1161	Tilia cordata		
553	Malus 'Jumbo'	1162	<i>Tilia cordata</i> 'Greenspire'		
554	Malus 'Jupiter'	1163	Tilia cordata 'Winter Orange'		
555	Malus 'Katy'	1164	Tilia euchlora		
556	Malus 'Keswick Codlin'	1165	Tilia henryana 'Arnold Select'		
557	Malus 'Kidd's Orange Red'	1166	Tilia platanoides		
558	Malus 'King of the Pippins'	1167	Tilia platanoides 'Tiltstone Filigree'		
559	Malus 'King's Acre Pippin'	1168	Tilia platyphyllos		
560	Malus 'Kingston Black'	1169	Trachelospermum		
561	Malus 'Lady Henniker'	1170	Tradescantia		
562	Malus 'Lane's Prince Albert'	1171	Tricyrtis		
563	Malus 'Laura'	1172	Trollius		
564	Malus 'Laxton's Superb'	1173	Tsuga heterophylla		
565	Malus 'Limelight'	1174	Ulex		
566	Malus 'Little Pax'	1175	Ulmus		
567	Malus 'Lord Derby'	1176	<i>Ulmus</i> × <i>hollandica</i> 'Wredei'		
568	Malus 'Lord Lambourne'	1177	Ulmus glabra		
569	<i>Malus</i> 'Louisa'	1178	<i>Ulmus</i> × Wingham		
570	Malus 'Major'	1179	Uncinia		
571	Malus 'Marble NUVAR'	1180	Vaccinium 'Bluecrop'		
572	Malus 'Melrose Belmonte'	1181	<i>Vaccinium</i> 'Chandler'		
573	Malus 'Meridian'	1182	Vaccinium 'Darrow'		
574	Malus 'Michelin'	1183	<i>Vaccinium</i> 'Duke'		
575	Malus 'Newton Wonder'	1184	Vaccinium 'Liberty'		
576	Malus 'Orleans Reinette'	1185	Vaccinium 'Northland'		
577	Malus 'Paradice Gold'	1186	Vaccinium 'Patriot'		
578	Malus 'Peasgood's Nonsuch'	1187	Vaccinium 'Pink Lemonade'		
579	Malus 'Pink Glow'	1188	Vaccinium 'Sunshine Blue'		
580	Malus 'Pink Perfection'	1189	Verbena		
581	Malus 'Pinot Prince SUPERNOVA'	1190	Veronica		
582	Malus 'Pitmaston Pine Apple'	1191	Viburnum		
583	Malus 'Pixie'	1192	Viburnum lantana		

TABLE C.1 (Continued)

Number	Plant taxa	Number	Plant taxa
584	Malus 'Porters Perfection'	1193	Viburnum opulus
585	Malus 'Prairie Fire'	1194	Viburnum opulus 'Roseum'
586	Malus 'Prince William'	1195	Viburnum plicatum 'Kilimanjaro'
587	Malus 'Professor Sprenger'	1196	Vinca
588	Malus 'Queen Cox S.F 18'	1197	Vitis Bacchus'
589	Malus 'Queen of the Realm'	1198	<i>Vitis</i> 'Dornfelder'
590	Malus 'Red Devil'	1199	Vitis 'Lakemont'
591	Malus 'Red Falstaff'	1200	Vitis 'Muscat Bleu'
592	Malus 'Red Foxwhelp'	1201	<i>Vitis</i> 'Phoenix'
593	Malus 'Red Jonaprince'	1202	Vitis 'Polo Muscat'
594	Malus 'Red Obelisk'	1203	Vitis 'Regent'
595	Malus 'Red Topaz'	1204	Vitis 'Strawberry'
596	Malus 'Red Windsor'	1205	Vitis 'Suffolk Red'
597	Malus 'Reverend W. Wilks'	1206	Weigela
598	Malus 'Ribston Pippin'	1207	Wisteria brachybotrys 'Golden Dragon'
599	Malus 'Rosehip'	1208	Wisteria brachybotrys 'Kapiteyn Fugi'
600	Malus 'Rosemary Russet'	1209	Wisteria brachybotrys 'Okayama'
601	Malus 'Rosette'	1210	Wisteria brachybotrys 'Shiro Beni'
602	Malus 'Royal Beauty'	1211	Wisteria 'Burford'
603	Malus 'Royalty'	1212	Wisteria floribunda 'Black Dragon'
604	Malus 'Rudolph'	1213	Wisteria floribunda 'Hon-beni'
605	Malus 'Santana'	1214	Wisteria sinensis 'Prolific'
606	Malus 'Saturn'	1215	Xanthocyparis nootkatensis 'Pendula'
607	Malus 'Scarlet Brandywine'	1216	Yucca
608	Malus 'Scarlett'	1217	Zelkova serrata 'Kiwi Sunset'
609	Malus 'Scotch Bridget'		

APPENDIX D

Water used for irrigation

All mains water used meets the UK standard Water Supply (Water quality) regulation 2016 and the WHO/EU potable water standards, (Drinking water Directive (98/83/EC and the revised Drinking Water Directive 2020/2184) which includes a total freedom from both human and plant pathogens (Article 2-(7)). All mains water conducting pipework fully complies with the UK Water Supply (Water Fittings) regulations of 1999 and the amendments of 2019. Irrigation water used is not stored in any open tanks where air borne contamination could take place and is entirely isolated from any outside exposure (Dossier Sections 1.1, 1.2 and 1.3).

Bore hole water supply: in some cases, where the underlying geology permits, nurseries can draw water directly from bore holes drilled into underground aquafers. The water that fills these aquafers is naturally filtered through the layers of rock (e.g. limestone) over long periods of time, many millennia in some cases. The water from such supplies is generally of such high quality that it is fit for human consumption with little to no further processing and is often bottled and sold as mineral water (Dossier Sections 1.1, 1.2 and 1.3).

Rainwater or freshwater watercourse supply: some nurseries contributing to this application for both environmental and efficiency reasons use a combination of rain capture systems or abstract directly from available watercourses. All water is passed through a sand filtration system to remove contaminants and is contained in storage tanks prior to use. One nursery that operates this approach is currently in the process of installing additional nanobubble technology to treat the water (Dossier Sections 1.1, 1.2 and 1.3).

APPENDIX E

List of pests that can potentially cause an effect not further assessed

TABLE E.1 List of potential pests not further assessed.

N	Pest name	EPPO Code	Group	Pest present in the UK	Present in the EU	<i>Populus</i> confirmed as a host (reference)	Pest can be associated with the commodity	Impact	Justification for inclusion in this list
1	Elsinoe populi		Fungi	Yes	Limited	Populus nigra (Farr & Rossman, 2024)	Yes	No data	Uncertainty on the impact
2	Meloidogyne mali	MELGMA	Nematodes	Yes	Limited	Wide host range (Dossier)	Uncertain	Uncertain	Uncertainty on the association with <i>Populus</i> and its impact on Populus
3	Pemphigus populitransversus	PEMPPO	Insects	Yes	Limited	<i>Populus nigra</i> (Aphids on World's Plants, 2024)	Yes	Uncertain	There is uncertainty on the impact on <i>Populus</i>

APPENDIX F

Excel file with the pest list of Populus alba, Populus nigra and Populus tremula

Appendix F is available under the Supporting Information section.



